


Depicting the Implication of miR-378a in Cancers

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Abstract

MicroRNA-378a (miR-378a), including miR-378a-3p and miR-378a-5p, are encoded in *PPARGC1B* gene. miR-378a is essential for tumorigenesis and is an independent prognostic biomarker for various malignant tumors. Aberrant expression of miR-378a affects several physiological and pathological processes, including proliferation, apoptosis, tumorigenesis, cancer invasion, metastasis, and therapeutic resistance. Interestingly, miR-378a has a dual functional role in either promoting or inhibiting tumorigenesis, independent of the cancer type. In this review, we comprehensively summarized the role and regulatory mechanisms of miR-378a in cancer development, hoping to provide a direction for its potential use in cancer therapy.

Keywords

miR-378a, tumorigenesis, migration, invasion, proliferation, targets, clinical applications

Abbreviations

ALCAM, activated leukocyte cell adhesion molecule; AML, acute myelocytic leukemia; ATG12, autophagy-related protein 12; BC, breast cancer; BL, Burkitt lymphoma; BM, brain metastasis; BMP2, bone morphogenetic proteins 2; BMNCs, bone marrow mononuclear cells; BRAF, B-Raf proto-oncogene serine/threonine kinase; BYSL, bystin like; CC, cervical cancer; CDC40, cell division cycle 40; CDDP, cisplatin; CDK, cyclin dependent kinase; CML, chronic myeloid leukemia; CRC, colorectal cancer; DCTPPI, dCTP pyrophosphatase 1; DFS, disease-free survival; EMT, epithelial-mesenchymal transition; EHD1, EH domain containing 1; EPA, eicosapentaenoic acid ethyl ester; ESCC, esophageal carcinoma; FOXG1, Fork head-box gene 1; FOXN3, Forkhead box N3; FUS-1, FUS RNA binding protein; FOXN3, forkhead box N3; GC, gastric cancer; GLUT1, glucose transporter 1; GOLT1A, golgi Transport 1A; HCC, hepatocellular carcinoma; HMOX1, heme oxygenase-1; HOXD10, homeobox D10; IRAK4, interleukin-1 receptor associated kinase-4; IRG1, immune responsive gene 1; KLF9, Fruppel-like factor 9; KLK, Kallikrein-related peptidase; KISS1, tumor metastasis suppressor kiss-1; LXR, liver X receptor; miR-378a, MicroRNA-378a; MAPK1, mitogen-activated protein kinase 1; MB, medulloblastoma; MED19, mediator complex subunit 19; MNT, MAX network transcriptional repressor; NPC, nasopharyngeal carcinoma; NSCL, non-small cell lung carcinoma; OC, ovarian cancer; OCT4, organic cation/carnitine transporter 4; OS, overall survival; OSCC, oral squamous carcinoma; PA, pituitary adenoma; PCa, prostate cancer; PDGFR β , platelet derived growth factor receptor beta; PFS, progression-free

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survival; POLR2A, RNA polymerase II subunit A; Ppargc1 β , peroxisome proliferator-activated receptor γ coactivator 1 beta; PSA, prostate-specific antigen; RB, retinoblastoma; RBX1, ring-box 1; RMS, rhabdomyosarcoma; RNF31, ring finger protein 31; RUNX1, runt related transcription factor 1; SCLU, secreted form clusterin; SDAD1, SDAI domain containing 1; SERPINE1, serpin family E member 1; SKP2, S-phase kinase associated protein 2; SMAD4, SMAD family member 4; SOX2, SRY-box transcription factor 2; SOX7, SRY-box transcription factor 7; STAMPB, STAM binding protein; ST7L, suppression of tumorigenicity 7 like; STAT3, signal transducer and activator of transcription 3; SUFU, SUFU negative regulator of hedgehog signaling; TOB2, transducer of ERBB2; TSPAN17, tetraspanin 17; TNBC, triple-negative breast cancer; TUSC2, tumor suppressor 2, mitochondrial calcium regulator; UHRF1, ubiquitin like with PHD and ring finger domains 1; VEGF, vascular endothelial growth factor; ZBTB20, zinc finger and BTB domain containing 20

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Introduction

Micro RNAs (miRNAs) are small non-coding RNAs (19-24 nucleotides) produced from an endogenous transcript having a local hairpin structure with the help of RNase III-type enzyme.¹ miRNAs can repress the translation of target proteins by cleaving or clinging to the 3' untranslated region (UTR) of the corresponding messenger RNAs (mRNAs).² The first miRNA was discovered over 20 years ago in mammals.³ Based on their genomic origin, miRNAs are divided into intergenic, intronic, and exonic.⁴ They are essential regulators of various critical biological processes, such as cell differentiation,^{5,6} apoptosis,^{7,8} proliferation,⁹ cell division,¹⁰ protein secretion,¹¹ and viral infection.¹² Recently, several studies have shown that the aberrant expression of miRNA is tightly associated with the progression of human diseases, especially cancers,¹³ as they can suppress or promote tumor growth by regulating the target gene mRNA.¹⁴⁻¹⁶

The miRNA miR-378a is one such essential tumor-regulating miRNA. It is located on chromosome 5q32 and embedded within the first intron of the *PPARGC1 β* gene, which encodes the PGC1 β protein. miR-378a is divided into miR-378a-3p (usually identified as miR-378) and miR-378a-5p (usually identified as miR-378*), which are the guide and passenger strands of miR-378a, respectively.¹⁷ Both miR-378a-3p and miR-378a-5p are synchronously transcribed with PGC1 β ,¹⁸⁻²⁰ although there is an exception. Recently, it has been demonstrated that the over-expression (Table 1) or under-expression (Table 2) of miR-378a regulates the expression of its target genes and affects the proliferation,²¹ invasion, and metastasis²² of cancers. For example, low miR-378a expression is usually correlated to a significantly poor overall survival (OS)²³ in colorectal cancer patients and associated with significantly decreased OS and disease-free survival (DFS) in gastric cancer patients.²⁴ However, its expression and role in different cancers, such as lung²⁵ and cervical cancers,²⁶ is still controversial. miR-378a was identified as an onco-RNA in various tumors, including lung cancer,^{22,25,27-29} ovarian cancer,³⁰⁻³² cervical cancer,³³ nasopharyngeal carcinoma,³⁴ melanoma,³⁵ osteosarcoma,³⁶ cholangiocarcinoma,³⁷ acute myeloid leukemia,³⁸ chronic myeloid leukemia,²¹ and Burkitt lymphoma.³⁹ In contrast, it acted as an tumor suppressor

in gastric cancer,⁴⁰ colorectal cancer,⁴¹ liver cancer,⁴² glioblastoma,⁴³ prostate cancer,⁴⁴ breast cancer (BC),⁴⁵ medulloblastoma (MB),⁴⁶ pituitary adenoma (PA),⁴⁷ oral squamous cell carcinomas,⁴⁸ bladder cancer,⁴⁹ esophageal carcinoma,⁵⁰ rhabdomyosarcoma (RMS),⁵¹ and retinoblastoma (RB).⁵² Compared with normal tissues and no lymph node metastasis cancer tissues, miR-378a was upregulated in cervical cancer tissues, especially in CIN III and lymph node metastasis cervical cancer.³³ Additionally, a study of 120 cholangiocarcinoma tissues and adjacent noncancerous tissues found that miR-378a expression increased with the development of the TNM (T-tumor, N-nodes, M-metastasis) stages.³⁷ Positive miR-378a expression is also tightly linked to a low OS in cholangiocarcinoma patients.³⁷ However, in colorectal cancer patients, miR-378a is downregulated and low miR-378a correlated with poor OS,⁴ suggesting an inhibiting effect of miR-378a.²⁴ Thus, miR-378a can drive cancer cell proliferation,^{29,53} invasion, and migration^{27,32} in some tumors, while inhibiting these processes in other tumors.^{23,24,32,54,55} Further, a study on the radiation response in a glioblastoma xenograft model found that the overexpression of miR-378a was associated with enhanced local tumor response to radiotherapy by increasing the vascular density and perfusion, thereby prolonging the survival of tumor-bearing hosts and acting as a novel therapeutic intervention.⁴³ To summarize, the abnormal expression of miR-378a is crucial for the occurrence and development of malignant tumors. Therefore, it is important to understand the functions of miR-378a in tumor biology to enhance the efficiency of early screening and diagnosis of cancer, strengthen anticancer therapy, and develop novel targeted methods for tumor therapy. Hence, this review focuses on the roles and targets of miR-378a in different cancers to elucidate the specific molecular mechanisms of miR-378a during tumor progression.

Dual Function of miR-378a in Cancer Development

Growing research studies have elucidated the critical role of miR-378a in cancer progression, and most studies focused on the specific targets of miR-378a in tumor cell proliferation,

Table 1. miR-378a Plays Tumorigenic Roles as an Oncogene.

Tumor	Sample Type	Expression of MiR-378a in Tumor sample	Role of MiR-378a Upregulation in Vitro	Role of miR-378a Upregulation in Vivo	Reference
Lung cancer	Cancer tissue and cell	Lung cancer tissue with brain metastasis (BM)-↓, BM + ↑; A549↑	Cell survival↑, migration and invasion↑, angiogenesis↑	Growth↑, angiogenesis↑	22
	Cell	A549↑, SK-LU-1↓	Invasion↑	Metastasis↑, angiogenesis↑	27
	Cancer tissue and cell	Lung cancer tissue↑; A549↑, H1299↑, H460↑, H520↑, H1975↑, PC9↑	Cell growth↑, apoptosis↓	—	28
	Cell	SK-MES-1↓, A549↓, NCI-H460↑, NCI-H292↑	Proliferation↑, migration↑, angiogenesis↑	Growth ↑, metastasis↑	29
	Serum	Serum exosomal miR-378a↑	—	—	57
	Cell	—	Proliferation↑, migration↑	—	59
OC	Cancer tissue	Lung cancer tissue↑	Proliferation↑	—	60
	Cell	A549↑, A549/CDDP↓, Anip973↑, Anip973/CDDP↓	CDDP sensitivity↑, apoptosis↑	Growth↓	25
	Cell	SKOV3↑, CAOV3↑	Proliferation↑, migration↑, invasion↑, angiogenesis↑	—	32
CC	Cell	SKOV3↑, A2780↑, H281↓, H252↓, H254↓	—	—	62
	Cell	A2780↑, CAOV3↑, OVCAR3↓	—	—	63
	Cancer tissue	Normal tissue↓; cancer tissue↑	Migration↑, invasion↑	Metastasis↑	33
	Cancer tissue and cell	CC tissue↑; HeLa↑, C33A↑, SiHa↑, CaSKi↑, End1/E6E7↓	Growth↑, apoptosis↓, cell cycle progression↑, Wnt/β-catenin pathway↑	—	65
Melanoma	Serum and cancer tissue	Serum of CC patients↓; CC tissue↓	Proliferation↓	Growth↓	26
	Cell	C33A↓, ME-1800↓	Proliferation↓, Migration↓, invasion↓	—	64
	Cancer tissue	Melanoma tissue↑, lymph node metastasis↑	Migration↑, invasion↑, EMT↑	Growth↑	35
Melanoma	Cancer tissue	Melanoma tissue↑	Proliferation↑, migration↑, angiogenesis↑	Angiogenesis↑	67
NPC	Cell	5-8F↑, 6-10B↑, CNE1↑ CNE2↑	Proliferation↑, migration and invasion↑	Growth↑	34
Osteosarcoma	Cancer tissue and cell	Osteosarcoma tissue↑; Saos2↑, U2OS↑, MG-63↑, HOS↑	Proliferation↑	—	71
Cholangiocarcinoma	Cancer tissue and cell	Cholangiocarcinoma tissue↑; QBC939↑, RBE↑	Proliferation↑, migration and invasion↑	—	37
Renal carcinoma	Serum	RCC patients↑, patients after radical nephrectomy↓	—	—	73
	Serum	RCC patients↑	—	—	74,75
BL	Cell	ST486↑, Ramos↑, CA46↑DG75↑	Growth↑	—	39
AML	Cancer sample and cell	AML samples↓; bone marrow mononuclear cells (BMNCs)↑	—	—	38
CML	BMMNCs	BMMNCs↑	Proliferation↑, apoptosis ↓, drug resistance to 5-FU↑, ability of stem cell sphere formation↑	—	21

↑indicates upregulation (in case of expression level) or promoting (in case of functional consequences).

↓indicates downregulation (in case of expression level) or suppressing (in case of functional consequences).

migration, invasion, and regulatory pathways. Here, we summarize the roles and targets of miR-378a in tumorigenesis. Interestingly, both oncogenic (Table 1) and tumor-suppressive

(Table 2) roles have been reported for miR-378a, implicating its dual function in cancer progression. By regulating downstream target genes, miR-378a promotes (Table 3) or inhibits

Table 2. miR-378a Plays Inhibiting Roles by Acting as a Tumor Suppressor.

Tumor	Sample Type	Expression of MiR-378a in Tumor Sample	Role of miR-378a Upregulation in Vitro	Role of miR-378a Upregulation in Vivo	Reference
GC	Cancer tissue and cell	GC tissue↓; BGC-823↓, MKN-28↓, MGC-803↓, SGC-7901↓, AGS↓	Invasion↓, migration↓, EMT↓	–	24
	Cancer tissue and cell	GC tissue↓; SGC-7901↓, AGS↓, MGC-803↓	Proliferation↓, G2/M phases↑	–	40
	Cancer tissue and cell	GC tissue↓; KATO III↑, HGC-27↓, SNU-1↓	Proliferation↓, delay the cell cycle	–	54
	Cancer tissue	GC tissue↓	–	–	82
	Serum and cancer tissue	Serum level of GC patients↑, GC tissue↓	–	–	83
CRC	Cancer tissue and cell	CRC tissue↓; SW620↓, SW480↓, HT29↓, HCT116↓	Progression↓, invasion↓	Growth↓	23
	Cancer tissue and cell	CRC tissue↓; HCT116↓, HT29↓, DLD-1↓, LOVO↓, SW620↓, SW480↓	Proliferation↓, cell of G0/G1 phase↑ and S-phase↓, cisplatin-induced apoptosis↑	Growth↓	41
	Cancer tissue and cell	CRC tissue↓; LoVo↓, CaCo2↓, SW1116↓, SW480↓, HCT-116↓	Proliferation↓, apoptosis↑	–	55
	Cancer tissue and cell	CRC and cell↓; HT29↓, HCT116↓, SW480↓, SW1116↓, SW620↓, LOVO↓	Proliferation↓, apoptosis↑	–	86
	Plasma	Plasma level of CRC patient↓	–	–	88
	Cancer tissue	CRC tissue↓	Invasion↓	–	91
	Cancer tissue and cell	CRC tissue↓; HT-29↓, SW620↓, HT55↓	–	Growth↓	92
	Cancer tissue and cell	CRC tissue↓; LoVo↓, CaCo2↓, SW1116↓, SW480↓, HCT-116↓	Proliferation↓, apoptosis↑, migration and invasion↓	–	93
	–	–	Proliferation↓, migration and invasion↓	–	94
	Liver cancer	Cell	–	Proliferation↓	Growth↓
	Cancer tissue and cell	Liver cancer tissue↓; HepG2↓, HuH7↓, SMMC-7721↓, Li-7↓, PLC/PRF5↓, SK-Hep-1↓, MHCC97L↓, and MHCC97↓	Growth↓, migration and invasion↓, PLAGL2/β-catenin signaling↓	Growth↓, distant metastasis↓	96
	Cancer tissue and cell	HCC tissue↓; SMMC-7721↓, MHCC-97H↓, MHCC-97L↓, HCCLM3↓	Angiogenesis↓, NF-κB signaling↓	Angiogenesis↓	97
	Cancer tissue	Liver cancer tissue↓	Proliferation↓, invasion ↓, sorafenib-based chemotherapies↑	–	98
	Cancer tissue and cell	–	Migration and invasion↓	–	101
	Cancer cell	Sorafenib resistance cell↓	Response to sorafenib↑	–	102
Glioma	Cancer tissue	HCC tissue↓	HCC cell viability↓, apoptosis↑, migration and invasion↓	–	103
	Cancer tissue and cell	Glioma tissue↓; SHG44↓, A172↓, LN229↓, LN18↓, T98↓	Migration and invasion↓, EMT↓	Growth↓	108
	Cancer tissue and cell	Glioma tissue↓; U87↓, U251↓, MT330↓, SJ-G2↓	Migration and invasion↓	–	109
	Cancer tissue and cell	Glioma tissue↓; U87MG↓, MT-330↓	Proliferation↓, apoptosis↑, migration and invasion↓	–	110
PCa	Cancer tissue and cell	PCa tissue↓; LNCAP↓, PC-3M1E8↓, PC-3M↓, PC-3M2B4↓, du145↓	Proliferation↓, apoptosis↑, migration and invasion↓, cell in G1/S phase↑	Growth↓	44
	Cancer tissue	PCa tissue↓	–	–	113

(continued)

Table 2. (continued).

Tumor	Sample Type	Expression of MiR-378a in Tumor Sample	Role of miR-378a Upregulation in Vitro	Role of miR-378a Upregulation in Vivo	Reference
	Cancer tissue	PCa tissue↓	–	–	115
	Cell	22Rv1↓, LNCaP↓, PC3↓, DU145↓	Proliferation↓, glucose metabolism↓	–	114
BC	Cancer tissue from mouse model and BC cell of human	BC tissue↓; MDA-231↓	Migration and invasion↓	–	45
	Cell	Tamoxifen-resistant MCF-7 cells↓, long-term estrogen-deprived MCF-7 cells↓	Growth↓ and apoptosis↑	–	116
	–	–	Proliferation↓	–	118
MB	Cancer tissue	MB tissue↓	Proliferation↓, apoptosis↑	–	46
PA	Cancer tissue	PA tissue↓	Proliferation↓, migration↓	–	47
OSCC	Cancer tissue	OSCC tissue↓	–	–	121
	–	–	Migration and invasion↓, EMT↓	Metastasis↓	48
	Cancer tissue and cell	OSCC tissue↓; Tca8113↓, CAL-27↓	Proliferation↓, apoptosis↑	–	121
Bladder cancer	Cancer tissue	Bladder cancer tissue↓	–	–	49
	Cancer tissue	Bladder cancer tissue↓	Cell viability↓, migration and invasion↓	–	123
ESCC	Cancer tissue and cell	ESCC tissue↓; TE-1↓, KYSE-150↓, Eca-109↓, TE-8	Proliferation↓, migration and invasion↓	–	50
	Cancer tissue and cell	ESCC tissue↓; EC109↓, KYSE150↓	Proliferation↓, apoptosis↑, cells in G1 phase↑, migration and invasion↓	–	124
	Cancer tissue and cell	ESCC tissue↓; Eca109↓, KYSE150↓, KYSE30↓, KYSE450↓, KYSE70↓	Proliferation↓, invasion↓	–	126
RMS	Cancer tissue and cell	RMS tissue↓; ARMS RH30↓, ERMS RD↓	Apoptosis↑, migration↓	–	51
RB	Cancer tissue	RB tissue↓	Cell viability↓, apoptosis↑	Growth↓	52

↑indicates upregulation (in case of expression level) or promoting (in case of functional consequences).

↓indicates downregulation (in case of expression level) or suppressing (in case of functional consequences).

tumors (Table 4), while it is regulated by its upstream regulators (Figure 1).

Upregulation of miR-378a Promotes Malignant Transformation and Progression

Lung Cancer

Lung cancer is a common malignant tumor with a high mortality rate.⁵⁶ Despite advances in cancer therapy, the OS rate of patients with advanced lung cancer is low. The expression of miR-378a both in tumor tissues and cell lines was significantly higher than in adjacent normal tissues or cells, and upregulation of miR-378a promoted the tumor progression.^{22,25,27-29} Further, miR-378a was significantly overexpressed in serum exosomes of non-small cell lung cancer (NSCLC) patients. The aberrant expression of miR-378a was closely associated with an advanced TNM stage, positive metastasis, negative therapeutic response, and poor OS.^{25,57} miR-378a can promote NSCLC cell

migration, invasion, and tumor angiogenesis.^{22,27} Moreover, the circular RNA has-circ-0007059 was identified as an upstream regulatory factor of miR-378a, which can inhibit the proliferation and epithelial-mesenchymal transition (EMT) of A549 and H1975 cells via inhibiting miR-378a.⁵⁸ The lncRNA ACTA2-AS1 suppressed malignancy by decoying miR-378a-3p, thus enhancing SOX7 expression.⁵⁹ As for the downstream regulation, miR-378a works on different targets and pathways to produce a plethora of effects on lung cancer progression. Studies on stimulating miR-378a for promoting lung cancer progression and constructing miR-378a overexpression cell lines have shown that miR-378a can strengthen metastasis by regulating EMT.²⁷ Further, miR-378a targets RBX1, and miR-378 upregulation boosts NSCLC cell invasion by downregulating RBX1, but it did not have any major effect on the angiogenesis signaling pathway. Likewise, HMOX1 was another target of miR-378a, and the interplay between HMOX1 and miR-378a²⁹ modulated the miRNA transcriptome and the 3' UTR of HMOX1 in tumors. Further experiments in

Table 3. miR-378a Functions as an Onco-miRNA in Cancers by Regulating the Downstream Targets.

Cancer Type	Expression of Downstream Target When miR-378a is Upregulated	Effects of Changes in Target Genes	Reference
Lung cancer	SOX7↓	Promote proliferation, invasion, migration and EMT but inhibited cell apoptosis	59
	RBX1↓	Promote invasion	27
	HMOX1↓	Promote proliferation, migration and angiopoiesis	29
	FOXG1↓	Promote proliferation and cell cycle distribution at G0/G1 phase but attenuated cell apoptosis	28
	SCLU↓	Promote chemoresistance of cisplatin and apoptosis	25
	CDK4↓	Inhibit proliferation	60
	CDK6↓	Inhibit proliferation	60
	TUSC2↓	Promote growth	61
OC	ALCAM↑	A marker for better response to antiangiogenic therapy	62
	EHD1↓	A marker for worse response to antiangiogenic therapy	62
CC	SMAD4↓	Promote proliferation, migration, invasion, and angiogenesis	32
	CPEB3↓	Promote proliferation, migration, invasion and EMT but decrease apoptosis	64
	ATG12↓	Promote migration and invasion	33
	ST7L↓	Promote proliferation by regulating the cell cycle process but inhibit apoptosis and Wnt/β-catenin pathway	65
Melanoma	SUFU↓	Unknown	67
	FUS-1↓	Unknown	67
	KLF9↓	Unknown	67
	STAMBP↓	Unknown	67
	HOXD10↓	Unknown	67
NPC	TOB2↓	Promote proliferation, migration, invasion in vitro and tumor growth vivo	71
Osteosarcoma	KLF9↓	Promote proliferation	36
	BYSL↓	Inhibit EMT and invasion	72
Renal cell carcinoma	POLR2A↓	Promote apoptosis	77
	RUNX2↓	Promote apoptosis	77
BL	IRAK4↓	Promote cell growth	39
	MNT↓	Promote cell growth	39
AML	ZBTB20↑	Promote growth and invasion	79
CML	FUS1↓	Promote proliferation	80
	FUS1↓	Promoted proliferation	21

↑indicates upregulation; ↓indicates downregulation.

NCI-H292 cells with stable overexpression of miR-378a indicated that miR-378a enhanced the proliferation, migration, and angiogenic capabilities of the cells both *in vitro* and *in vivo* by downregulating HMOX1 mRNA and protein expression. However, when HMOX1 was diminished, miR-378a could not modulate the cell's resistance to chemotherapeutic drugs and oxidative stress. Another study noted a reverse correlation between miR-378a and FOXG1.²⁸ Silencing FOXG1 led to tumor suppression in NSCLC cells by downregulating miR-378a, thereby inhibiting NSCLC cell proliferation, promoting apoptosis, and increasing the length of the G0/G1 phase in the cell cycle. Hence, all the above studies show a positive relation between miR-378a and tumorigenesis.

However, some studies have shown the opposite result. Cisplatin (CDDP)-sensitive patients have high expression of miR-378a and low expression of small cell lung cancer (SCLU).²⁵ Forced upregulation of miR-378a inhibits neoplasm growth and SCLU expression and makes cells more sensitive to CDDP. Overexpression of miR-378a-3p can inhibit cell

proliferation and decrease the expression of proliferation-related proteins CDK4 and CDK6. Since lncRNA OIP5-AS1 functioned as a competing endogenous RNA of miR-378a-3p, overexpressed wild-type OIP5-AS1 increased CDK4 and CDK6 expression, thereby promoting tumor growth.⁶⁰ In addition, SCP inhibits tumorigenesis by upregulating TUSC2 by targeting miR-378a-5p.⁶¹ Thus, the aberrant miR-378a expression in lung cancer significantly affects tumors. However, the controversy over whether it is the low or high expression of miR-378a that increases lung cancer progression is yet to be resolved.

Gynecological Cancer

Ovarian cancer (OC) and cervical cancer (CC) are the most common gynecological tumors. In OC and CC, miR-378a is overexpressed both in the tumor tissues and cancer cell lines.^{31,33} Among recurrent OC patients treated with bevacizumab, higher miR-378a correlated to longer PFS. miR-378a and

Table 4. miR-378a Functions as a Suppressor in Various Cancers by Regulating the Downstream Targets.

Cancer Type	Expression of Downstream Target When miR-378a is Upregulated	Effects of Changes in Target Genes	Reference
GC	BMP2↓	Inhibit migration, invasion and EMT	24
	MAPK1↓	Inhibit proliferation, migration, invasion and cell cycle progression	84
	VEGF↓	Unknown	85
CRC	MED19↓	Inhibit proliferation, migration, invasion and EMT, decrease cells in S phase and increase cells in G0 phase	91
	IGF1R↓	Unknown	86
	CDC40↓	Inhibit cell growth	41
	Vimentin↓	Unknown	23
	BRAF↓	Inhibit proliferation and induce apoptosis	55
	SDAD1↓	Inhibit proliferation, migration and invasion	93
	KISS1↑	Inhibit proliferation, migration and invasion	94
	VEGFR↓	Unknown	98
	PDGFRβ↓	Unknown	98
	C-Raf↓	Unknown	98
Liver cancer	FUS↓	Unknown	100
	VEGF↓	Inhibit migration, invasion and proliferation	101
	IGF1R↓	Promote sorafenib sensitivity	102
	IRG1↓	Inhibit metastasis and EMT	108
	TSPAN17↓	Inhibit proliferation migration, invasion and decreased apoptosis	110
	SUFU↓	Increase cell survival, tumor growth and angiogenesis	112
Glioma	FUS-1↓	Increase cell survival, tumor growth and angiogenesis	112
	GLUT1↓	Inhibit glycolytic metabolism and decrease cell survival	114
	KLK2↑	Unknown	113
PCa	KLK4↑	Unknown	113
	MAPK↓	Inhibit proliferation, invasion and migration, induce apoptosis and cause cell cycle arrest	44
	YY1↓	Unknown	117
	DCTPP1↓	Inhibit proliferation, induce apoptosis and activate DNA Repair mediated signaling pathway in vitro, repress tumorigenesis in vivo	118
BC	RUNX1↓	Inhibit migration	45
	SUFU↓	Decrease apoptosis, increase chemotherapy-resistant	119
	KLK4↓	Unknown	120
MB	UHRF1↓	Inhibit proliferation and apoptosis	46
PA	RNF31↓	Inhibit proliferation and migration	47
OSCC	KLK4↓	Inhibit migration and invasion	48
	GLUT1↓	Inhibit glycolysis and cell proliferation	122
	NCAPG↓	Inhibit proliferation and decrease apoptosis	123
ESCC	Rab10↓	Inhibit proliferation, invasion and migration	125
	GLUT1↓	Inhibit proliferation, invasion and induce apoptosis	126
	SPHK1↓	Inhibit proliferation and invasion	127
RMS	IGF1R↓	Unknown	51
RB	FOXG1↓	Inhibit proliferation and promote apoptosis	52

↑indicates upregulation; ↓indicates downregulation

its downstream targets ALCAM and EHD1 can be used as markers for anti-angiogenic therapy.⁶² In OC tissues and cells, it has been shown that circATRNL1 is under-expressed while miR-378a is overexpressed.³² Further study revealed that circATRNL1 could adhere to miR-378a while miR-378a directly acts on SMAD4. High circATRNL1 expression also reduces miR-378a, which, in turn, inhibits SMAD4 expression, resulting in the suppression of angiogenesis, cell proliferation, invasion, and migration, both *in vitro* and *in vivo*. Hence, in OC, circATRNL1 acts as a miR-378a sponge, thereby accelerating SMAD4 signaling and attenuating angiogenesis and metastasis.³²

miR-378a is also a target of circ-LOPD2. circ-LOPD2 expression is high in both OC tissue and cells and is responsible for promoting OC cell proliferation.⁶³ Several studies have revealed that circ-RNAs worked as molecule sponges of miR-378a to regulate cancer development.

As for CC, miR-378a acts as an oncogene since high miR-378a expression significantly enhances cancer migration and invasion *in vitro*, and metastasis *in vivo*, while downregulating miR-378a produces the opposite effect *in vitro*.³³ The lncRNA LINC00641 inhibits CC progression by diminishing miR-378a-3p. Further, knockout of CPEB3, a downstream target of miR-378a-3p, can reverse the effects induced by

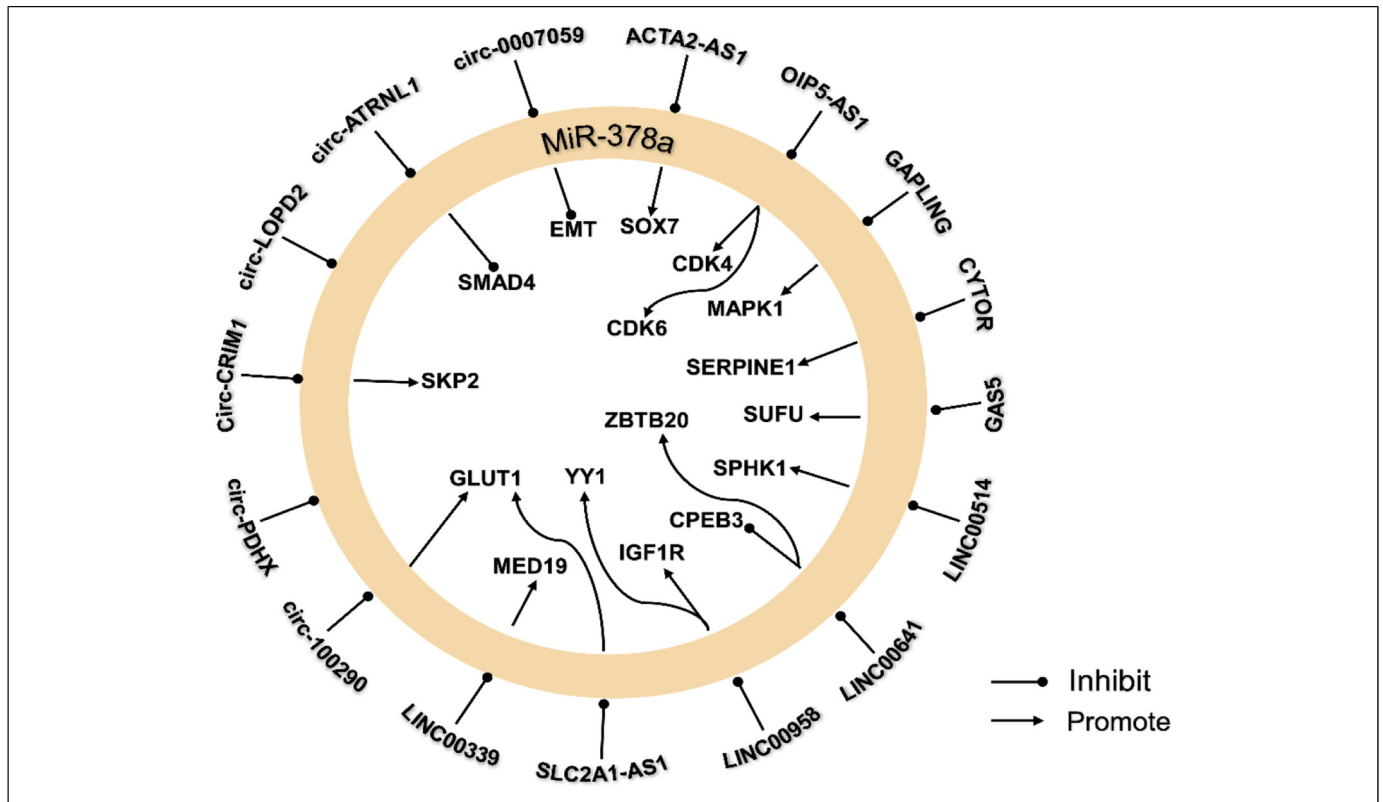


Figure 1. The regulatory factors upstream of miR-378a and the downstream target genes regulated by miR-378a in human cancer. miR-378a can be targeted and regulated by lncRNAs and circRNAs, thereby affecting its downstream gene expression and playing an important role in tumors.

LINC00641 overexpression.⁶⁴ Likewise, ATG12 targets miR-378a and their expression is inversely correlated, thereby promoting tumor cell progression.³³ Moreover, overexpressed miR-378a facilitates cell cycle progression, reduces apoptosis, and promotes cell growth by attenuating ST7L expression. miR-378a upregulation also activates the Wnt/ β -catenin pathway to regulate tumor progression.⁶⁵

However, miR-378a-3p is lower in the serum and tissues of CC patients than in the serum and tissues of healthy control subjects and normal tissues, leading to the poor prognosis of patients with CC.²⁶ In this study, high miR-378a-3p expression reduced the proliferation and migration of CC cells both *in vitro* and *in vivo*. In short, miR-378a functions as an onco-miRNA by being regulated by circ-RNAs and then regulating its downstream targets and pathways, which is crucial in the malignancy of gynecological tumors. However, due to some inconsistent results, more studies are needed to elucidate the exact function of miR-378a in gynecological tumors.

Melanoma

Melanoma is the most aggressive type of skin cancer with high mortality.⁶⁶ The miRNA miR-378a shows the highest upregulation in advanced melanoma patients and those with lymph node metastasis³⁵ and miR-378a-5p is upregulated in metastatic melanoma patients, especially those resistant to targeted

therapy. High miR-378a-5p expression enhances cell invasion, migration, and activates EMT in melanoma cells, and promotes angiogenesis. Further, several studies implementing various approaches have demonstrated that miR-378a-5p plays a carcinogenic role by regulating downstream targets such as SUFU, FUS-1, KLF9, STAMPB, and HOXD10.⁶⁷ However, when miR-378a targets the 3' UTR of FOXN3, it mitigates the stimulation of miR-378a for proliferation, migration, and invasion. Moreover, FOXN3 interacts with β -catenin to downregulate the Wnt/ β -catenin signaling proteins,⁵³ and this pathway is crucial for cell proliferation, self-regeneration, differentiation, and tissue homeostasis.⁶⁸⁻⁷⁰

Other Cancers

Apart from the positive function of miR-378a in the tumors mentioned above, similar results have been reported in nasopharyngeal carcinoma (NPC), osteosarcoma, cholangiocarcinoma, renal carcinoma, Burkitt lymphoma (BL), acute lymphoblastic leukemia (AML), and chronic lymphoblastic leukemia (CML). In NPC, miR-378a is upregulated in NPC tissues and cell lines.³⁴ Elevating the expression of miR-378a dramatically promotes the capability of NPC to proliferate, migrate, and invade *in vitro*, as well as grow *in vivo* by restraining the expression of the TOB2 transducer. TOB2 is a potential tumor suppressor which inhibits cell proliferation by arresting

the progression of the G0/G1 phase cells to the S phase.⁷¹ In osteosarcoma cells and patient-derived tumor specimens, overexpression of miR-378a promotes osteosarcoma cell proliferation by diminishing the levels of KLF9. Forced upregulation of KLF9 extraneously reverses the tumor proliferation-promoting effect of miR-378a.³⁶ miR-378a-3p upregulation can repress cell invasion and suppress EMT by suppressing BYSL.⁷²

Elevated miR-378a exerts a similar promoting effect in cholangiocarcinoma, which is significantly associated with an advanced TNM stage, positive lymph node metastasis, and short OS, and a miR-378a knockout inhibits cell proliferation, migration, and invasion.³⁷ Similarly, miR-378a was identified as a diagnostic biomarker when renal carcinoma cancer tissues and cells showed increased proliferation,⁷³⁻⁷⁶ and increased miR-378a inhibited POLR2A and RUNX2 expression and subsequently promoted cell apoptosis.⁷⁷

In BL, miR-378a-3p can facilitate tumor growth by targeting IRAK4 and MNT.³⁹ miR-378a dysregulation is also detected in non-solid tumors, such as AML and CML. The upregulation of miR-378a is positively correlated with poor survival in AML patients.³⁸ The 5' flanking region of miR-378a is hypomethylated in AML. Thus, miR-378a can be reactivated by demethylation after 5-aza-dC treatment, but this is unlikely to provide helpful prognostic information in AML patients.⁷⁸ Knockdown of LINC00641 inhibited cell malignancy and promoted apoptosis by downregulating miR-378a by promoting ZBTB20 expression in AML.⁷⁹ Besides, lower FUS-1 expression, whose expression inversely correlates with miR-378a, is linked to the poor prognosis of AML patients.⁸⁰ In CML, enhanced expression of miR-378a in leukemia cell line K562 facilitates cell proliferation, clonality, and drug resistance to 5-FU, boosts expression of stem cell self-renewal markers OCT4 and c-Myc, and suppresses apoptosis.²¹ As in AML, FUS-1 was also identified as a target of miR-378a in CML, but its role has not been elucidated yet.²¹ Together, miR-378a overexpression exerts a stimulative effect on tumors mentioned above by regulating its different targets.

Downregulation of miR-378a in Several Malignancies Suggestive of its Antitumorigenic Roles

Unlike the cancers overexpressing miR-378a, it is worth noting that miR-378a is downregulated in many cancers. An inverse relationship between miR-378a expression and tumors progression has been reported in various carcinomas, including gastric cancer (GC), colorectal cancer (CRC), liver cancer, glioma, prostate cancer (PCa), BC, MB, pituitary cancer, oral squamous carcinoma (OSCC), bladder cancer, esophageal carcinoma (ESCC), RMS, and RB.

Gastric Cancer

GC has a high incidence worldwide and severely affects the quality of life.⁸¹ Several studies have explored the underlying

molecular alterations of miR-378a mediating tumorigenesis and GC development. A marked reduction of miR-378a was detected in GC tissues and cells compared to their controls.²⁴ Underexpression of miR-378a led to poor prognosis and malignant clinicopathologic features in GC patients. Circulating miR-378a level is considered the best biomarker for GC detection since it has a higher sensitivity and specificity than that of the serum levels of CEA and CA199.^{82,83} Low miR-378a expression in GC patients is also associated with an advanced TNM stage, poor tumor differentiation, high lymph node metastasis rate, and poor OS and DFS. Further, GC cells are arrested in the G0/G1 or G2/M phase after miR-378 overexpression.⁴⁰

miR-378a regulates various targets to exert its function as a tumor suppressor. For example, miR-378a targets BMP2, which has a high expression in GC patients with a shorter OS and DFS. Reverse-validation experiment confirmed that miR-378a overexpression inhibits invasion, migration, and EMT of GC by modulating BMP2, suggesting that components of the miR-378a/BMP2 axis might be potential therapeutic targets of GC.²⁴ Additionally, two other targets of miR-378a in GC—MAPK1 and VEGF—are closely associated with cancer malignancy. miR-378 inhibits MAPK1 in GC.⁸⁴ Depletion of MAPK1 by RNA interference in MC-803 cells reduces cell growth, increases apoptosis, and suppresses cell migration and invasion, which corresponded to the tumor-inhibiting effects caused by miR-378a restoration. Furthermore, miR-378a can directly bind to GAPLINC and reduce its expression, thereby repressing MAPK1 expression.⁵⁴ Upregulated miR-378a suppresses MAPK1 expression, cell proliferation, and cell cycle progression in GC cells, while these effects are reversed by increasing the expression of the lncRNA GAPLINC. Another miR-378 target in GC is VEGF,⁸⁵ as confirmed by the evidence that VEGF is downregulated by exogenously overexpressed miR-378.⁴⁰ Together, these findings suggest that miR-378 regulates GC development and can act as a promising biomarker for the treatment of GC patients.

Colorectal Cancer

Decreased miR-378a expression significantly correlates with shorter OS in CRC patients.^{23,86} Hence, miR-378a is a potential biomarker for the early detection and diagnosis of patients with CRC.^{23,87,88} Interestingly, miR-378a might also function as a novel biomarker to predict the efficacy of vaccines against CRC.^{89,90} The overexpression of miR-378a in CRC repressed cell growth,²³ induced apoptosis,⁸⁶ and inhibited migration and invasion by restraining EMT.⁵⁵ Studies also showed that miR-378a mitigated the malignant phenotypes of CRC cells by inhibiting the Wnt/ β -catenin pathway.⁹¹

Two lncRNAs were found to regulate miR-378a-3p. SP1-activated LINC00339 decreased miR-378a-3p but enhanced MED19 expression, contributing to cell proliferation, cell cycle progression, migration, and invasion in CRC cells.⁹¹ Further, the lncRNA CYTOR decreased miR-378a-5p and led

to the increase in SERPINE1, which inhibited L-OHP resistance and EMT.⁹²

Several other genes are the downstream targets of miR-378a, including IGF1R, CDC40, vimentin, BRAF, SDAD1, and KISS1. IGF1R protein expression significantly negatively correlated with miR-378a in CRC tissues.⁸⁶ Further, luciferase reporter assays confirmed CDC40 as a direct target of miR-378a.⁴¹ miR-378a represses cell growth and G1/S transition in CRC cells, and consistent with the inhibitory effect of miR-378a on tumor growth, CDC40 knockdown reduces proliferation and represses cell cycle progression by regulating G1/S and G2/M phases and pre-mRNA splicing.

High miR-378a expression restrains CRC cell growth and invasion and downregulates vimentin while silencing miR-378a produces the inverse effects.²³ miR-378 diametrically targets the 3' UTR of vimentin to inhibit tumor progression. Similarly, BRAF⁵⁵ and SDAD1⁹³ were significantly reduced by miR-378a-5p overexpression in CRC. SDAD1 expression increased upon blocking miR-378a, suggesting that miR-378a and SDAD1 have a direct but negative regulatory relationship. Further, the expression of KiSS1 is enhanced in CRC cells with miR-378a elevation, leading to decreased proliferation, migration, and invasion abilities of the cancer cells.⁹⁴

Liver Cancer

Low expression of miR-378a⁹⁵⁻⁹⁷ is related to a poor prognosis of liver cancer.⁹⁷ Enhanced miR-378a expression inhibits the proliferation and invasion⁹⁸ but increases the apoptosis⁹⁹ in liver cancer. Moreover, miR-378a elevated the sensitivity of sorafenib treatment by targeting VEGFR, PDGFR β , and c-Raf.⁹⁸

miR-378a also enhances hepatocellular carcinoma (HCC) development by downregulating FUS expression,¹⁰⁰ while miR-378a-5p targets VEGF.¹⁰¹ Moreover, miR-378a targets TRAF1 and weakens NF- κ B signaling, consequently downregulating VEGF.⁹⁷ IGF1R was also identified as a novel target of miR-378a in sorafenib-resistant HCC cells, with high miR-378a expression activating the PI3K/AKT and the Ras/Raf/MAP kinase pathways to mediate cell survival¹⁰² and IGF1R knockout significantly restrained the miR-378a-mediated sorafenib resistance. miR-378a also suppresses HCC tumorigenesis by downregulating PD-L1 and STAT3 expression.¹⁰³ Moreover, the overexpressed miR-378a suppressed HCC cell proliferation by arresting G2/M and inhibited tumor growth *in vivo*.⁴² Interestingly, in HCC, circCRIM1 acts as an upstream regulator of miR-378a that upregulates SKP2 by acting as a sponge for miR-378a and facilitates cell proliferation, angiogenesis, and the transition from the G1 to the S phase in the cell cycle.¹⁰⁴

Glioma

Even with the advancement of therapeutic strategies, glioma still shows a poor prognosis due to its high tumor aggressiveness and unlimited proliferation.^{105,106} Several clinical studies

in glioma have shown that miR-378a is remarkably decreased in cancer tissue and can potentially act as a diagnostic biomarker.¹⁰⁷⁻¹⁰⁹ Glioma patients with lower miR-378a expression showed poorer OS, and miR-378a overexpression restrained cell migration and invasion.¹⁰⁹ Downregulated miR-378a increased the target IRG1 expression, which enhanced glioma cell growth, invasion, migration, and EMT.¹⁰⁸ Another study showed that TSPAN17 was a direct target of miR-378a and correlated with poor prognosis in glioblastoma patients. Overexpressed miR-378a weakened TSPAN17 expression, consequently promoting apoptosis and reducing proliferation, migration, and invasion. These effects were attenuated in rescue experiments increasing TSPAN17 expression.¹¹⁰ Interestingly, interfilamentous vimentin can also mediate miR-378a self-renewal by regulating SOX2 transcription factor expression.¹¹¹ Further, miR-378a promotes cell survival, tumor growth, and angiogenesis by targeting SUFU and FUS-1 expression.¹¹²

Prostate Cancer

miR-378a is notably underexpressed in PCa tissues.⁴⁴ The reduction of miR-378a levels resulted in higher Gleason score, larger diameter tumors, and elevated serum prostate-specific antigen in PCa.^{113,114} miR-378a improved risk stratification based on the Gleason score or tumor stage and led to a higher risk of recurrence in patients with low miR-378a levels. Thus, miR-378a is a promising independent predictor of short-term recurrence in patients at high and very high risk of recurrence.¹¹³

CircRNA PDHX boosts PCa progression by sponging miR-378a.¹¹⁵ MiR-378a also inhibits prostate cell proliferation and glucose metabolism by repressing GLUT1.¹¹⁴ Forced miR-378a overexpression represses prostate cancer cell migration and invasion but promotes cell apoptosis *in vitro*.⁴⁴ Further, studies have shown that KLK2, KLK4,¹¹³ and MAPK1 are the targets of miR-378a in PCa, and the ectopic expression of MAPK1 rescues miR-378a-inhibited cell migration and invasion capacity.⁴⁴

Breast Cancer

Even in BC, miR-378a shows a low expression and correlates with the unfavorable prognosis of BC patients upon tamoxifen treatment.¹¹⁶ GOLT1A is regulated by miR-378a and is critical for the mechanisms underlying BC endocrine resistance.¹¹⁶ The m6A-mediated increase in LINC00958 expression enhanced tumorigenesis through the miR-378a/YY1 pathway.¹¹⁷ Further, DCTPP1 is modulated by miR-378a, and decreased miR-378a expression elevates the DCTPP1 level, which facilitates cell proliferation by activating DNA repair signaling.¹¹⁸

RUNX1 is significantly increased upon inhibition of miR-378a expression, which also enhances the invasion and migration of the triple-negative breast cancer (TNBC) cells. Further, the regulation of RUNX1 is mediated by the PPARGC1B/miR-378a/RUNX1 regulatory pathway.⁴⁵ It has

also been revealed that lncRNA GAS5 and miR-378a bind to each other, and the target of miR-378a-5p, SUFU, promotes GAS5-induced apoptosis of TNBC cells. This implicates that GAS5 stimulates apoptosis in TNBC cells by regulating miR-378a-5p/SUFU signaling.¹¹⁹ Moreover, although no significant effect of miR-378a was found on KLK4 expression, both miR-378a and KLK4 represent unfavorable prognostic markers in TNBC patients.¹²⁰

Other Cancers

In addition to the above-mentioned tumors, decreased miR-378a expression has also been reported in other cancers. miR-378a is significantly reduced in MB. miR-378a negatively regulates UHRF1 expression in MB by binding to the 3' UTR of its transcript.⁴⁶ Therefore, miR-378a overexpression inhibits UHRF1, but increased expression of UHRF1 reverses the miR-378a-induced suppression of cell proliferation and promotion of apoptosis.

In PA, RNF31 is highly expressed, and its expression is negatively regulated by miR-378a.⁴⁷ Knocking out RNF31 inhibits the proliferation and migration of the pituitary tumor cell line GH3. Similarly, miR-378a plays an inhibitory role in oral squamous cell carcinoma (OSCC). miR-378a is downregulated¹²¹ in OSCC and also targets KLK4.⁴⁸ Overexpressed KLK4 reverses the miR-378a-induced suppression of migration and invasion by enhancing MMP-9, MMP-2, and N-cadherin expression while decreasing E-cadherin levels. CircRNA-100290 can also rescue the miR-378a-induced suppression of GLUT1 by functioning as a competing endogenous RNA (ceRNA), thereby accelerating cell proliferation and glycolysis in OSCC.¹²² NCAPG, a target of miR-378a in OSCC, promotes cell proliferation and cell cycle progression but inhibits apoptosis by activating the GSK-3 β / β -catenin signaling in OSCC.¹²³

In bladder cancer, miR-378a is downregulated and can act as an independent prognostic factor for the OS and recurrence.⁴⁹ LINC00958, an upstream regulator of miR-378, promotes the proliferation and metastasis of bladder cancer cells by increasing IGF1R, a downstream target of miR-378a, by sponging miR-378a.¹²⁴ miR-378a is lower in esophageal squamous cell carcinoma (ESCC) cells and tissues than in normal cells and adjacent tissues. Low miR-378a expression leads to an unfavorable prognosis and shorter OS in ESCC patients. Further, low miR-378a levels elevate cell proliferation, migration, and invasion.⁵⁰ However, miR-378a overexpression exerts the opposite effect by inhibiting Rab10.¹²⁵ Mechanistically, the lncRNAs SLC2A1-AS1 and LINC00514 are essential in ESCC progression by acting as miR-378a sponges. SLC2A1-AS1 inhibits miR-378a-5p by regulating glycolysis, thereby restoring GLUT1 expression and enhancing ESCC.¹²⁶ LINC00514, acting as a ceRNA, upregulates SPHK1 by sponging miR-378a-5p and activates adipogenesis-related pathways, thereby promoting the proliferation and invasion of ESCC cells.¹²⁷

In a RMS-derived cell line, miR-378a upregulation inhibited IGF1R expression and impacted phosphorylated-Akt protein levels.⁵¹ Moreover, ectopic expression of miR-378a modulated

apoptosis, cell migration, cytoskeleton organization, and the expression of the muscle markers MyoD1, MyoR, desmin, and MyHC in RMS. DNA demethylation by 5-aza-2-deoxycytidine (5-aza-dC) also elevates miR-378a levels which correlate with increased apoptosis, cell viability reduction, and cell cycle arrest in G2-phase.⁵¹ In RB, upregulated miR-378a-3p inhibits cell proliferation by targeting FOXG1.⁵²

The Role of miR-378a in Anti-Cancer Therapeutic Resistance

Unfortunately, cancer cells cumulatively get resistant to miscellaneous anti-tumor therapies, leading to disease recurrence and metastasis. Hence, understanding the internal molecular mechanism of cancer drug resistance is essential for finding new therapeutic approaches. Given that miR-378 is involved in various biological processes, the abnormal expression of miR-378a might be responsible for the anti-cancer resistance. For instance, miR-378a expression is preternatural in chemotherapy resistance in esophageal cancer cells.¹²⁸ Many studies have also shown miR-378a as a promising candidate for anti-tumor treatment. In lung cancer, elevated miR-378a recovered CDDP chemosensitivity by targeting SCLU and downregulating Bcl-2, pCas-3, p-Erk1/2, and p-Akt.²⁵ In ovarian cancer, low miR-378a expression can elevate the efficacy of bevacizumab treatment in patients with recurrent ovarian cancer, thereby lengthening PFS. Further, multivariate analysis showed miR-378a level as an efficient predictor of PFS after anti-angiogenic therapy.⁶²

In CRC cells with upregulated miR-378a, sensitivity to cetuximab was restored in all BRAF mutants and half of KRAS mutants, and lauric acid improved the sensitization of Cetuximab in KRAS/BRAF mutated CRC cells by regaining miR-378a expression.¹²⁹ Moreover, EPA-induced upregulation of miR-378 led to the significant restoration of sensitivity to cetuximab in the KRAS-mutant cells.¹³⁰ In glioma, the inhibitory effect of curcumin was enhanced in miR-378a-expressing stable U87 cells.⁴³ Likewise, increased miR-378a expression enhances radiation response by promoting radiation-induced TGD in glioblastoma cells.¹⁰⁷ However, reduced miR-378a expression was found in sorafenib-resistant HCC cells than in the sorafenib-sensitive group. Further, LXR stimulated miRNA-378a-3p transcription and hence can be used as a potential combinable treatment strategy with sorafenib to suppress HCC progression.¹⁰² Metformin also induces miR-378a to downregulate CDK1, leading to suppression of cell proliferation and induction of the G2/M cell cycle arrest in HCC.⁴² Moreover, the biologically active component isolated from the fungus *Ganoderma lucidum* can also overcome the drug resistance conferred by miR-378.¹³¹

Conclusion

Cancer is the leading cause of death worldwide. The high mortality rate of cancers can be ascribed to its inefficient early

detection, inherent or acquired therapeutic resistance, and poor predictive power of conventional screening techniques and clinicopathological parameters. Hence, searching for novel molecular targets is necessary to solve this problem.

The miRNA miR-378a is a promising biomarker and therapeutic target for cancers. As discussed in this paper, miR-378a expression is aberrant in more than 23 cancers and is responsible for affecting the cell's capability to proliferate, invade, metastasize, and resist multiple anti-tumor treatment regimens. However, there is still some controversy and ambiguity over its expression and roles in some cancers. miR-378a plays a dual role in tumors and can produce different impacts on different tumors based on whether it is overexpressed or underexpressed. The fact that it acts as an onco-miRNA makes it an attractive therapeutic target in lung cancer,^{22,25,27-29} CC,³³ OC,³⁰⁻³² NPC,³⁴ melanoma,³⁵ osteosarcoma,³⁶ cholangiocarcinoma,³⁷ AML,³⁸ CML,²¹ renal carcinoma,⁷³⁻⁷⁶ and BL.³⁹ However, it acts as a tumor suppressor in GC,⁸⁴ CRC,⁴¹ liver cancer,⁴² glioma,⁴³ PCa,⁴⁴ BC,⁴⁵ MB,⁴⁶ PA,⁴⁷ OSCC,⁴⁸ bladder cancer,⁴⁹ ESCC,⁵⁰ RMS,⁵¹ and RB.⁵² Notably, the expression and role of miR-378a are still controversial in malignant tumors, including lung cancer⁶⁰ and CC.²⁶ All these effects may be due to the different expression of the two strands of miR-378a and it is also due to the different specific expression of miR-378a in tissues and cells of different tumors. Further, the differences and limitations in the source of cell and tissue samples might be the key influencing factors.

The expression of circulating miR-378a can be used for cancer screening and diagnosis.^{22-24,31,33} The identification of the signaling pathways involved in cell proliferation,³⁴ migration,³⁵ invasion,³³ and apoptosis⁴¹ also provides evidence for the different functions of miR-378a. miR-378a has also shown value as a therapy sensitizer²⁵ and drug target.^{25,131} All the results mentioned above might be due to the regulation of miR-378a by various factors, such as circRNA and lncRNA, and the interaction between miR-378a and its targets that constitute complex regulatory networks. Depending on the cellular environment, miR-378a might have various biological functions and clinical applications and can even be expressed differently and have varied roles in different tumors.

Because of the dual function of miR-378a in cancer development and whether targeting miR-378a is an effective and feasible approach for individual tumor patient-based treatment, more detailed investigations and understanding of the mechanisms underlying how miR-378a regulates tumor progression and therapy resistance are required in the future. Different analysis methods and intervention strategies for the role of the miRNA in different cancers are essential. Further, considering that miR-378a can effectively regulate the response of cancer cells to chemotherapy^{25,128} and radiotherapy,⁴³ a combination of miR-378a targeted therapy and chemotherapy or radiotherapy might achieve better therapeutic effects in some cancer types.

Thus, in this review, we provide insights into the rationale underlying the dual functions of miRNAs in tumor onset and progression. We show that miR-378 is critical for key biological and pathological processes via its complex network regulation

mechanism in human cancers. In the future, miR-378 has great potential to become a clinically effective approach for cancer diagnosis and prognosis by refining cancer types and subtypes and an effective therapeutic strategy against cancer.

Author Contributions

YQ and RL conceived the idea, YQ and LL performed literature retrieval, YQ conducted literature reading and sorting, YQ wrote initial article and revised the manuscript, RL provided helpful ideas and comments on this article, PL played an essential role in manuscript revision and responding to comments, XZ supervised the study. All the authors have read and approved the final version of this article. YQ and RL have contributed equally to this work as co-first author.


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References

- Hutvagner G, Zamore PD. A microRNA in a multiple-turnover RNAi enzyme complex. *Science*. 2002;297(5589):2056-2060.
- He L, Hannon GJ. MicroRNAs: Small RNAs with a big role in gene regulation. *Nat Rev Genet*. 2004;5(7):522-531.
- Rosalind C, Lee RLF, Ambrost V. The *C. elegans* heterochronic gene *lin-4* encodes small RNAs with antisense complementarity to *lin-4*.pdf. *Cell*. 1993;75(5):843-854.
- Shukla V, Varghese V, Kabekkodu S, et al. Enumeration of deregulated miRNAs in liquid and tissue biopsies of cervical cancer. *Gynecol Oncol*. 2019;155(1):135-143.
- Wang CH, Lee DY, Deng Z, et al. MicroRNA miR-328 regulates zonation morphogenesis by targeting CD44 expression. *PLoS One*. 2008;3(6):e2420.
- Kawasaki H, Taira K. Retraction: Hes1 is a target of microRNA-23 during retinoic-acid-induced neuronal differentiation of NT2 cells. *Nature*. 2003;426(6962):100.
- Chan JA, Krichevsky AM, Kosik KS. MicroRNA-21 is an anti-apoptotic factor in human glioblastoma cells. *Cancer Res*. 2005;65(14):6029-6033.
- Xu C LY, Pan Z, Chu W, et al. The muscle-specific microRNAs miR-1 and miR-133 produce opposing effects on apoptosis by targeting HSP60, HSP70 and caspase-9 in cardiomyocytes. *J Cell Sci*. 2011;124(Pt 18):3187.
- Shatseva T, Lee DY, Deng Z, Yang BB. MicroRNA miR-199a-3p regulates cell proliferation and survival by targeting caveolin-2. *J Cell Sci*. 2011;124(Pt 16):2826-2836.

10. Croce CM, Calin GA. miRNAs, cancer, and stem cell division. *Cell*. 2005;122(1):6-7.
11. Mello C, Czech M. Micromanaging insulin secretion. *Nat Med*. 2004;10(12):1297-1298.
12. Chellappan P, Vanitharani R, Fauquet C. MicroRNA-binding viral protein interferes with Arabidopsis development. *Proc Natl Acad Sci U S A*. 2005;102(29):10381-10386. doi:10.1073/pnas.0504439102.
13. Iorio M, Croce C. Causes and consequences of microRNA dysregulation. *Cancer journal (Sudbury, Mass)*. 2012;18(3):215-222.
14. Visone R, Croce C. MiRNAs and cancer. *Am J Pathol*. 2009;174(4):1131-1138.
15. Wu W, Lee C, Cho C, et al. MicroRNA dysregulation in gastric cancer: A new player enters the game. *Oncogene*. 2010;29(43):5761-5771.
16. Labbaye C, Testa U. The emerging role of MIR-146A in the control of hematopoiesis, immune function and cancer. *J Hematol Oncol*. 2012;5:13.
17. Kasashima K, Nakamura Y, Kozu T. Altered expression profiles of microRNAs during TPA-induced differentiation of HL-60 cells. *Biochem Biophys Res Commun*. 2004;322(2):403-410.
18. Carrer M, Liu N, Grueter C, et al. Control of mitochondrial metabolism and systemic energy homeostasis by microRNAs 378 and 378*. *Proc Natl Acad Sci U S A*. 2012;109(38):15330-15335.
19. Eichner L, Perry M, Dufour C, et al. miR-378(*) mediates metabolic shift in breast cancer cells via the PGC-1 β /ERR γ transcriptional pathway. *Cell Metab*. 2010;12(4):352-361.
20. Yu J, Kong X, Liu J, et al. Expression profiling of PPAR γ -regulated microRNAs in human subcutaneous and visceral adipogenesis in both genders. *Endocrinology*. 2014;155(6):2155-2165.
21. Ma J, Wu D, Yi J, et al. MiR-378 promoted cell proliferation and inhibited apoptosis by enhanced stem cell properties in chronic myeloid leukemia K562 cells. *Biomedicine Pharmacotherapy = Biomedecine Pharmacotherapie*. 2019;112:108623.
22. Chen L, Xu S, Xu H, Zhang J, Ning J, Wang S. MicroRNA-378 is associated with non-small cell lung cancer brain metastasis by promoting cell migration, invasion and tumor angiogenesis. *Med Oncol (Northwood, London, England)*. 2012;29(3):1673-1680.
23. Zhang G, Zhou H, Xiao H, Li Y, Zhou T. MiR-378 is an independent prognostic factor and inhibits cell growth and invasion in colorectal cancer. *BMC Cancer*. 2014;14:109.
24. Yang Y, Luo S, Wang L. Effects of microRNA-378 on epithelial-mesenchymal transition, migration, invasion and prognosis in gastric carcinoma by targeting BMP2. *Eur Rev Med Pharmacol Sci*. 2019;23(12):5176-5186.
25. Chen X, Jiang Y, Huang Z, et al. miRNA-378 reverses chemoresistance to cisplatin in lung adenocarcinoma cells by targeting secreted clusterin. *Sci Rep*. 2016;6:19455.
26. Zhang L, Wu Z. MicroRNA-378a-3p downregulation as a novel biomarker with poor clinical outcomes in cervical cancer. *Biomed Environ Sci: BES*. 2021;34(3):213-221.
27. Ho C, Noor S, Nagoor N. RBX1MiR-378 And MiR-1827 regulate tumor invasion, migration and angiogenesis in human lung adenocarcinoma by targeting and, respectively. *J Cancer*. 2018;9(2):331-345.
28. Ji K, Cui F, Qu D, et al. MiR-378 promotes the cell proliferation of non-small cell lung cancer by inhibiting FOXG1. *Eur Rev Med Pharmacol Sci*. 2018;22(4):1011-1019.
29. Skrzypek K, Tertilt M, Golda S, et al. Interplay between heme oxygenase-1 and miR-378 affects non-small cell lung carcinoma growth, vascularization, and metastasis. *Antioxid Redox Signaling*. 2013;19(7):644-660.
30. Heldin C, Moustakas A. Role of Smads in TGF β signaling. *Cell Tissue Res*. 2012;347(1):21-36.
31. Kim Y, Kim E, Jeon D, et al. Differential microRNA expression signatures and cell type-specific association with Taxol resistance in ovarian cancer cells. *Drug Des Devel Ther*. 2014;8:293-314.
32. Wang J, Li Y, Zhou J, Shen F, Shi X, Chen Y. CircATRNL1 activates Smad4 signaling to inhibit angiogenesis and ovarian cancer metastasis via miR-378. *Mol Oncol*. 2021;15(4):1217-1233.
33. Tan D, Zhou C, Han S, Hou X, Kang S, Zhang Y. MicroRNA-378 enhances migration and invasion in cervical cancer by directly targeting autophagy-related protein 12. *Mol Med Rep*. 2018;17(5):6319-6326.
34. Yu B, Peng X, Zhao F, et al. MicroRNA-378 functions as an onco-miR in nasopharyngeal carcinoma by repressing TOB2 expression. *Int J Oncol*. 2014;44(4):1215-1222.
35. Sun M, Ma X, Tu C, et al. MicroRNA-378 regulates epithelial-mesenchymal transition and metastasis of melanoma by inhibiting FOXN3 expression through the Wnt/ β -catenin pathway. *Cell Biol Int*. 2019;43(10):1113-1124.
36. Peng N, Miao Z, Wang L, Liu B, Wang G, Guo X. MiR-378 promotes the cell proliferation of osteosarcoma through downregulating the expression of Kruppel-like factor 9. *Biochem Cell Biol = Biochimie Biologie Cellulaire*. 2018;96(5):515-521.
37. Zhou Z, Ma J. miR-378 serves as a prognostic biomarker in cholangiocarcinoma and promotes tumor proliferation, migration, and invasion. *Cancer Biomarkers: Section A of Disease Markers*. 2019;24(2):173-181.
38. Qian J, Lin J, Qian W, et al. Overexpression of miR-378 is frequent and may affect treatment outcomes in patients with acute myeloid leukemia. *Leuk Res*. 2013;37(7):765-768.
39. Niu F, Dzikiewicz-Krawczyk A, Koerts J, et al. MiR-378a-3p is critical for Burkitt lymphoma cell growth. *Cancers*. 2020;12(12):3546.
40. Deng H, Guo Y, Song H, et al. MicroRNA-195 and microRNA-378 mediate tumor growth suppression by epigenetic regulation in gastric cancer. *Gene*. 2013;518(2):351-359.
41. Wang K, Ma J, Zhang F, Yu M, Xue J, Zhao J. MicroRNA-378 inhibits cell growth and enhances L-OHP-induced apoptosis in human colorectal cancer. *IUBMB life*. 2014;66(9):645-654.
42. Zhou J, Han S, Qian W, Gu Y, Li X, Yang K. Metformin induces miR-378 to downregulate the CDK1, leading to suppression of cell proliferation in hepatocellular carcinoma. *Oncotargets Ther*. 2018;11:4451-4459.
43. Li W, Yang W, Liu Y, et al. MicroRNA-378 enhances inhibitory effect of curcumin on glioblastoma. *Oncotarget*. 2017;8(43):73938-73946.

44. Chen Q, Zhou W, Han T, et al. MiR-378 suppresses prostate cancer cell growth through downregulation of MAPK1 in vitro and in vivo. *Tumour Biol: J Int Soc Oncodevelop Biol Med.* 2016;37(2):2095-2103.
45. Browne G, Dragon J, Hong D, et al. MicroRNA-378-mediated suppression of Runx1 alleviates the aggressive phenotype of triple-negative MDA-MB-231 human breast cancer cells. *Tumour Biol: J Int Soc Oncodevelop Biol Med.* 2016;37(7):8825-8839.
46. Zhang Z, Zhu B, Zhao X, et al. Regulation of UHRF1 by microRNA-378 modulates medulloblastoma cell proliferation and apoptosis. *Oncol Rep.* 2017;38(5):3078-3084.
47. Qiu P, Xu T, Lu X, Yang W, Zhang Y, Xu G. MicroRNA-378 regulates cell proliferation and migration by repressing RNF31 in pituitary adenoma. *Oncol Lett.* 2018;15(1):789-794.
48. Cui Z, Sun S, Liu Q, et al. MicroRNA-378-3p/5p suppresses the migration and invasiveness of oral squamous carcinoma cells by inhibiting KLK4 expression. *Biochem Cell Biol = Biochimie Biologie Cellulaire.* 2020;98(2):154-163.
49. Zaravinos A, Radojicic J, Lambrou G, et al. Expression of miRNAs involved in angiogenesis, tumor cell proliferation, tumor suppressor inhibition, epithelial-mesenchymal transition and activation of metastasis in bladder cancer. *J Urol.* 2012;188(2):615-623.
50. Jin W, Wang L, Cheng S, Lv H. Prognostic value of microRNA-378 in esophageal cancer and its regulatory effect on tumor progression. *Exp Ther Med.* 2021;22(1):704.
51. Megiorni F, Cialfi S, McDowell H, et al. Deep Sequencing the microRNA profile in rhabdomyosarcoma reveals down-regulation of miR-378 family members. *BMC Cancer.* 2014;14:880.
52. Zhang C, Wu S. microRNA – 378a-3p restrains the proliferation of retinoblastoma cells but promotes apoptosis of retinoblastoma cells via inhibition of FOXG1. *Invest Ophthalmol Visual Sci.* 2020;61(5):31.
53. Dai Y, Wang M, Wu H, Xiao M, Liu H, Zhang D. Loss of FOXN3 in colon cancer activates beta-catenin/TCF signaling and promotes the growth and migration of cancer cells. *Oncotarget.* 2017;8(6):9783-9793.
54. Diao L, Wang S, Sun Z. Long noncoding RNA GAPLINC promotes gastric cancer cell proliferation by acting as a molecular sponge of miR-378 to modulate MAPK1 expression. *Oncotargets Ther.* 2018;11:2797-2804.
55. Wang Z, Ma B, Ji X, et al. MicroRNA-378-5p suppresses cell proliferation and induces apoptosis in colorectal cancer cells by targeting BRAF. *Cancer Cell Int.* 2015;15:40.
56. Li R, Liu Y, Wang T, et al. The characteristics of lung cancer in Xuanwei County: A review of differentially expressed genes and noncoding RNAs on cell proliferation and migration. *Biomedicine Pharmacotherapy = Biomedecine Pharmacotherapie.* 2019;119:109312.
57. Zhang Y, Xu H. Serum exosomal miR-378 upregulation is associated with poor prognosis in non-small-cell lung cancer patients. *J Clin Lab Anal.* 2020;34(6):e23237.
58. Gao S, Yu Y, Liu L, Meng J, Li G. Circular RNA hsa_circ_0007059 restrains proliferation and epithelial-mesenchymal transition in lung cancer cells via inhibiting microRNA-378. *Life Sci.* 2019;233:116692.
59. Ying K, Wang L, Long G, Lian C, Chen Z, Lin W. ACTA2-AS1 Suppresses lung adenocarcinoma progression via sequestering miR-378a-3p and miR-4428 to elevate SOX7 expression. *Cell Biol Int.* 2020;44(12):2438-2449.
60. Wang M, Sun X, Yang Y, Jiao W. Long non-coding RNA OIP5-AS1 promotes proliferation of lung cancer cells and leads to poor prognosis by targeting miR-378a-3p. *Thorac Cancer.* 2018;9(8):939-949.
61. Mao J, Zhang Z, Chen Y, et al. Sea cucumber peptides inhibit the malignancy of NSCLC by regulating miR-378a-5p targeted TUSC2. *Food Funct.* 2021;12(24):12362-12371.
62. Chan J, Kiet T, Blansit K, et al. MiR-378 as a biomarker for response to anti-angiogenic treatment in ovarian cancer. *Gynecol Oncol.* 2014;133(3):568-574.
63. Wei X, Lv H, Yang S, Yang X. CircRNA PLOD2 enhances ovarian cancer propagation by controlling miR-378. *Saudi J Biol Sci.* 2021;28(11):6260-6265.
64. Zhang Y, Yu R, Li L. LINC00641 Hinders the progression of cervical cancer by targeting miR-378a-3p/CPEB3. *J Gene Med.* 2020;22(9):e3212.
65. Li S, Yang F, Wang M, Cao W, Yang Z. miR-378 functions as an onco-miRNA by targeting the ST7L/Wnt/ β -catenin pathway in cervical cancer. *Int J Mol Med.* 2017;40(4):1047-1056.
66. Siegel R, Miller K, Jemal A. Cancer statistics, 2017. *CA Cancer J Clin.* 2017;67(1):7-30.
67. Tupone M, D'Aguzzo S, Di Martile M, et al. microRNA-378a-5p is a novel positive regulator of melanoma progression. *Oncogenesis.* 2020;9(2):22.
68. Huo J, Zhang Y, Li R, Wang Y, Wu J, Zhang D. Upregulated MicroRNA-25 mediates the migration of Melanoma cells by targeting DKK3 through the WNT/ β -Catenin pathway. *Int J Mol Sci.* 2016;17(11):1124.
69. Kovacs D, Migliano E, Muscardin L, et al. The role of Wnt/ β -catenin signaling pathway in melanoma epithelial-to-mesenchymal-like switching: Evidences from patients-derived cell lines. *Oncotarget.* 2016;7(28):43295-43314.
70. Wang E, Wang D, Li B, et al. Capn4 promotes epithelial-mesenchymal transition in human melanoma cells through activation of the Wnt/ β -catenin pathway. *Oncol Rep.* 2017;37(1):379-387.
71. Winkler G. The mammalian anti-proliferative BTG/Tob protein family. *J Cell Physiol.* 2010;222(1):66-72.
72. Zhang J, Tang H, Jiang X, Huang N, Wei Q. Hypoxia-induced miR-378a-3p Inhibits osteosarcoma invasion and epithelial-to-mesenchymal via *BYSL* transition regulation. *Front Genet.* 2021;12:804952.
73. Fedorko M, Stanik M, Iliev R, et al. Combination of MiR-378 and MiR-210 serum levels enables sensitive detection of renal cell carcinoma. *Int J Mol Sci.* 2015;16(10):23382-9.
74. Hauser S, Wulfken L, Holdenrieder S, et al. Analysis of serum microRNAs (miR-26a-2*, miR. 191, miR-337-3p and miR-378) as potential biomarkers in renal cell carcinoma. *Cancer Epidemiol.* 2012;36(4):391-3914.
75. Redova M, Poprach A, Nekvindova J, et al. Circulating miR-378 and miR-451 in serum are potential biomarkers for renal cell carcinoma. *J Transl Med.* 2012;10:55.

76. Li H, Li J, Wang Z, et al. Identification of angiogenesis-related miRNAs in a population of patients with renal clear cell carcinoma. *Oncol Rep*. 2014;32(5):2061-2069.
77. Weng W, Yu K, Jhan J, Pang S, Chiou Y, Pang S. Micro-RNA378a-3p induces apoptosis in sarcomatoid renal cell carcinoma and regulates POLR2A and RUNX2 expression. *Anticancer Res*. 2022;42(2):811-825.
78. Zhu X, Wen X, Zhang Y, et al. The 5' flanking region of miR-378 is hypomethylated in acute myeloid leukemia. *Int J Clin Exp Pathol*. 2015;8(5):4321-4331.
79. Wang J, Liu Z, Yu L. Long non-coding RNA LINC00641 promotes cell growth and migration through modulating miR-378a/ZBTB20 axis in acute myeloid leukemia. *Eur Rev Med Pharmacol Sci*. 2019;23(17):7498-7509.
80. Tang X, Zhou J, Zhang J, et al. Low expression of FUS1 is negatively correlated with miR-378 and may predict adverse prognoses in acute myeloid leukemia. *Acta Haematol*. 2018;139(2):89-95.
81. Lordick F, Janjigian Y. Clinical impact of tumour biology in the management of gastroesophageal cancer. *Nature Rev Clin Oncol*. 2016;13(6):348-360.
82. Guo J, Miao Y, Xiao B, et al. Differential expression of microRNA species in human gastric cancer versus non-tumorous tissues. *J Gastroenterol Hepatol*. 2009;24(4):652-657.
83. Liu H, Zhu L, Liu B, et al. Genome-wide microRNA profiles identify miR-378 as a serum biomarker for early detection of gastric cancer. *Cancer Lett*. 2012;316(2):196-203.
84. Fei B, Wu H. MiR-378 inhibits progression of human gastric cancer MGC-803 cells by targeting MAPK1 in vitro. *Oncol Res*. 2012;20(12):557-564.
85. Hua Z, Lv Q, Ye W, et al. MiRNA-directed regulation of VEGF and other angiogenic factors under hypoxia. *PLoS One*. 2006;1(1):e116.
86. Li H, Dai S, Zhen T, et al. Clinical and biological significance of miR-378a-3p and miR-378a-5p in colorectal cancer. *European Journal Of Cancer (Oxford, England : 1990)*. 2014;50(6):1207-1221. doi:10.1016/j.ejca.2013.12.010.
87. Peng J, Xie Z, Cheng L, et al. Paired design study by real-time PCR: miR-378* and miR-145 are potent early diagnostic biomarkers of human colorectal cancer. *BMC Cancer*. 2015;15:158.
88. Zanutto S, Pizzamiglio S, Ghilotti M, et al. Circulating miR-378 in plasma: A reliable, haemolysis-independent biomarker for colorectal cancer. *Br J Cancer*. 2014;110(4):1001-1007.
89. Tanaka H, Hazama S, Iida M, et al. miR-125b-1 and miR-378a are predictive biomarkers for the efficacy of vaccine treatment against colorectal cancer. *Cancer Sci*. 2017;108(11):2229-2238.
90. Shindo Y, Hazama S, Nakamura Y, et al. miR-196b, miR-378a, and miR-486 are predictive biomarkers for the efficacy of vaccine treatment in colorectal cancer. *Oncol Lett*. 2017;14(2):1355-1362.
91. Ye H, Li W, Wu K, et al. The SP1-induced long noncoding RNA, LINC00339, promotes tumorigenesis in colorectal cancer via the miR-378a-3p/MED19 axis. *Onco Targets Ther*. 2020;13:11711-11724.
92. Yang J, Ma Q, Zhang M, Zhang W. LncRNA CYTOR drives L-OHP resistance and facilitates the epithelial-mesenchymal transition of colon carcinoma cells via modulating miR-378a-5p/SERPINE1. *Cell Cycle (Georgetown, Tex)*. 2021;20(14):1415-1430.
93. Zeng M, Zhu L, Li L, Kang C. miR-378 suppresses the proliferation, migration and invasion of colon cancer cells by inhibiting SDAD1. *Cell Mol Biol Lett*. 2017;22:12.
94. Zheng Y, Liu Y, Lin Y, et al. MicroRNA-124 and microRNA-378 inhibit the proliferation and invasion of colorectal cancer by upregulating KiSS1. *Transl Cancer Res*. 2020;9(4):2838-2846.
95. An J, Liu J, Liu L, et al. A genetic variant in primary miR-378 is associated with risk and prognosis of hepatocellular carcinoma in a Chinese population. *PLoS One*. 2014;9(4):e93707.
96. Qian F, Wang J, Wang Y, et al. MiR-378a-3p as a putative biomarker for hepatocellular carcinoma diagnosis and prognosis: Computational screening with experimental validation. *Clin Transl Med*. 2021;11(2):e307.
97. Zhu B, Chen J, Feng Y, et al. DNMT1-induced miR-378a-3p silencing promotes angiogenesis via the NF- κ B signaling pathway by targeting TRAF1 in hepatocellular carcinoma. *J Exp Clin Cancer Res: CR*. 2021;40(1):352.
98. Fu H, Zhang J, Pan T, Ai S, Tang L, Wang F. Mir-378a enhances the sensitivity of liver cancer to sorafenib by targeting VEGFR, PDGFR β and c-Raf. *Mol Med Rep*. 2018;17(3):4581-4588.
99. Wang J, Li Y, Ma Q, Huang J. Mir-378 in combination with ultrasonic irradiation and SonoVue microbubbles transfection inhibits hepatoma cell growth. *Mol Med Rep*. 2020;21(6):2493-2501.
100. Ma J, Lin J, Qian J, et al. MiR-378 promotes the migration of liver cancer cells by down-regulating Fus expression. *Cell Physiol Biochem: Int J Exp Cell Physiol, Biochem, Pharm*. 2014;34(6):2266-2274.
101. Zou H, Yang L. miR-378a-5p improved the prognosis and suppressed the progression of hepatocellular carcinoma by targeting the VEGF pathway. *Transl Cancer Res*. 2020;9(3):1558-1566.
102. Lin Z, Xia S, Liang Y, et al. LXR Activation potentiates sorafenib sensitivity in HCC by activating microRNA-378a transcription. *Theranostics*. 2020;10(19):8834-8850.
103. Li Y, Zhou T, Cheng X, Li D, Zhao M, Zheng W. microRNA-378a-3p regulates the progression of hepatocellular carcinoma by regulating PD-L1 and STAT3. *Bioengineered*. 2022;13(3):4730-4743.
104. Ji Y, Yang S, Yan X, et al. CircCRIM1 promotes hepatocellular carcinoma proliferation and angiogenesis by sponging miR-378a-3p and regulating SKP2 expression. *Front Cell Dev Biol*. 2021;9:796686.
105. Onishi M, Ichikawa T, Kurozumi K, Date I. Angiogenesis and invasion in glioma. *Brain Tumor Pathol*. 2011;28(1):13-24.
106. Xue S, Hu M, Li P, et al. Relationship between expression of PD-L1 and tumor angiogenesis, proliferation, and invasion in glioma. *Oncotarget*. 2017;8(30):49702-49712.
107. Li W, Liu Y, Yang W, et al. MicroRNA-378 enhances radiation response in ectopic and orthotopic implantation models of glioblastoma. *J Neuro-Oncol*. 2018;136(1):63-71.
108. Shi H, Wang D, Sun X, Sheng L. MicroRNA-378 acts as a prognosis marker and inhibits cell migration, invasion and epithelial-mesenchymal transition in human glioma by targeting IRG1. *Eur Rev Med Pharmacol Sci*. 2018;22(12):3837-3846.

109. Li B, Wang Y, Li S, et al. Decreased expression of miR-378 correlates with tumor invasiveness and poor prognosis of patients with glioma. *Int J Clin Exp Pathol.* 2015;8(6):7016-7021.
110. Guo X, Zhang X, Chen P, Ma L, Shen Z. Mir-378a-3p inhibits cellular proliferation and migration in glioblastoma multiforme by targeting tetraspanin 17. *Oncol Rep.* 2019;42(5):1957-1971.
111. Deng Z, Du W, Fang L, et al. The intermediate filament vimentin mediates microRNA miR-378 function in cellular self-renewal by regulating the expression of the Sox2 transcription factor. *J Biol Chem.* 2013;288(1):319-331.
112. Lee D, Deng Z, Wang C, Yang B. MicroRNA-378 promotes cell survival, tumor growth, and angiogenesis by targeting SuFu and Fus-1 expression. *Proc Natl Acad Sci U S A.* 2007;104(51):20350-20355.
113. Avgeris M, Stravodimos K, Scorilas A. Loss of miR-378 in prostate cancer, a common regulator of KLK2 and KLK4, correlates with aggressive disease phenotype and predicts the short-term relapse of the patients. *Biol Chem.* 2014;395(9):1095-1104.
114. Cannistraci A, Hascoet P, Ali A, et al. MiR-378a inhibits glucose metabolism by suppressing GLUT1 in prostate cancer. *Oncogene.* 2022;41(10):1445-1455.
115. Mao Y, Li W, Hua B, et al. Circular RNA_PDHX promotes the proliferation and invasion of prostate cancer by sponging MiR-378a-3p. *Front Cell Dev Biol.* 2020;8(602707).
116. Ikeda K, Horie-Inoue K, Ueno T, et al. miR-378a-3p modulates tamoxifen sensitivity in breast cancer MCF-7 cells through targeting GOLT1A. *Sci Rep.* 2015;5(13170).
117. Rong D, Dong Q, Qu H, et al. mA-induced LINC00958 promotes breast cancer tumorigenesis via the miR-378a-3p/YY1 axis. *Cell Death Discov.* 2021;7(1):27.
118. Niu M, Shan M, Liu Y, et al. DCTPP1, an oncogene regulated by miR-378a-3p, promotes proliferation of breast cancer via DNA repair signaling pathway. *Front Oncol.* 2021;11(641931).
119. Zheng S, Li M, Miao K, Xu H. lncRNA GAS5-promoted apoptosis in triple-negative breast cancer by targeting miR-378a-5p/SUFU signaling. *J Cell Biochem.* 2020;121(3):2225-2235.
120. Gong W, Zhu C, Liu Y, et al. Elevated levels of both microRNA 378 (miR-378) and kallikrein-related peptidase 4 (KLK4) mRNA are associated with an unfavorable prognosis in triple-negative breast cancer. *Am J Transl Res.* 2021;13(3):1594-1606.
121. Scapoli L, Palmieri A, Lo Muzio L, et al. MicroRNA expression profiling of oral carcinoma identifies new markers of tumor progression. *Int J Immunopathol Pharmacol.* 2010;23(4):1229-1234.
122. Chen X, Yu J, Tian H, et al. Circle RNA hsa_circRNA_100290 serves as a ceRNA for miR-378a to regulate oral squamous cell carcinoma cells growth via glucose transporter-1 (GLUT1) and glycolysis. *J Cell Physiol.* 2019;234(11):19130-19140.
123. Li J, Sun S, Li J, et al. NCAPG, mediated by miR-378a-3p, regulates cell proliferation, cell cycle progression, and apoptosis of oral squamous cell carcinoma through the GSK-3 β / β -catenin signaling. *Neoplasma.* 2021;68(6):1201-1211.
124. Cui Y, Xie M, Zhang Z. LINC00958 Involves in bladder cancer through sponging miR-378a-3p to elevate IGF1R. *Cancer Biother Radiopharm.* 2020;35(10):776-788.
125. Ding N, Sun X, Wang T, Huang L, Wen J, Zhou Y. Mir-378a-3p exerts tumor suppressive function on the tumorigenesis of esophageal squamous cell carcinoma by targeting Rab10. *Int J Mol Med.* 2018;42(1):381-391.
126. Liu H, Zhang Q, Song Y, et al. Long non-coding RNA SLC2A1-AS1 induced by GLI3 promotes aerobic glycolysis and progression in esophageal squamous cell carcinoma by sponging miR-378a-3p to enhance Glut1 expression. *J Exp Clin Cancer Res: CR.* 2021;40(1):287.
127. Wang X, Liu H, Zhang Q, et al. LINC00514 promotes lipogenesis and tumor progression in esophageal squamous cell carcinoma by sponging miR-378a-5p to enhance SPHK1 expression. *Int J Oncol.* 2021;59(5):86.
128. Hummel R, Sie C, Watson D, et al. MicroRNA signatures in chemotherapy resistant esophageal cancer cell lines. *World J Gastroenterol.* 2014;20(40):14904-14912.
129. Weng W, Leung W, Pang Y, Hsu H. Lauric acid can improve the sensitization of Cetuximab in KRAS/BRAF mutated colorectal cancer cells by retrievable microRNA-378 expression. *Oncol Rep.* 2016;35(1):107-116.
130. Weng W, Leung W, Pang Y, Kuo L, Hsu H. EPA Significantly improves anti-EGFR targeted therapy by regulating miR-378 expression in colorectal cancer. *Oncol Lett.* 2018;16(5):6188-6194.
131. Wu Q, Xie Y, Deng Z, et al. Ergosterol peroxide isolated from *Ganoderma lucidum* abolishes microRNA miR-378-mediated tumor cells on chemoresistance. *PloS One.* 2012;7(8):e44579.