

ORIGINAL RESEARCH

Gene Mutation Annotation and Pedigree for Pulmonary Arterial Hypertension Patients in Han Chinese Patients

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Background: The etiology of pulmonary arterial hypertension (PAH) in the Han Chinese population is poorly understood.

Objectives: The aim of this study was to assess gene variants and associated functional annotations for PAH in Han Chinese patients.

Methods: This is an ethnicity-based multi-centre study. Blood samples were collected from 20 PAH patients who volunteered for the study, and genetic tests were performed. The DAVID database was used to functionally annotate the genes *BMPR2*, *ALK1*, *KCNK3*, *CAV1*, and *ENG*. Associated diseases, functional categories, gene ontology, and protein interactions were analysed using the Functional Annotation Tool in the DAVID database. GEO and ClinVar databases were also used for further comparison with gene mutations in our study.

Results: PAH patient with gene mutations were female predominant except for a single male with a *BMPR2* mutation. Locus variants in our study included 'G410DfsX1' in *BMPR2*, 'ex7 L300P', 'ex4 S110PfsX40', and 'ex7 E295Afs96X' in *ALK1*, 'c.-2C>A (IVS1-2 C>A)' in *CAV1*, and 'ex8 D366Q' in *ENG* were not found in the ClinVar database associated with PAH. In addition to BMP and TGF- β pathways, gene ontology of input genes in the DAVID database also included pathways associated with nitric oxide signaling and regulation.

Conclusions: This Multi-centre study indicated that 'G410DfsX1' in *BMPR2*, 'ex7 L300P', 'ex4 S110PfsX40', 'ex7 E295Afs96X' in *ALK1*, 'c.-2C>A (IVS1-2 C>A)' in *CAV1*, and 'ex8 D366Q' in *ENG* were identified in Han Chinese patients with PAH. Females were more susceptible to PAH, and a relatively young age distribution was observed for patients with *BMPR2* mutations.

Keywords: ClinVar database; DAVID database; gene annotation; gene mutation; pulmonary arterial hypertension; heritable pulmonary arterial hypertension

Introduction

Pulmonary arterial hypertension (PAH) is defined by the presence of pre-capillary pulmonary hypertension, with right heart catheterisation showing mean pulmonary arterial pressure ≥ 20 mmHg, pulmonary artery wedge pressure ≤ 15 mmHg and a pulmonary vascular resistance > 3 Wood units [1, 2]. The National Organization for Rare Disorders (NORD) classified pulmonary hypertension into three subtypes, including idiopathic pulmonary arterial hypertension (IPAH), heritable pulmonary hypertension (HPH), and associated pulmonary hypertension [3]. Patients with a family history of PAH were grouped under the term 'heritable

PAH (HPAH) in group 1 PAH. This is an autosomal-dominant vascular disorder that predominantly affects pulmonary arterioles [4, 5]. Furthermore, genetic mutations have been identified in sporadic primary PAH [6]. Some gene variants were considered to have potential effects on individual susceptibility to pulmonary hypertension for Chinese Han Chinese [3], suggesting that the variants of the PAH gene may be associated with the development of PAH.

Bone morphogenetic protein receptor type II (*BMPR2*) gene mutation is the single most common causal factor for HPAH, however, approximately 25% of idiopathic PAH patients have pathogenic mutations without prior family history of disease [7, 8]. Previous large survey confirmed that *BMPR2* (15.3%), *ACVRL1* (activin receptor-like kinase 1 (*ALK1*)) (0.9%), *ENG* (endoglin) (0.6%), and *KCNK3* (potassium channel sub-family K member 3) (0.4%) are causal mutations of PAH [9]. *CAV1* (caveolin-1) functions to physically colocalize BMP receptors, and is associated with both lipodystrophy and PAH [10, 11]. Although the mutated gene has been identified in white or Hispanic patients, these mutations in Han Chinese patients with PAH remain to be elucidated. The aim of this study is to analyse the gene variants and their associated functional annotations in Han Chinese PAH patients.

Methods

Data source and study population

This is a multi-centre ethnicity-based study that investigates PAH gene mutations in the Han Chinese. Twenty PAH patients were enrolled into this study. Informed consent was obtained from all participants and their family members for collection of blood samples and genetic analysis. The Institutional Review Board (IRB) of Kaohsiung Veterans General Hospital approved this study (IRB number: KSVGH21-CT1-21).

Whole exome sequencing, alignment, variant calling and annotation

Genomic DNA was isolated from peripheral blood leukocytes of PAH patients. Polymerase chain reaction (PCR) was used to amplify the exons and flanking intronic bases of the five genes, including *BMPR2* (13 exons), *ALK1* (10 exons), *CAV1* (3 exons), *ENG* (14 exons), and *KCNK3* (2 exons). The primers used for PCR were designed using reference sequences deposited in the GenBank database. Standard DNA sequencing reactions were performed using the fluorescence-labelled dideoxy chain termination method with the Big-Dye Terminator ABI Prism Kit and the ABI PRISM™ 3700 DNA Analyser (Applied Biosystems, Foster City, CA, USA), according to the manufacturer's instructions.

Gene mutation and expression analysis

The DAVID database (<https://david.ncicrf.gov>) was used to functionally annotate mutated genes of PAH patients in this study [12–13]. *BMPR2*, *ALK1*, *KCNK3*, *CAV1*, and *ENG* were entered into the gene list, with 'Official_Gene_Symbol' as the selected identifier, and human/homo sapiens as the selected species; background population was set as homo sapiens. Associated diseases, functional categories, gene ontology, and protein interaction were analysed using the Functional Annotation Tool in the DAVID database. The aforementioned genes were also entered into the GEO profiles database, where search results on mutations and expressions related to PAH were manually reviewed. Pulmonary hypertension was also used as a key word for the ClinVar database [14]; genetic locus mutations were downloaded from the website in text format, which were transformed to Excel format for further comparisons with gene mutations in our study.

Statistical analyses

The SPSS version 22 (IBM, Chicago, IL, USA) was used for data analysis. Percentile values were used to express categorical data, and were analysed using the chi-square test. Mean (μ) and standard deviation values were used for continuous variables using the Student's unpaired *t*-tests. A *p*-value of < 0.05 (–Log *P* value > 1.3) was considered to be statistically significant.

Results

Patient characteristics

All patients' basic characteristics are listed in **Table 1**. There was one variant at 'G410DfsX1' in *BMPR2*, as well as three locus variants in *ALK1*, at 'ex7 L300P,' 'ex4 S110PfsX40,' and 'ex7 E295Afs96X.' In addition, one variant at 'c.-2C>A (IVS1–2 C>A)' in *CAV1*, and one at 'ex8 D366Q' in *ENG* were also found. There were 11 patients without gene mutations, whereas patient 15 (P15) had gene mutations at both ex7 L300P in *ALK1*

Table 1: Basic characteristics and gene mutations for pulmonary artery hypertension patients.

Number	Sex	Age	BMPR2	ALK-1	KCNK3	CAV1	ENG
1	M	54	(-)	(-)	(-)	(-)	(-)
2	F	67	(-)	(-)	(-)	(-)	(-)
3	F	55	(-)	(-)	(-)	ex3 A216P	(-)
4	F	37	(-)	(-)	(-)	(-)	(-)
5	F	48	(-)	(-)	(-)	(-)	(-)
6	F	49	(-)	(-)	(-)	(-)	(-)
7	F	39	(-)	(-)	(-)	(-)	(-)
8	F	36	(-)	(-)	(-)	(-)	(-)
9	F	45	(-)	(-)	(-)	(-)	(-)
10	F	71	(-)	(-)	(-)	(-)	(-)
11	F	43	(-)	(-)	c.-2C>A (IVS1-2 C>A)	(-)	(-)
12	M	48	(-)	(-)	(-)	(-)	ex8 D366Q
13	M	25	G410DfsX1	(-)	(-)	(-)	(-)
14	F	57	(-)	(-)	(-)	(-)	(-)
15	F	48	(-)	ex7 L300P	(-)	(-)	ex8 D366Q
16	F	62	(-)	ex4 S110PfsX40	(-)	(-)	(-)
17	F	56	(-)	(-)	(-)	(-)	(-)
18	F	42	(-)	ex7 E295Afs96X	(-)	(-)	(-)
19	F	51	(-)	(-)	(-)	(-)	(-)
20	F	39	(-)	(-)	(-)	(-)	ex8 D366Q

and D366Q in *ENG*. **Table 2** shows the comparison between gene subgroups, including gender, age, body height, and body weight. Out of the 20 patients, there were three (15%) males and 17 (85%) females, with a mean age of 48.6 ± 11.1 . There was a 25-year-old male in the *BMPR2* group, who was considerably younger than patients in other groups.

DAVID database analysis

The DAVID database was used for functional annotation, and the output data are shown in **Figure 1**. Mutated genes in this study, including *BMPR2*, *ALK1*, *KCNK3*, *CAV1*, and *ENG*, are displayed in the gene list. Commonly associated diseases for these mutations included pulmonary hypertension (-Log P value: 1.9), liver cirrhosis, and associated hepatopulmonary syndrome (-Log P value: 2.8) (Fig. 1A). Mutated genes and their associated proteins included transforming growth factor beta receptor 1 (*TGF- β 1*) (-Log P value: 2.0), bone morphogenetic protein 7 (*BMP7*) (Log P value: 2.6), and activin A receptor type 1 (*ACVR1*) (-Log P value: 2.7), as shown in **Figure 1B**.

Gene cluster and ontologies were analysed, and the results are shown in **Figure 2**. All mutated genes in this study were found in the gene cluster of 'disease mutation' (-Log P value: 2.72) (**Figure 2A**). Moreover, gene ontologies of input genes included 'negative regulation of nitric-oxide synthase activity' (-Log P value: 2.85), 'negative regulation of endothelial cell proliferation' (-Log P value: 2.28), 'positive regulation of BMP signalling pathway' (-Log P value: 2.26), 'positive regulation of pathway-restricted Smad protein phosphorylation' (-Log P value: 2.07), 'vasculogenesis' (-Log P value: 2.00), 'negative regulation of TGF- β receptor signalling pathway' (-Log P value: 1.96), 'regulation of cell proliferation' (-Log P value: 1.48), and 'BMP binding' (-Log P value: 2.74), as shown in **Figure 2B**.

GEO profile database analysis

GEO profile database associated with gene mutations and expressions was shown in **Figure 3**. The data discussed in **Figure 3A** were deposited into the NCBI's Gene Expression Omnibus (Edgar et al., 2002),

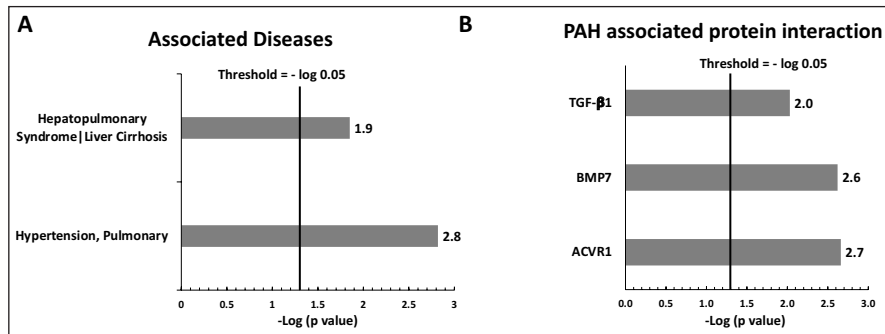


Figure 1: The DAVID database was used for functional annotation. Panel A. Mutated genes in this study, including *BMPR2*, *ALK1*, *KCNK3*, *CAV1*, and *ENG* were entered into the gene list. Common associated diseases included pulmonary hypertension, liver cirrhosis, as well as associated hepatopulmonary syndrome. Panel B. Mutated genes in our study and their associated protein interactions. Activin A receptor type 1 (ACVR1); bone morphogenetic protein 7 (BMP7); transforming growth factor beta receptor 1 (TGF-β1).

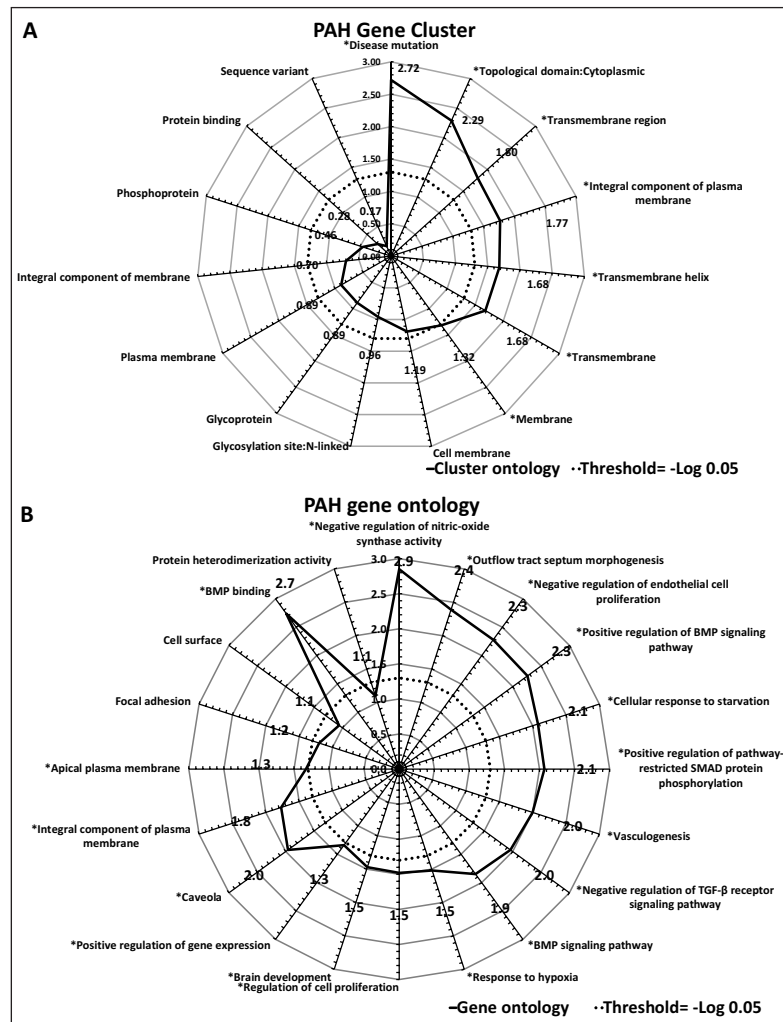


Figure 2: Gene cluster and ontology outputs of the five locus variants, as analysed by the DAVID database. Panel A. All mutated genes in this study belonged to the 'disease mutation' gene cluster (-log P value: 2.72). Panel B. PAH gene ontology of mutated genes in this study. Gene ontology reports 'negative regulation of nitric-oxide synthase activity' (-log P value: 2.85), 'negative regulation of endothelial cell proliferation' (-log P value: 2.28), 'positive regulation of BMP signalling pathway' (-log P value: 2.26), 'positive regulation of pathway-restricted Smad protein phosphorylation' (-log P value: 2.07), 'vasculogenesis' (-log P value: 2.00), 'negative regulation of TGF-β receptor signalling pathway' (-log P value: 1.96), 'regulation of cell proliferation' (-log P value: 1.48), and 'BMP binding' (-log P value: 2.74). Bone morphogenetic protein (BMP); transforming growth factor beta receptor 1 (TGF-β1).

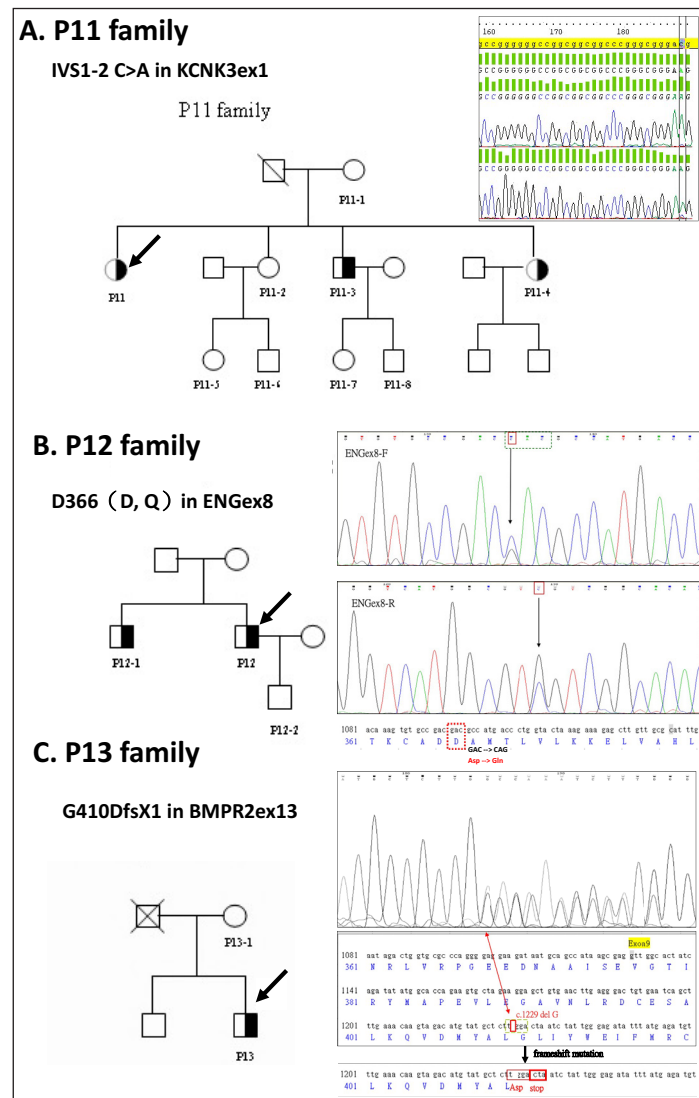


Figure 4: Location of gene mutation and family pedigree of volunteer PAH patients. Arrows indicate PAH patients in our study. Panel A. Gene mutation location of P11 is at IVS1–2 C>A in the KCNK3ex1 fragment. Panel B. P12 has a gene mutation located at D366(D, Q) in the ENGex8 fragment. Panel C. P13 has a gene mutation located at G410DfsX1 in the BMPR2ex13 fragment.

hypertension, liver cirrhosis, and associated hepatopulmonary syndrome. Mutated genes were involved in TGF- β , BMP7, and ACVR1 interactions.

Gene mutation for PAH was associated with age and gender difference

Compared with PAH patients without *BMPR2* mutations, those with *BMPR2* mutations were younger, with a mean age of 35.4 years [8]. Previous study indicated a mean age of 42 years for *BMPR2* non-carriers [8]. Patients with gene mutations other than *BMPR2* in our study were older than previously reported (49.7–62.8 years in *ALK1*, *KCNK3*, *CAV1*, and *ENG* subgroups). Our study also indicated that patients with *BMPR2* mutations were younger than patients with other gene mutations.

The occurrence of *BMPR2* mutations in sporadic PAH cases without a family history can be attributed to low penetrance of *BMPR2* mutations (20%–30%) and *de novo* mutations [15]. The estimated penetrance in male and female carriers is 14% and 42%, respectively [15, 16]. It has been shown that female is the single most important determinant for the penetrance of *BMPR2* mutations in PAH [15, 16]; male patients were significantly more likely to have *BMPR2* mutations than female patients [17]. Similarly, in our study, only one male was found to have the *BMPR2* mutation.

PAH occurs predominantly in females; the sex ratio of female-to-male is 2.4:1. It has been suggested that oestrogen and its metabolism may be associated with the pathogenesis of PAH [16, 18, 19]. However, mortality rate of PAH is higher in males as compared with that of females, particularly in male *BMPR2* mutation carriers [20]. The Registry to Evaluate Early and Long-Term Pulmonary Arterial Hypertension Disease Management

(REVEAL) showed a higher female predominance of PAH irrespective of *BMPR2* status, and found that the female-to-male ratio for PAH was 3.9:1 in races other than whites, black and Hispanic [21]. Our study results agreed with previous evidence that suggested female dominance in IPAH or HPAH (**Table 1**) [17].

Gene ontologies from the Han Chinese patients correlated with PAH mechanism

Mutated genes in our study, including *BMPR2*, *ALK1*, *KCNK3*, *CVA1*, and *ENG* were considered to have evidence of mutation in patients with PAH [22]. Among which, *BMPR2*, *ALK1* and *ENG* were clearly recognized for their biological functions in PAH [22]. *BMPR2* is particularly highly expressed on the cell surface of pulmonary vascular endothelium [23], and *BMP9* functions as a circulating vascular quiescence factor to counterbalance cell apoptosis and excessive proliferation in endothelial cells [24]. In *BMPR2* mutated pulmonary artery smooth muscle cells, there was a loss of antiproliferative effects from BMPs, resulting in excessive Smad1/5 signalling, which led to hyper-proliferative cells [24]. The *BMPR2* protein forms a complex with *ALK1*; *ENG* plays a co-receptor to form a complex on the membrane and signal specifically in response to the circulating BMP ligands [25]. *CAV1* is highly expressed in endothelial cells, and is an important constituent protein of caveolae. BMP receptors are localized in caveolae, and loss of *CAV1* inhibits *BMPR2* membrane localization and signalling [26, 27]. *KCNK3* encodes a potassium channel that generates the membrane potential needed to regulate pulmonary vascular tone [28]. *BMPR2*, composed of an extracellular motif and transmembrane kinase domains, and is a part of the TGF- β receptor superfamily [29]; mutated genes associated with the TGF- β signalling pathway include *ALK1*, *CVA1*, and *ENG* [30]. All of the genes analyzed in our study encode for membrane or transmembrane proteins [22].

The TGF- β pathway processes angiogenesis via two distinct signalling pathways, including the ALK5-Smad2/3 pathway and the ALK1-Smad1/5/8 pathway [31]. Endoglin counterbalances the stabilizing role of ALK5 to stabilise the vessel and inhibit endothelial cell overproliferation [32]. However, *BMPR2* or *ACVRL1* mutation disrupt the SMAD1/5/8 pathway and BMP signalling. This inhibits SMC apoptosis, which leads to SMC proliferation, vascular remodelling, and ultimately results in PAH [33]. Our study (**Figure 2**) reported gene ontologies, including 'positive regulation of BMP signaling pathway' and 'negative regulation of TGF- β signaling pathway,' which confirmed the imbalance between the ALK5-Smad2/3 pathway and ALK1-Smad1/5/8 pathways in these gene mutations, resulting in PAH. Gene ontology of input genes in our study also suggested that in addition to their roles in BMP and TGF- β pathways, they also function in pathways associated with nitric oxide signalling and regulation, which coincides with the pathobiology of PAH; the results of this study was supported by a previous study [22].

New pathogenic mutations for IPAH might exist in the Han Chinese population

There are differences in genetic, physiological, and anatomic factors between races, which affected the structure and function of the right ventricle [34]. However, studies focused on the Han Chinese are limited. The REVEAL trial enrolled only 3.3% Asians, and it showed a higher PAH prevalence in Hispanic patients as compared with that of earlier registries [21]. A Chinese registry reported a lower 1-year survival rate for Chinese PAH patients [35]. Early molecular genetics studies can strengthen clinical diagnosis and assist decision-making in adopting effective treatment [36].

Locus variants in our study included 'G410DfsX1' in *BMPR2*, 'ex7 L300P,' 'ex4 S110PfsX40,' and 'ex7 E295Afs96X' in *ALK1*, 'c.-2C>A (IVS1-2 C>A)' in *CAV1*, and 'ex8 D366Q' in *ENG*, which were not found in the ClinVar database associated with PAH. Further studies are needed to determine whether these new mutations are associated with sporadic primary PAH or HPAH in the Han Chinese. This study also revealed similar gender and age distribution with previous large studies, which implies that other genetic mutations or environmental factors may contribute to the poor survival of Han Chinese PAH patients [35]. Further studies on offspring of these PAH patients may be needed to confirm the association between these new mutations in Han Chinese patients with PAH.

Study limitations

The sample size of this study was small, and not all family members of enrolled PAH patients received whole genome sequencing. Although the characteristics of patients included in this study were similar to those of previous large studies. Further investigations are needed for PAH patients from the Han Chinese population.

Study strengths

This is a multi-centre ethnicity-based study to determine gene mutations in PAH patients. Current trials are focused mostly on white or Hispanic patients; studies of PAH patients in the Han Chinese population are rare. This study focused on the Han Chinese and used databases, including DAVID, ClinVar, and GEO profiles

to analyse the relationship between mutated genes and their functional ontologies. This study offers valuable genetic data for the Han Chinese population.

Conclusions

This multi-centre ethnicity-based analysis revealed five new locus variants that is potentially associated with PAH in the Han Chinese, including 'G410DfsX1' in *BMPR2*, ex7 L300P,' 'ex4 S110PfsX40,' and 'ex7 E295Afs96X' in *ALK1*, 'c.-2C>A (IVS1-2 C>A)' in *CAV1*, and 'ex8 D366Q' in *ENG*. This study reports that females have greater susceptibility to PAH in the Han Chinese; moreover, in addition to BMP and TGF- β pathways, changes in nitric oxide signalling and regulation have also been reported to be associated with PAH. This study further enriched the gene database for PAH in the Han Chinese, which may be used to advance PAH therapy at the molecular genetic level.

Data Accessibility Statement

The data underlying this article cannot be shared publicly due to legal restrictions imposed by the government of Taiwan in relation to the 'Personal Information Protection Act.'

Ethics and Consent

All authors give their consent for publication.

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Competing Interests

The authors have no competing interests to declare.

Author Contributions

Concept and design: Huang WC, Chi PL.

Acquisition, analysis, or interpretation of data: Wang MT, Cheng CC, Hung CC.

Drafting of the manuscript: Wang MT.

Critical revision of the manuscript for important intellectual content: Huang WC.

Statistical analysis: Wang MT.

Administrative, technical, or material support: Charng MJ.

Supervision: Huang WC.

References

1. **Galiè N, Humbert M, Vachiery JL**, et al. 2015 ESC/ERS Guidelines for the diagnosis and treatment of pulmonary hypertension: The Joint Task Force for the Diagnosis and Treatment of Pulmonary Hypertension of the European Society of Cardiology (ESC) and the European Respiratory Society (ERS): Endorsed by: Association for European Paediatric and Congenital Cardiology (AEPC), International Society for Heart and Lung Transplantation (ISHLT). *Eur Heart J*. 2016; 37: 67–119. 1 September 2015. DOI: <https://doi.org/10.1093/eurheartj/ehv317>
2. **Galiè N, McLaughlin VV, Rubin LJ**, et al. An overview of the 6th World Symposium on Pulmonary Hypertension. *Eur Respir J*. 2019; 53. 16 December 2018. DOI: <https://doi.org/10.1183/13993003.02148-2018>
3. **Yin C, Li K, Yu Y**, et al. Genome-wide association study identifies loci and candidate genes for non-idiopathic pulmonary hypertension in Eastern Chinese Han population. *BMC Pulm Med*. 2018; 18: 158–158. DOI: <https://doi.org/10.1186/s12890-018-0719-0>
4. **Hoepfer MM, Bogaard HJ, Condliffe R**, et al. Definitions and diagnosis of pulmonary hypertension. *J Am Coll Cardiol*. 2013; 62: D42–50. 21 December 2013. DOI: <https://doi.org/10.1016/j.jacc.2013.10.032>
5. **Tuder RM, Archer SL, Dorfmueller P**, et al. Relevant issues in the pathology and pathobiology of pulmonary hypertension. *J Am Coll Cardiol*. 2013; 62: D4–12. 21 December 2013. DOI: <https://doi.org/10.1016/j.jacc.2013.10.025>
6. **Thomson JR, Machado RD, Pauciulo MW**, et al. Sporadic primary pulmonary hypertension is associated with germline mutations of the gene encoding BMPR-II, a receptor member of the TGF-beta family. *J Med Genet*. 2000; 37: 741–745. 4 October 2000. DOI: <https://doi.org/10.1136/jmg.37.10.741>

7. **Machado RD, Southgate L, Eichstaedt CA**, et al. Pulmonary Arterial Hypertension: A Current Perspective on Established and Emerging Molecular Genetic Defects. *Hum Mutat.* 2015; 36: 1113–1127. 22 September 2015. DOI: <https://doi.org/10.1002/humu.22904>
8. **Evans JD, Girerd B, Montani D**, et al. BMPR2 mutations and survival in pulmonary arterial hypertension: An individual participant data meta-analysis. *Lancet Respir Med.* 2016; 4: 129–137. 23 January 2016. DOI: [https://doi.org/10.1016/S2213-2600\(15\)00544-5](https://doi.org/10.1016/S2213-2600(15)00544-5)
9. **Gräfs S, Haimel M, Bleda M**, et al. Identification of rare sequence variation underlying heritable pulmonary arterial hypertension. *Nat Commun.* 2018; 9: 1416. 14 April 2018. DOI: <https://doi.org/10.1038/s41467-018-03672-4>
10. **Austin ED, Ma L, LeDuc C**, et al. Whole exome sequencing to identify a novel gene (caveolin-1) associated with human pulmonary arterial hypertension. *Circ Cardiovasc Genet.* 2012; 5: 336–343. 5 April 2012. DOI: <https://doi.org/10.1161/CIRCGENETICS.111.961888>
11. **Morrell NW, Aldred MA, Chung WK**, et al. Genetics and genomics of pulmonary arterial hypertension. *Eur Respir J.* 2019; 53. 14 December 2018. DOI: <https://doi.org/10.1183/13993003.01899-2018>
12. **Huang da W, Sherman BT, Lempicki RA.** Systematic and integrative analysis of large gene lists using DAVID bioinformatics resources. *Nat Protoc.* 2009; 4: 44–57. 10 January 2009. DOI: <https://doi.org/10.1038/nprot.2008.211>
13. **Huang da W, Sherman BT, Lempicki RA.** Bioinformatics enrichment tools: Paths toward the comprehensive functional analysis of large gene lists. *Nucleic Acids Res.* 2009; 37: 1–13. 27 November 2008. DOI: <https://doi.org/10.1093/nar/gkn923>
14. **Landrum MJ, Lee JM, Benson M**, et al. ClinVar: Improving access to variant interpretations and supporting evidence. *Nucleic Acids Res.* 2018; 46: D1062–D1067. 23 November 2017. DOI: <https://doi.org/10.1093/nar/gkx1153>
15. **Larkin EK, Newman JH, Austin ED**, et al. Longitudinal analysis casts doubt on the presence of genetic anticipation in heritable pulmonary arterial hypertension. *Am J Respir Crit Care Med.* 2012; 186: 892–896. 28 August 2012. DOI: <https://doi.org/10.1164/rccm.201205-0886OC>
16. **Austin ED, Cogan JD, West JD**, et al. Alterations in oestrogen metabolism: Implications for higher penetrance of familial pulmonary arterial hypertension in females. *European Respiratory Journal.* 2009; 34: 1093. DOI: <https://doi.org/10.1183/09031936.00010409>
17. **Ge X, Zhu T, Zhang X**, et al. Gender differences in pulmonary arterial hypertension patients with BMPR2 mutation: A meta-analysis. *Respiratory Research.* 2020; 21: 44. DOI: <https://doi.org/10.1186/s12931-020-1309-2>
18. **West J, Cogan J, Geraci M**, et al. Gene expression in BMPR2 mutation carriers with and without evidence of pulmonary arterial hypertension suggests pathways relevant to disease penetrance. *BMC Med Genomics.* 2008; 1: 45. 1 October 2008. DOI: <https://doi.org/10.1186/1755-8794-1-45>
19. **Mair KM, Yang XD, Long L**, et al. Sex affects bone morphogenetic protein type II receptor signaling in pulmonary artery smooth muscle cells. *Am J Respir Crit Care Med.* 2015; 191: 693–703. 22 January 2015. DOI: <https://doi.org/10.1164/rccm.201410-1802OC>
20. **Humbert M, Sitbon O, Chaouat A**, et al. Survival in patients with idiopathic, familial, and anorexigen-associated pulmonary arterial hypertension in the modern management era. *Circulation.* 2010; 122: 156–163. 30 June 2010. DOI: <https://doi.org/10.1161/CIRCULATIONAHA.109.911818>
21. **Frost AE, Badesch DB, Barst RJ**, et al. The Changing Picture of Patients With Pulmonary Arterial Hypertension in the United States: How REVEAL Differs From Historic and Non-US Contemporary Registries. *Chest.* 2011; 139: 128–137. DOI: <https://doi.org/10.1378/chest.10-0075>
22. **Garcia-Rivas G, Jerjes-Sánchez C, Rodriguez D**, et al. A systematic review of genetic mutations in pulmonary arterial hypertension. *BMC Med Genet.* 2017; 18: 82. 5 August 2017. DOI: <https://doi.org/10.1186/s12881-017-0440-5>
23. **David L, Mallet C, Mazerbourg S**, et al. Identification of BMP9 and BMP10 as functional activators of the orphan activin receptor-like kinase 1 (ALK1) in endothelial cells. *Blood.* 2006; 109: 1953–1961. DOI: <https://doi.org/10.1182/blood-2006-07-034124>
24. **Yang X, Long L, Southwood M**, et al. Dysfunctional Smad signaling contributes to abnormal smooth muscle cell proliferation in familial pulmonary arterial hypertension. *Circ Res.* 2005; 96: 1053–1063. 23 April 2005. DOI: <https://doi.org/10.1161/01.RES.0000166926.54293.68>
25. **Upton PD, Davies RJ, Trembath RC**, et al. Bone morphogenetic protein (BMP) and activin type II receptors balance BMP9 signals mediated by activin receptor-like kinase-1 in human pulmonary artery endothelial cells. *J Biol Chem.* 2009; 284: 15794–15804. 16 April 2009. DOI: <https://doi.org/10.1074/jbc.M109.002881>

26. **Bonor J, Adams EL, Bragdon B**, et al. Initiation of BMP2 signaling in domains on the plasma membrane. *J Cell Physiol.* 2012; 227: 2880–2888. DOI: <https://doi.org/10.1002/jcp.23032>
27. **Wertz JW, Bauer PM.** Caveolin-1 regulates BMPRII localization and signaling in vascular smooth muscle cells. *Biochem Biophys Res Commun.* 2008; 375: 557–561. 30 August 2008. DOI: <https://doi.org/10.1016/j.bbrc.2008.08.066>
28. **Ma L, Roman-Campos D, Austin ED**, et al. A Novel Channelopathy in Pulmonary Arterial Hypertension. *New England Journal of Medicine.* 2013; 369: 351–361. DOI: <https://doi.org/10.1056/NEJMoa1211097>
29. **Aldred MA, Comhair SA, Varella-Garcia M**, et al. Somatic chromosome abnormalities in the lungs of patients with pulmonary arterial hypertension. *Am J Respir Crit Care Med.* 2010; 182: 1153–1160. 29 June 2010. DOI: <https://doi.org/10.1164/rccm.201003-0491OC>
30. **Star GP, Giovinazzo M, Langleben D.** ALK2 and BMPR2 knockdown and endothelin-1 production by pulmonary microvascular endothelial cells. *Microvasc Res.* 2013; 85: 46–53. 13 November 2012. DOI: <https://doi.org/10.1016/j.mvr.2012.10.012>
31. **Vorselaars VMM, Hosman AE, Westermann CJJ**, et al. Pulmonary Arterial Hypertension and Hereditary Haemorrhagic Telangiectasia. *Int J Mol Sci.* 2018; 19. 20 October 2018. DOI: <https://doi.org/10.3390/ijms19103203>
32. **Li C, Hampson IN, Hampson L**, et al. CD105 antagonizes the inhibitory signaling of transforming growth factor beta1 on human vascular endothelial cells. *Faseb j.* 2000; 14: 55–64. 8 January 2000. DOI: <https://doi.org/10.1096/fasebj.14.1.55>
33. **van der Bruggen CE, Happé CM, Dorfmueller P**, et al. Bone Morphogenetic Protein Receptor Type 2 Mutation in Pulmonary Arterial Hypertension: A View on the Right Ventricle. *Circulation.* 2016; 133: 1747–1760. 18 February 2016. DOI: <https://doi.org/10.1161/CIRCULATIONAHA.115.020696>
34. **Medrek SK, Sahay S.** Ethnicity in Pulmonary Arterial Hypertension: Possibilities for Novel Phenotypes in the Age of Personalized Medicine. *Chest.* 2018; 153: 310–320. DOI: <https://doi.org/10.1016/j.chest.2017.08.1159>
35. **Jing Z-C, Xu X-Q, Han Z-Y**, et al. Registry and Survival Study in Chinese Patients With Idiopathic and Familial Pulmonary Arterial Hypertension. *Chest.* 2007; 132: 373–379. DOI: <https://doi.org/10.1378/chest.06-2913>
36. **Chiou KR, Charng MJ.** Genetic diagnosis of familial hypercholesterolemia in Han Chinese. *J Clin Lipidol.* 2016; 10: 490–496. 22 May 2016. DOI: <https://doi.org/10.1016/j.jacl.2016.01.009>

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