RP-HPLC method for simultaneous estimation of tenofovir disoproxil fumarate, lamivudine, and efavirenz in combined tablet dosage form

Abstract

Background: A simple, precise, accurate, and rapid reverse phase-high performance liquid chromatography (RP-HPLC) method with UV-Visible detector has been developed and subsequently validated for the simultaneous determination of tenofovir disoproxil fumarate (TDF), lamivudine (LAMI), and efavirenz (EFV) in their combined tablet dosage form. **Materials and Methods:** The separation was based on the use of a Kromasil C_{18} analytical column (150 × 4.6 mm, i.d., 5 μm). The mobile phase consisted of a mixture of 70 volumes of methanol and 30 volumes of 10 mM phosphate buffer (pH 5.0). The separation was carried out at 40°C temperature with a flow rate of 1 ml/min. **Results:** Quantitation was achieved with UV detection at 254 nm, with linear calibration curves at concentration ranges of 1–6 μg/ml for TDF and LAMI and 2–12 μg/ml for EFV. The recoveries obtained were 99.46–101.36% for LAMI, 99.57–101.42% for TDF, and 99.96–100.87 for EFV. **Conclusion:** The method was validated according to International conference of harmonisation guidelines in terms of accuracy, precision, specificity, robustness, limits of detection and quantitation, and other aspects of analytical validation.

Key words: Efavirenz, lamivudine, RP-HPLC, tenofovir disoproxil fumarate, validation

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INTRODUCTION

Tenofovir disoproxil fumarate (TDF) {9-[(R)-2-[[bis [[isopropoxycarbonyl] oxy] methoxy] phosphonyl] methoxy] popyl] adenine fumarate} [Figure 1] is a nucleotide analog reverse transcriptase inhibitor (NRTI) and is used for treating HIV infection in adults, in combination with other antiretroviral agents. Lamivudine (LAMI) {4-amino-1-[(2R,5S)-2-(hydroxyl methyl)-1,3-oxathiolan-5-yl]-1,2-dihydro pyrimidin-2-one} [Figure 2] is an NRTI used in the treatment of HIV infection and chronic hepatitis B virus (HBV). Efavirenz (EFV) [(4S)-6-chloro-4-(cyclopropylethynyl)-1,4-dihydro-4-(trifluoromethyl)-2H-3,1-benzoxazin-2-one] [Figure 3] is a non-nucleotide reverse transcriptase inhibitor (NNRTI) used in the combination treatment of HIV infection (AIDS).[1-6] The combination of TDF, LAMI, and EFV (300 mg TDF, 300 mg LAMI, and 600 mg EFV) was approved in July 2008 by Central Drug Standard Control Organization (CDSCO) and tentatively approved by US Food and Drug Administration (USFDA) on 9 March 2009 for the treatment of HIV infection in adults.^[7,8] Literature survey reveals that TDF is estimated individually by titrimetric, UV, reverse phase-high performance liquid chromatography (RP-HPLC) in tablet formulation and by RP-HPLC methods in human plasma. [9-14] Few UV, RP-HPLC, high performance thin layer chromatography (HPTLC), and liquid chromatography with tandem mass spectrometry (LC/MS/MS) methods have been reported for simultaneous estimation of emtricitabine and TDF in pharmaceutical formulation.[15-18] Similarly, estimation of LAMI by titrimetric, UV, HPTLC, RP-HPLC methods in tablet formulation and by RP-HPLC method in human plasma has been reported.[19-23] Few UV, RP-HPLC, HPTLC, and LC/MS/MS methods have been reported for the simultaneous estimation of LAMI with nevirapine, stavudine,

Figure 1: Chemical structure of tenofovir disoproxil fumarate

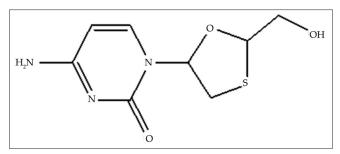


Figure 2: Chemical structure of lamivudine

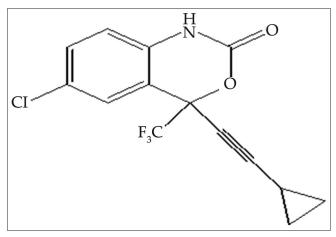


Figure 3: Chemical structure of efavirenz

and zidovudine.^[24-28] Also, for the estimation of EFV, UV, HPTLC, and isocratic HPLC methods in tablet formulation and HPLC method in human plasma have been reported.^[25-33] Also, RP-HPLC method is reported for the simultaneous estimation of EFV with TDF and emtricitabine.^[34] Only one method, that is UV spectrophotometric method, is used for the simultaneous estimation of this combination of TDF, LAMI, and EFV in their combined tablet dosage form.^[35] The purpose of this study was to develop a simple, rapid, precise, and accurate RP-HPLC method for the simultaneous estimation of these drugs in combined tablet dosage form.

MATERIALS AND METHODS

Instrumentation

Liquid chromatographic Shimadzu (LC-2010 $C_{\rm HT}$) system was manufactured by Shimadzu, Kyoto, Japan, and is equipped with auto-sampler, UV and Photodiode Array (PDA) detector, and Rheodyne injector with 20 μ l loop volume. Weighing was done on a Digital Micro Balance an Acculab ALC 210.4 analytical balance, and pH of buffer was maintained by pH analyzer, Chemiline CL 180 μ c based pH meter.

Chemicals and reagents

Reference standards of TDF, LAMI, and EFV were obtained from Cipla Pharmaceuticals (Mumbai, India) and as a gift sample, whereas their combined tablet was obtained from local market. HPLC grade methanol, water (Finar Chemicals Pvt. Ltd., Ahmadabad, India), and HPLC grade orthophosphoric acid (80%) (Finar Chemicals Pvt. Ltd.) were also procured.

Chromatographic conditions

The mobile phase consisted of methanol: Phosphate buffer (sodium dihydrogen orthophosphate, 10 mMol, pH 5.0) in the ratio of 70:30 (v/v) at a flow rate of 1.0 ml/min. Kromasil C_{18} column (150 mm \times 4.6 mm i.d., 5 μ m) was used as the stationary phase. By considering the chromatographic parameter, sensitivity, and selectivity of the method for each of three drugs, 254 nm was selected as the detection wavelength for UV-PDA detector. The HPLC system was operated at a room temperature of 40°C.

Preparation of standard solution

Standard stock solution

Standard stock solutions were prepared by dissolving separately 10 mg of LAMI and TDF and 20 mg of EFV in 100 ml volumetric flask. Dissolve and dilute with methanol up to the mark to get concentrations of 100 μ g/ml of each of LAMI and TDF and 200 μ g/ml of EFV.

Working standard solution

Working standard solutions were prepared by taking 0.1, 0.2, 0.3, 0.4, 0.5, and 0.6 ml into 10 ml volumetric flask and diluted up to the mark with the mobile phase to get 1–6 μ g/ml each for LAMI and TDF and 2–12 μ g/ml for EFV.

Preparation of sample solution

Twenty tablets of combined dosage form of LAMI, TDF, and EFV were weighed and ground to a fine powder. Take powder equivalent to 10 mg of LAMI, TDF, and 20 mg of EFV, mixed, and transferred to a 100-ml volumetric flask. The solution was sonicated to dissolve the powder in 60 ml methanol and diluted up to the mark with the same. The solution was filtered through a Whatman filter paper no. 41. Suitable dilutions with the diluent were made to prepare tablet solutions containing 3 μ g/ml of each of LAMI and TDF and 6 μ g/ml of EFV and then analyzed.

RESULTS AND DISCUSSION

Optimization of chromatographic condition

It was observed from the UV spectra that all the three drugs have considerable absorbances at 254 nm wavelength. So, 254 nm was selected as the detection wavelength. Various combinations of methanol, acetonitrile, and buffers of different pH were tried initially to separate LAMI, TDF, and EFV on C₁₈ column. Preliminary experiments indicated that use of different combinations of acetonitrile or methanol with water was not able to separate the peaks of LAMI, TDF, and EFV and to obtain suitable retention times and peak symmetry. In order to achieve acceptable peak symmetry and separation with good resolution, various buffer systems were tried systematically. Finally, a mobile phase consisting of methanol and phosphate buffer of pH 5.0 (adjusted with 10% solution of orthophosphoric acid) in a ratio of 70:30 v/v and a Kromasil C_{18} column (150 mm × 4.6 mm i.d., 5 μm particle size) were selected to achieve good resolution and acceptable peak symmetry. Flow rates between 0.5-and 1.2 ml/min were tried. Flow rate of 1.0 ml/min was observed to be enough to get both the drugs eluted within less than 10 min. The column temperature was set at 40°C.

System suitability

The retention times for LAMI, TDF, and EFV using optimum conditions were 2.76, 3.96, and 10.5 min, respectively. For three of them, the peak symmetries were <1.5 and the theoretical plates' numbers were >2000. These values are within the acceptable range of United state pharmacopoeia definition and the chromatograms obtained under optimized chromatographic conditions. Figure 4 clearly shows the ability of the method to assess the analyte in the presence of other excipients. The results obtained are shown in Table 1.

Method validation^[36]

The developed method for simultaneous estimation of LAMI, TDF, and EFV has been validated in accordance with the ICH guidelines.^[11]

Linearity

Linearity was checked by preparing standard solutions at six different concentration levels of each of LAMI, TDF, and EFV, ranging from 1 to 6 μ g/ml for each of LAMI and TDF and from 2 to 12 μ g/ml for EFV. Triplicates of 20- μ l injections were made for each concentration and were chromatographed under the chromatographic conditions mentioned above. Peak areas were plotted against the corresponding concentrations to obtain the

Table 1: System suitability parameters					
Parameter	LAMI	TDF	EFV		
Retention time (min)*	2.76±0.002	3.96±0.004	10.5±0.005		
Number of theoretical plates*	2005±25.0	2056±38.0	5352±45.0		
Tailing factor*	1.2±0.032	1.0±0.021	1.2±0.014		
HETP*	116.23±0.34	77.40±0.42	28.02±0.25		
Resolution*	3.37±0.05	3.51±0.01	3.21±0.01		

^{*}Each value is the mean±SD of six determinations

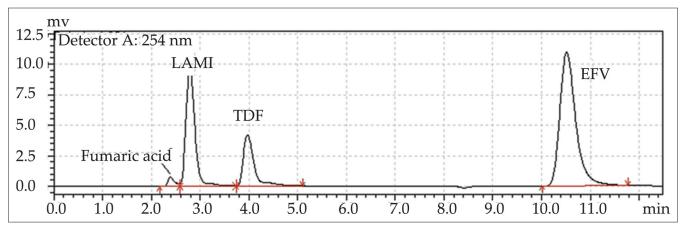


Figure 4: Chromatograms of LAMI (3 μg/ml), TDF (3 μg/ml), and EFV (6 μg/ml) reference substances

calibration graph for each compound. The regression analysis data are given in Table 2.

Sensitivity

The sensitivity of the analytical method was evaluated by determining the limits of detection (LOD) and quantitation (LOQ). The values of LOD and LOQ for LAMI, TDF, and EFV are given in Table 2.

Accuracy

The accuracy of the method for assay determination was checked at three concentration levels of 80%, 100%, and 120% for LAMI, TDF, and EFV. The percentage recoveries are tabulated in Table 3. The recovery was calculated from the slope and intercept of the calibration curve of each drug. As per the ICH guideline, the % recovery must be between 98% and 102%.

Precision

Repeatability: System repeatability was determined by replicate applications and measurements of peak area for LAMI, TDF, and EFV. One dilution in six replicates was analyzed on the same day for repeatability and results were found within acceptable limits (RSD <2) as shown in Table 4.

Intermediate precision: Intermediate precision was assessed by the assay of sample sets on three different days (inter-day precision). Three dilutions in three replicates were analyzed and results were found within acceptable limits (RSD <2) as shown in Table 4.

Robustness

As per the ICH norms, small, but deliberate variations are obtained by altering the temperature at 38°C and 42°C and changing the flow rate to 0.8 and 1.2 ml/min. The change in the flow rate of mobile phase does not affect the peak area; it may only change the retention time of peak of each drug. Results were found within acceptable limits (RSD <2), which are summarized in Table 5.

Stability of sample solution

The sample solution stability was analyzed by injecting the same solution at 0, 6, 12, and 24 h. Identical change was not observed in the developed method. Also, results were found within acceptable limits (RSD <2), which are summarized in Table 6.

Specificity and selectivity

The specificity test of the proposed method demonstrated that the excipients from sample do

Table 2: Linearity data					
Parameter	LAMI	TDF	EFV		
Linearity range (μg/ml)	1–6	1–6	2–12		
Regression	y = 36827x	y = 23574x -	y =45385x –		
equation	+20194	3072	4786.2		
Correlation coefficient (r ²)	0.9991	0.9992	0.9995		
LOD (μg/ml)	0.018	0.01	0.02		
LOQ (μg/ml)	0.05	0.1	0.07		

Table 3: Repeatability precision					
Type of precision LAMI	% RSD				
	TDF	EFV			
Repeatability	0.03	0.013	0.005263		
Intermediate precision					
Intra-day	0.64	0.92	1.0		
Inter-day	0.61-1.04	1.24-1.47	0.90-1.64		

Table 4: Result of recovery studies with static evaluation						
Drug	Amount taken (μg/ml)	Amount added (µg/ml)	Amount recovered (μg/ml)	%Recovery ± SD	%RSD	
LAMI	2.0	0				
	2.0	1.0	0.995	99.46±0.09	0.09	
	2.0	2.0	1.999	99.97±0.05	0.15	
	2.0	3.0	3.041	101.36±0.05	0.18	
TDF	2.0	0				
	2.0	1.0	1.014	101.42±0.08	0.17	
	2.0	2.0	2.018	100.89±0.07	0.25	
	2.0	3.0	2.987	99.57±0.09	0.31	
EFV	4.0	0				
	4.0	2.0	1.999	99.96±0.03	0.11	
	4.0	4.0	4.035	100.87±0.06	0.14	
	4.0	6.0	5.987	99.78±0.06	0.28	

Table 5: Robustness (column temperature related changes and mobile phase flow rate changes, concentration: 3 μ g/ml for LAMI and TDF each and 6 μ g/ml for EFV)

Type of precision	% RSD		
	LAMI	TDF	EFV
Column temperature (±2°C)			
38.0	0.30	0.40	0.11
40.0			
42.0			
Flow rate of mobile phase (±0.2 ml/min)			
0.8	0.20	0.18	0.17
1.0			
1.2			

not interfere with the drug peak. Figure 5 shows the complete separation of LAMI, TDF, and EFV in the presence of tablet excipients.

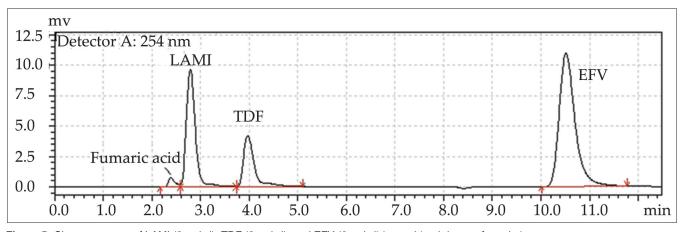


Figure 5: Chromatograms of LAMI (3 µg/ml), TDF (3 µg/ml), and EFV (6 µg/ml) in combined dosage formulation

Time	Assay (%)			%Difference		
	LAMI (3 µg/ml)	TDF (3 µg/ml)	EFV (6 μg/ml)	LAMI (3 µg/ml)	TDF (3 µg/ml)	EFV (6 μg/ml)
Initial	100.12	99.98	100.60			
After 6 h	100.05	99.86	100.54	0.08	0.12	0.12
After 12 h	99.96	99.66	100.17	0.16	0.32	0.49
After 18 h	99.92	99.30	100.11	0.21	0.68	0.55
After 24 h	99.73	99.14	100.02	0.39	0.84	0.64

Assay

The % assay was found to be 100.13% for LAMI, 99.95% for TDF, and 101.14% for EFV.

CONCLUSION

A simple, precise, reliable, sensitive, and accurate RP-HPLC method has been developed for the simultaneous determination of TDF, LAMI, and EFV. The developed method is suitable for the quantification of TDF, LAMI, and EFV in combined tablet dosage form.

REFERENCES

- Budawari S. The Merck Index. Whitehouse Station, NJ: Merck and Co. Inc.; 2006. p. 927, 1573.
- Martindale The complete drug reference. London: Pharmaceutical Press; 2005. p. 635-42.
- Rang HP, Dale MM, Pitter JM, Moore PK. In Pharmacology. New Delhi: Elesvier; 2005. p. 657-63.
- Indian Pharmacopeia. Vol. 2, 3. Government of India. New Delhi: The Controller of Publications; 2010. p. 372-374, 646-650, 1552-1564, 1780-4.
- British Pharmacopoeia, the Stationary Office. Vol. 1 and 2. UK: British Pharmacopoeia Commission; 2009. p. 3407.
- United States of Pharmacopoeia. Vol. 2. United States Pharmacopoeia Convention: Rockville, MD, USA; 2009. p. 2747.
- Central Drug Standard Control Organization. India: New approved drugs in India, Inc.; c2008-09. Available from: http://www.cdsco.nic. in/listofdrugapprovedmain.html. [Last updated on 2009 Jan 1; cited on 2011 Oct 9].
- U.S. Food and Drug Administration. United States: New approved drugs, Inc.; c2007-08. Available from: http://www.aegis.org/ news/fda/2009/FD090901.html [Last updated on 2008 Sep 4; cited 2011 Oct 9].

- Onah J, Ajima U. Quantitative Analysis of Tenofovir by Titrimetric, Extractive Ion pair Spectrophotometric and Charge-Transfer Complexation Methods. Trop J Pharm Res 2011;10:89-96.
- Nevase P, Nimje H, Oswal R, Antre R, Kshirsagar S. UV spectrophotometric method for estimation of Tenofovir disoproxil fumarate. Int J Pharma Res Dev 2011;3:73-5.
- Bapatla J, Sai N, Hari HD, Theja K, Ramalingam P, Reddy Y. Validated HPTLC Method for the Determination of Tenofovir as Bulk Drug and in Pharmaceutical Dosage Form. Pelagia Res Library 2011;2:163-8.
- Kandagal P, Manjunatha D, Seetharamappa J. RP-HPLC Method for the Determination of Tenofovir in Pharmaceutical Formulations and Spiked Human Plasma. Ana Lett 2008;41:561-70.
- Sentenac S, Fernandez C, Thuillier A, Lechat P, Aymard G. Sensitive determination of tenofovir in human plasma samples using reversed-phase liquid chromatography. J Chromatography B Analyt Trchnol Biomed Life Sci 2003;793:317-24.
- Malipatil S, Nandedher M. Determination of Tenofovir Disoproxil Fumarate by a Sensitive Simple Isocratic RP-HPLC Method. J Indian Counc Chem 2009;26:67-9.
- Sudha T, Saminathan J, Hemalatha P, Ravikumar V. Simultaneous Ultraviolate Specrophotometric Estimation of Emtricitabine and Tenofovir disopoxil fumarate in Bulk and Tablet Dosage form. Int J Biopharm 2010;1:26-30.
- Rao J, Gondkar S, Yadav S. Simultaneous HPTLC-Densitometric analysis of Tenofovir and Emtricitabine in Tablet dosage form. Int J Pharm Tech Res 2011;3:1430-4.
- Sharma R, Gupta P. A Validated RP HPLC method for simulataneous estimation of emtricitabine and tenofovir disoproxil fumarate in a tablet dosage Form. EJAC 2009;4:276-8.
- Delahunty T, Bushman L, Robbins B, Fletcher CV. The Simultaneous Assay of Tenofovir and Emtricitabine in Plasma using LC/MS/MS and Isotopically Labeled Internal Standards. J Chromatogr B Analyt Techno Biomed Life Sci 2009;877:20-1.
- Basavaiah K, Somashekar B. Spectrophotometric determination of lamivudine in pure and tablet forms. J Sci Ind Res 2006;65:349-54.
- Deepali G, Elvis M. UV spectrophotometric method for assay of the Antiretroviral drug Lamivudine in active pharmaceutical ingredient and its tablet formulation. J Young Pharm 2010;2:417-9.

- Kaul N, Agrawal H, Paradkar A, Mahadik K. The International Conference on Harmonisation Guidance in Practice: Stress Degradation Studies on Lamivudine and Development of a Validated Specific Stability-Indicating HPTLC Assay Method. Chromatographia 2009;60:213-21.
- Jayaseelan S, Suresh S, Sathishkumar G, Sekar V. Bioanalytical method development and validation of Lamivudine by RP-HPLC method. Int J Chem Tech Res 2010;2:163-7.
- Vijay Kumar B, Vidyadhari CH, Sravan kumar G, Tanuja T. Development and Validation of RP-HPLC method for determination of Lamivudine from pharmaceutical preparation International. J Pharm Ther 2011;2:37-42.
- Nagulwar V, Bhusari K. A validated UV spectrophotometric method for the simultaneous estimation of Lamivudine, Nevirapine and zidovudine in combined tablet dosage form. J Pharm Res 2009;2:666-9.
- Habte G. Simultaneous determination of lamivudine and zidovudine in pharmaceutical formulations by high performance thin layer chromatography (hptlc) densitometric method. Department of pharmaceutical chemistry. Addis Ababa, Ethiopia: Addis Ababa university: 2008.
- Balasekarreddy C, Awen B, Sreekanth N, Ramalingam P. Validated HPLC Method for determination of Lamivudine and Stavudine in their formulations. Pharmanest 2010;1:22-8.
- Samee W, Srilamai P, Ongart S, Suwannaratana R. Simultaneous Determination of Lamivudine, Stavudine and Nevirapine in the Presence of Their Acid-Induced Degradation Products by HPLC. Thai Pharma Health Sci J 2007;2:39-45.
- Kenney K, Wring S, Carr R, Wells G, Dunn J. Simultaneous determination of zidovudine and lamivudine in human serum using HPLC with tandem mass spectrometry. J Pharm Biomed Anal 2000;22:967-83.
- 29. Kumar AY, Rao RN. Development of Rapid UV Spectrophotometric

- Method for the Estimation of Efavirenz in Formulations. E J Chemist 2010:7:856-60
- Hamrapurkar P, PhalecM, Shah N. Quantitative estimation of Efavirenz by high performance thin layer chromatography. Pharmanalysis 2009;1:359-63.
- Potale L, Khodke A, Patolen S, Damle M. Development and validation of Stability indicating HPTLC method for determination of Efavirenz as bulk drug and in pharmaceutical formulation. J Adv Pharm Res 2010;1:115-22.
- Lakshmi Sailaja A, Kishore Kumar K, Ravi Kumar D, Mohan Kumar C. Development and validation of a liquid chromatographic method for determination of efavirenz in human plasma. Chromatogaphia 2010;65:359-61.
- Satyavathi G, Sidhakar M, Rajesh V, Satyanarayana K, Rama Krishna D, Prathap G. Validation of an isocratic HPLC assay of Efavirenz in pharmaceutical formulations of drug substance. Int J Res Rev Pharm App Sci 2011;1:20-6.
- Raju N, Begum S. Simultaneous RP-HPLC Method for the Estimation of the Emtricitabine, Tenofovir Disoproxil Fumerate and Efavirenz in Tablet Dosage Forms. Res J Pharm Tech 2008;1:522-5.
- Sharma R, Mehta K. Simultaneous Spectrophotometric estimation of tenofovir disoproxil fumarate and lamivudine in three component tablet formulation containing efavirenz. Indian J Pharm Sci 2010:72:527-30.
- ICH Guideline Q2 (R1), Validation of Analytical Procedures: Text and Methodology. Geneva: ICH; 2005.

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