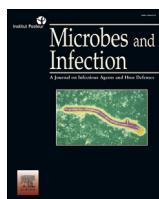




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Review

From hidden outbreaks to epidemic emergencies: the threat associated with neglecting emerging pathogens



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ABSTRACT

Not all infectious disease outbreaks undergo full epidemiological investigations. In certain situations, the resultant lack of knowledge has led to the development of epidemics and public health emergencies. This review will examine six emerging pathogens including their history, present status, and potential to expand to epidemics. Recommendations to improve our understanding of these hidden outbreaks and others also will be provided in the context of health systems policy.

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1. Introduction

Infectious disease outbreaks are common worldwide. While every occurrence deserves a proper epidemiological investigation, several constraints such as limited resources, political considerations, and an assumed familiarity with the pathogen may hinder the process [1,2]. This may result in what are known as "hidden outbreaks" in which spread is known to transpire in a localized environment (such as an endemic pathogen) but inquiries are not considered to be worthwhile in the larger context of global human health. This practice may be considered sound, however there is the potential an outbreak may expand to become an epidemic such that both caseload and costs are significantly increased [3,4].

The hidden outbreak begins no differently than isolated outbreaks with a single point-source such as a household, hospital, or restaurant. While several to dozens of people may be infected, the overall impact on society is considered to be low. Yet, as seen in several instances over the last decade, certain pathogens have become international juggernauts (Fig. 1). Public health authorities are caught off guard, calls for alarm are made by the scientific and medical communities, and the media must find a balance between objective reporting and the inevitable contagion of fear [5].

The quintessential example of such expansion is the Ebola virus epidemic in 2014 [6]. A small outbreak occurred in the small village of Meliandou in Guinea at the end of the previous year and eventually turned into a crisis in three West African nations and led to domestic cases in Europe and the United States. The spread was so troubling, the World Health Organization declared a Public Health

Emergency of International Concern (PHEIC) [7]. The event eventually was controlled yet the virus was considered to be too volatile to ignore any further. Any sign of Ebola in the human population regardless of population size was deemed serious enough to be met with a significant response [8] to control spread.

In addition to understanding the biological nature of a pathogen as the etiology of an outbreak, an examination of the anthropogenic effects facilitating amplification to epidemic status is needed. These factors have been reviewed elsewhere [9] and include the grouping of susceptibles in both the healthcare environment as well as the community, changes in human consumption of natural habitats, territories, and food sources, increased amount and ease of international travel, globalized trade, and political strife. In the context of expanding hidden outbreaks, two factors are considered the most important, travel and trade [10,11].

One of the best known examples of travel-related spread of a hidden outbreak into an epidemic is the SARS coronavirus [12]. The outbreak began in the small village of Foshan in China and expanded to various areas of the country as well as Hong Kong, Singapore, Vietnam, and Canada. Although a lack of proper containment of the virus at the source may have facilitated the spread of the virus into the community [13], travel was deemed as the most influential anthropogenic factor in the development of this epidemic [14].

Another travel-related expansion began in the same year as SARS in the province of Quebec, Canada. Known as the toxinotype III, North American PFGE type 1, and PCR-ribotype 027 (NAP1/027) strain of *Clostridium difficile* [15], the bacterium was found to produce up to 20 times the amount of toxin of other known subtypes and led to over 1,000 deaths [16]. This should have sounded alarms, yet the emergency was considered to be localized with no other regions reporting such activity [16]. Eventually, the outbreak was

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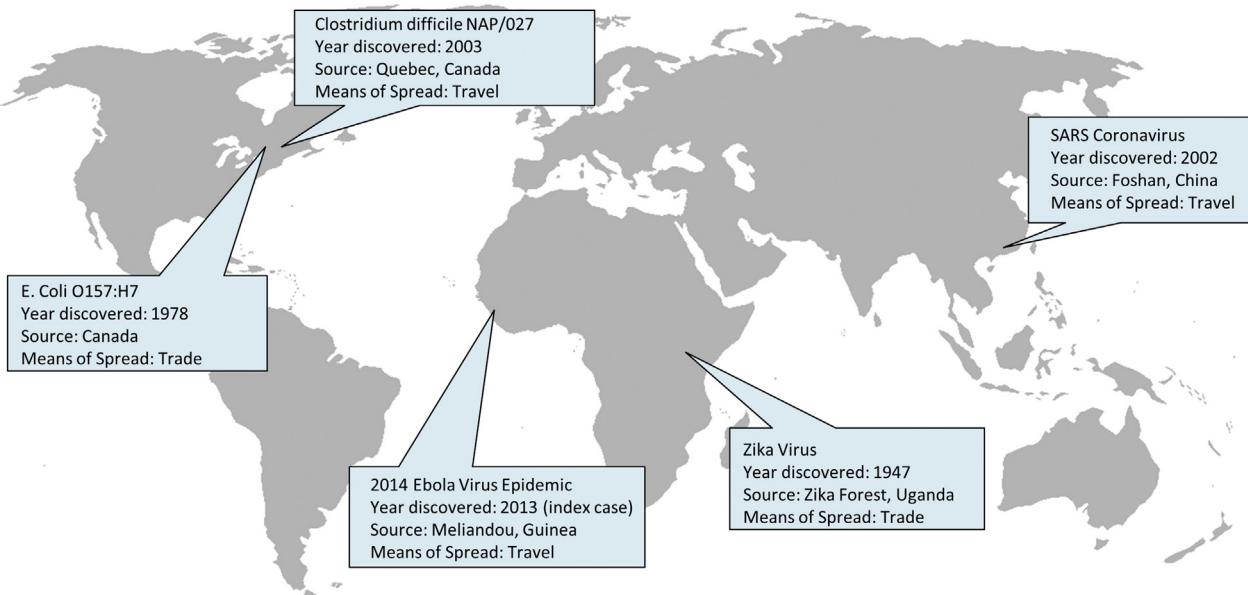


Fig. 1. Hidden Outbreak Source of Recent Global Epidemics. Source: Author edited copy of image, "The World Without Borders," Wikipedia Commons.

found to be part of a larger, unrecognized community-based spread of the pathogen. By 2010, NAP1/027 was found to be prevalent worldwide [17].

Travel of animals through trade has also proven to be a contributing factor. *Escherichia coli* O157:H7 was originally identified as a human pathogen in 1978 [18] but for many years thought to be a sporadic illness [19–22]. However, the strain spread throughout Canada and the United States and over time into other parts of the world due in part to travel as well as livestock movement through commercial routes [23,24].

Trade in general also can lead to inadvertent spread of a pathogen via arthropod vectors such as mosquitoes. This route has been seen as the means for the expansion of the Zika virus [25,26], which was originally found in Uganda in 1947. Due to international trade [25], the virus found its way to the Pacific and eventually the Americas leading to the now infamous epidemic in Brazil. While the exact vehicle has not been identified, a possible route may have involved the importation of mosquitoes in high enough numbers to meet the minimum requirements for establishment in the new environment [27].

Due to lessons learned, increased surveillance for these infectious agents and other well-known species, such as avian influenza viruses [28], polioviruses [29], and measles [30] are now in place. Yet, several pathogens given little attention have begun to show signs of expansion from hidden outbreaks into large scale epidemics. Although none currently poses a significant global threat, increased vigilance is needed to ensure history does not repeat itself. This review will examine six hidden outbreaks that have for the most part eluded widespread attention and will explore their potential to form epidemics. These analyses will include a list of questions that require answering to gain a better understanding of the potential of each pathogen to expand into significant pathogens of concern.

2. Current hidden outbreaks

2.1. *Candida auris*

In 2009, Satoh et al. [31] discovered a strain of yeast in the ear canal of a Japanese individual. Genetic analysis revealed the isolate

was a new species which was named *C. auris*. The species was closely associated with *C. ruelliae* and *C. haemulonii*, the latter of which had been known to cause fungemia [32]. *C. auris* was later isolated in other South Korean patients with otitis media [33]. The discovery was concerning yet assessing the significance in terms of risk for an epidemic was difficult at best. Two years later, the first cases of *C. auris* fungemia in South Korea [34] were detected, including one from an isolate taken 15 years earlier in 1996. The findings suggested the species already could possibly be endemic in the country. Moreover, an increased tolerance to fluconazole was identified suggesting future cases could be more difficult to control. Over the next three years, antimicrobial resistant strains of *C. auris* healthcare-acquired infections were observed in Kuwait [35], India [36], and South Africa [37]. In 2017, the yeast was discovered in the United Kingdom [38], South America [39], Europe [36] and Canada [40]. Genetic rDNA analysis of 24 isolates in the UK demonstrated several geographic origins [38] highlighting the importance of human travel in the spread of the species.

To date, *C. auris* remains for the most part a health-care associated infection affecting immunocompromised individuals. While there is increasing attention in the healthcare field to the risk of this pathogen [41], the international surveillance program SENTRY has demonstrated the contribution of *C. auris* to the overall burden of *Candida* species remains small and limited to healthcare facilities. However, the nearly simultaneous emergence of the species across several continents suggests the yeast is ubiquitous in the environment and may pose a threat to an increasingly immunocompromised population [42]. Additionally, the increase in antimicrobial resistance seen in numerous isolates confers significant hurdles to combat fungemia [43]. Guidelines to deal with *C. auris* are either in place [44] or in development [45] but as seen in the case of *C. difficile*, this may only have a limited effect in controlling its spread in the community and through travel. Public surveillance is necessary to determine the as of yet unknown prevalence of the yeast in the community and potential routes of transmission should also be examined to determine whether preventative strategies such as hygiene measures in the community may help to reduce the burden.

2.2. Coccidioides

Coccidioidomycosis has been a recognized disease since 1892 [46] although it is better known as Valley Fever [47] due to the geographic location of the first American cases, the San Joaquin Valley in California. The disease is caused by the fungal pathogen *Coccidioides immitis* [48] and is marked by skin lesions and the potentially fatal formation of granulomas in numerous organs. At first, the pathogen was considered to be limited to arid desert regions in the United States and Argentina [49], however, wider surveillance has detected the fungus in other geographic regions of the United States including Arkansas [50], Utah [51], Arizona [52], Texas [53] as well as in Canada [54,55], Central and South America [56]. In addition, a retrospective analysis of coccidioidomycosis in China revealed 38 cases involving no history or travel to endemic areas [57] suggesting the fungus already may be spreading globally with no defined routes identified.

Although the mortality of this pathogen is quite low at less than one case per million person-years [58], its expansion over the last half century highlights the potential for an increase in prevalence not only in the Americas but also Asia. Should the species become endemic in China, there is a greater potential for a rapid increase in outbreaks, and possible epidemics as evidenced by SARS. Moreover, the ability of the fungus to infect animals, such as dogs, cats, bats, rodents, and armadillos [48], potentiates even greater spread via domesticated and possibly feral reservoirs. Combined with a noted increase in planetary temperature and resultant increases in desertification [58], there is a need to increase surveillance and explore the modes of transmission including environmental dissemination of dust through wind currents [59].

2.3. HTLV-1

Human T-Lymphotropic Virus was first isolated in 1978 [60] from skin cancer and was considered an atypical oncovirus. Other diseases were eventually associated with this virus including adult T-cell leukemia, HTLV-1 associated myelopathy, tropical spastic paraparesis, uveitis, dermatitis, psychological disorders, and general immunosuppression [61]. Though the virus was detected worldwide, until recently, it was considered sporadic. That view has changed due to epidemics in several regions including Brazil, Spain, Sub-Saharan West Africa, the Middle East, the Caribbean, Japan, and Australia [62]. Due to the bloodborne nature of the virus, which can only be transmitted efficiently through unprotected sexual contact [63–65], the virus typically remains endemic to a region without significant geographic spread. However, historical analyses of HTLV-1 reveal the virus can be introduced into another area through large scale population movements from endemic regions [66,67]. For example, Trevino et al. [68] examined HTLV in Spain and discovered HTLV-1 has entered the country as a result of migration. While the virus has yet to become endemic in the Spanish population, the authors suggest this will occur in time.

Due to the continued effect of mass migrations from endemic areas into naïve ones, the risk for an increase in the prevalence of HTLV-1 exists. Surveillance for this virus needs to be recognized as a priority in countries receiving these individuals. While halting the spread of the virus requires a focus on safe sexual practices, there is a need to identify possible warning signs such as those in Spain prior to expansion of the virus.

2.4. Mycobacterium ulcerans

Discovered in 1948 as an atypical mycobacterium species, *M. ulcerans* has been widely seen as a rare infectious agent in comparison to other species such as *M. tuberculosis* and *M. leprae*.

According to Käser et al. [69], two different lineages of the bacterium exist with one being far more virulent than the other. The virulent strain is known to cause painful skin lesions known as Buruli ulcers, named after the Ugandan region where this infection was first described [70]. The bacterium and associated disease has subsequently been found in several other areas of Africa including Angola [71], Benin [72], the Democratic Republic of the Congo [71], Côte D'Ivoire [73], Ghana [74], Nigeria [75], and Togo [76]. The infection also has been identified in Australia [77], and most recently Jordan [78]. While the association between the bacterium and the symptoms are well established, the mode of transmission has yet to be elucidated. The infection is self-limiting and several reports suggest the highest risk is associated with living in a riverine region [79–83]. As to the potential route for inoculation, several theories have been suggested such as bites from water bugs [84,85] and the potential for colonization with the bacterium through contact with soil [86] and watersheds [87,88].

Without knowing the exact route of transmission, determining the risk of spread from one environment to another is difficult at best. In a genetic analysis of the spread of the pathogenic branch in Africa, Vandelannoote et al. [89] suggested the bacterium spread at the same time as populations were divided by European colonial rule. Infected individuals contaminated pristine water sources allowing for the growth and dissemination of the species. In this regard, the potential for spread of the bacterium relies on migration of those who are infected. While the morbidity associated with this disease should prevent such movements by those infected, political and socioeconomic strife may lead to forced travel during infection. Surveillance for ulcers should be conducted in healthcare facilities to identify new cases in non-endemic regions. In the event a case is discovered, efforts to minimize the potential spread to watersheds need to be in place to prevent the development of endemicity.

2.5. Invasive Streptococcus pyogenes

S. pyogenes has been a significant pathogen for centuries in the form of Scarlet Fever [90]. However, the bacterium has the ability to invade systemically causing bacteremia, endocarditis, toxic shock syndrome, necrotizing fasciitis and endometritis [91,92]. Several contributing factors are associated with these complications [93,94] including fibronectin binding proteins to facilitate invasion, cysteine proteases to escape immune attack, and superantigens that trigger a massive immune response leading to the potential for tissue damage and organ failure.

For the last two decades, a significant increase in the number of invasive infections has been seen in certain regions of North America and Europe. Gheradi et al. [95] have shown several serotypes have increased their circulation in these areas. While between 50 and 70% have been associated with one particular serotype, *emm1*, the risk of complications associated with the other serotypes, including known invasive members such as *emm3*, *emm12* and *emm28* [96] as well as *emm59* [97] and *emm89* [98].

The inability to identify a single serogroup to explain the rise in cases suggests the risk factors associated with the rise of this infection lies in the nature of the susceptible population. Nelson et al. [96] examined 9557 cases of invasive Group A *Streptococcus* infection and found several human factors associated with invasion and mortality including early childhood, advanced age, underlying chronic illness, and immunosuppression.

Given the population-based obstacles regarding prevention of invasive infection, emphasis needs to be placed on surveillance of all *S. pyogenes* infections. While this does occur in many regions of the developed world, more needs to be done to prepare for the inevitable introduction of an invasive serotype into a community. Although this likely will not prevent illnesses and small outbreaks,

public health authorities will have sufficient information to warn the public of a new invader and emphasize hygiene guidelines aimed at prevention of spread.

2.6. Yellow fever

Yellow Fever has been known for over four centuries as a serious illness with the potential to cause death [99]. The virus is present in both Africa and South America although limited to a few regions on these continents. However, due to the ubiquity of its mosquito vectors, *Aedes* and *Haemagogus* species, there is potential for escape from these regions.

Shearer et al. [100] performed a modeling analysis of this virus and determined numerous areas of the world including Southeast Asia, and Central America may be receptive to the virus. In addition, the virus may be transported to non-traditional regions such as the southern regions of China and the United States. In another example, Ibañez-Justicia et al. [101] reported on the introduction of *Aedes aegypti* into a Netherlands airport. While only six insects were identified, as Saarman et al. [27] point out, the number of mosquitoes required to develop a potentially endemic population may be as little as 25. Moreover, yellow fever can develop an urban transmission cycle in an unvaccinated population [102].

Historically, yellow fever has not caused significant widespread concern due to the usually remote nature of endemicity both in Africa and South America [103] and the availability of a vaccine. However, a recent outbreak in Brazil has raised the concerns for those traveling to the region [104,105]. Hamer et al. [106] describe 10 cases of yellow fever in travelers of whom 4 died. None were vaccinated. Moreover, due to the precedent of Zika virus [107], concerns have been raised regarding the potential of the virus to move north into Central and North America where populations are not vaccinated against the virus. The resultant scare has led to a concern for vaccine supply [108] as there is not enough to cover the entire American population living within zones where *Aedes* may thrive.

The most important tool in determining the risk of yellow fever expansion is surveillance. The identification of mosquitoes in Amsterdam [101] is one example of how proper environmental screening of packages from endemic areas may serve to prevent the introduction and eventual reception of a pathogen. In addition, promotion of vaccination will help to reduce the likelihood of infection in travelers. While human to human transmission has not been demonstrated for this virus, the virus may be brought into a region where competent mosquitoes thrive allowing for the initiation of the sylvatic and/or urban transmission cycles.

3. Future directions

The abundance of infectious disease outbreaks worldwide out-numbers the resources available to perform extensive investigations, mitigate morbidity, and determine prevention strategies. Decisions are not easy to make in light of the potential for misjudging a potential epidemic-causing strain as an isolated or sporadic case. Though these occurrences are rare, as demonstrated by previous epidemic of worldwide concern, the consequences may be drastic.

Moving forward, each outbreak associated with these six pathogens needs to be seen as a potential epidemic and algorithms for management need to be in place. Developing these decision-based policies require the use of advanced methods to improve confidence in results. The use of advanced molecular analysis techniques such as whole genome sequencing may provide a wider perspective on the nature of the outbreak and whether it truly is isolated or is a part of a larger problem. Molecular and syndromic

surveillance in at-risk areas can provide real time assessments and provide predictive information. The latter may be aided through the use of social media, which is currently being examined for its power in identifying and predicting outbreaks [109–112]. Mathematical modeling of anthropogenic effects apart from travel and trade, such as climate change, urbanization, and food supply and demand can enhance the fidelity and accuracy of these calculations. Finally, the appreciation of social and cultural practices can offer valuable insight into how healthcare workers and the community may react in the event of pathogen detection. This information also can offer best practices for collaborative efforts to reduce the chances for amplification and spread to wider areas.

In the case of the six pathogens listed in this review, there has yet to be an expansion of cases to epidemic status. However, this situation quickly can change and lead to an expansion into a larger public space. While we may be able to vaguely predict how such escapes may occur, they are only guesses with little contributory value to policy and health systems development. For this to occur, appropriate algorithms need to be put in place and used to assess each outbreak as it happens. This may require valuable resources and could be met with resistance from numerous levels of government and the public in general. However, in light of the economic burden already seen with what amount to preventable epidemics, the cost may be fully justified.

Conflict of interest

None.

References

- [1] Bottcher L, Woolley-Meza O, Araujo NA, Herrmann HJ, Helbing D. Disease-induced resource constraints can trigger explosive epidemics. *Sci Rep* 2015;5:16571.
- [2] van den Wijngaard K. Is this an outbreak? A retrospective evaluation of syndromic surveillance for emerging infectious disease. Erasmus University Rotterdam; 2010.
- [3] Marinović AB, Swaan C, van Steenbergen J, Kretzschmar M. Quantifying reporting timeliness to improve outbreak control. *Emerg Infect Dis* 2015;21:209–16.
- [4] Carpenter TE, O'Brien JM, Hagerman AD, McCarl BA. Epidemic and economic impacts of delayed detection of foot-and-mouth disease: a case study of a simulated outbreak in California. *J Vet Diagn Invest* 2011;23:26–33.
- [5] Towers S, Afzal S, Bernal G, Bliss N, Brown S, Espinoza B, et al. Mass media and the contagion of fear: the case of Ebola in America. *PLoS One* 2015;10:e0129179.
- [6] Agua-Agum J, Allegranzi B, Ariyarajah A, Aylward R, Blake IM, Barboza P, et al. After Ebola in West Africa – unpredictable risks, preventable epidemics. *N Engl J Med* 2016;375:587–96.
- [7] Mackey TK. The Ebola outbreak: catalyzing a "shift" in global health governance? *BMC Infect Dis* 2016;16:699.
- [8] Houlihan CF, Youree D, Brown CS. Novel surveillance methods for the control of Ebola virus disease. *Int Health* 2017;9:139–41.
- [9] Sattar SA, Tetro J, Springthorpe VS. Impact of changing societal trends on the spread of infections in American and Canadian homes. *Am J Infect Contr* 1999;27:S4–21.
- [10] Meloni S, Perra N, Arenas A, Gómez S, Moreno Y, Vespignani A. Modeling human mobility responses to the large-scale spreading of infectious diseases. *Sci Rep* 2011;1:62.
- [11] Saker L, Lee K, Cannito B, Gilmore A, Campbell-Lendrum D. Globalization and infectious diseases: a review of the linkages. Geneva: World Health Organization; 2004.
- [12] Baric RS. SARS-CoV: lessons for global health. *Virus Res* 2008;133:1–3.
- [13] Fidler DP. Germs, governance, and global public health in the wake of SARS. *J Clin Invest* 2004;113:799–804.
- [14] Lam WK, Zhong NS, Tan WC. Overview on SARS in Asia and the world. *Respirology* 2003;8:S2–5.
- [15] Werny M, Pepin J, Fang A, Killgore G, Thompson A, Brazier J, et al. Toxin production by an emerging strain of *Clostridium difficile* associated with outbreaks of severe disease in North America and Europe. *Lancet* 2005;366:1079–84.
- [16] Pepin J, Valiquette L, Cossette B. Mortality attributable to nosocomial *Clostridium difficile*-associated disease during an epidemic caused by a hyper-virulent strain in Quebec. *CMAJ* 2005;173:1037–42.

- [17] Gerding DN. Global epidemiology of *Clostridium difficile* infection in 2010. *Infect Control Hosp Epidemiol* 2010;31:S32–4.
- [18] Hockin JC, Lior H. Hemorrhagic colitis due to *Escherichia coli* O157:H7. A rare disease? *CMAJ* 1986;134:25–6.
- [19] Prevention USCFDCa Isolation of *E. Coli* O157:H7 from sporadic cases of hemorrhagic colitis –United States. *MMWR Morb Mortal Wkly Rep* 1982;31(580):5.
- [20] Johnson WM, Lior H, Bezanson GS. Cytotoxic *Escherichia coli* O157:H7 associated with haemorrhagic colitis in Canada. *Lancet* 1983;1:76.
- [21] Pai CH, Gordon R, Sims HV, Bryan LE. Sporadic cases of hemorrhagic colitis associated with *Escherichia coli* O157:H7. Clinical, epidemiologic, and bacteriologic features. *Ann Intern Med* 1984;101:738–42.
- [22] Wells JG, Davis BR, Wachsmuth IK, Riley LW, Remis RS, Sokolow R, et al. Laboratory investigation of hemorrhagic colitis outbreaks associated with a rare *Escherichia coli* serotype. *J Clin Microbiol* 1983;18:512–20.
- [23] Dallan TJ, Ashton PM, Byrne L, Perry NT, Petrovska L, Ellis R, et al. Applying phylogenomics to understand the emergence of Shiga-toxin-producing *Escherichia coli* O157:H7 strains causing severe human disease in the UK. *Microb Genom* 2015;1:e000029.
- [24] Hodges JR, Kimball AM. The global diet: trade and novel infections. *Glob Health* 2005;1:4.
- [25] Haddow AD, Schuh AJ, Yasuda CY, Kasper MR, Heang V, Huy R, et al. Genetic characterization of Zika virus strains: geographic expansion of the Asian lineage. *PLoS Neglected Trop Dis* 2012;6:e1477.
- [26] Duffy MR, Chen TH, Hancock WT, Powers AM, Kool JL, Lanciotti RS, et al. Zika virus outbreak on Yap Island, Federated States Of Micronesia. *N Engl J Med* 2009;360:2536–43.
- [27] Saarman NP, Gloria-Soria A, Anderson EC, Evans BR, Pless E, Cosme LV, et al. Effective population sizes of a major vector of human diseases, *Aedes aegypti*. *Evol Appl* 2017;10:1031–9.
- [28] Noah DL, Noah JW. Adapting global influenza management strategies to address emerging viruses. *Am J Physiol Lung Cell Mol Physiol* 2013;305: L108–17.
- [29] Gardner TJ, Diop OM, Jorba J, Chavan S, Ahmed J, Anand A. Surveillance to track progress toward polio eradication – worldwide, 2016–2017. *MMWR Morb Mortal Wkly Rep* 2018;67:418–23.
- [30] Xu W, Zhang Y, Wang H, Zhu Z, Mao N, Mulders MN, et al. Global and national laboratory networks support high quality surveillance for measles and rubella. *Int Health* 2017;9:184–9.
- [31] Satoh K, Makimura K, Hasumi Y, Nishiyama Y, Uchida K, Yamaguchi H. *Candida auris* sp. nov., a novel ascomycetous yeast isolated from the external ear canal of an inpatient in a Japanese hospital. *Microbiol Immunol* 2009;53: 41–4.
- [32] Silva CM, Carvalho-Parahym AM, Macedo DP, Lima-Neto RG, Francisco EC, Melo AS, et al. Neonatal candidemia caused by *Candida haemulonii*: case report and review of literature. *Mycopathologia* 2015;180:69–73.
- [33] Kim MN, Shin JH, Sung H, Lee K, Kim EC, Ryoo N, et al. *Candida haemulonii* and closely related species at 5 university hospitals in Korea: identification, antifungal susceptibility, and clinical features. *Clin Infect Dis* 2009;48: e57–61.
- [34] Lee WG, Shin JH, Uh Y, Kang MG, Kim SH, Park KH, et al. First three reported cases of nosocomial fungemia caused by *Candida auris*. *J Clin Microbiol* 2011;49:3139–42.
- [35] Emara M, Ahmad S, Khan Z, Joseph L, Al-Obaid Im, Purohit P, et al. *Candida auris* candidemia in Kuwait, 2014. *Emerg Infect Dis* 2015;21:1091–2.
- [36] Chowdhary A, Sharma C, Meis JF. *Candida auris*: a rapidly emerging cause of hospital-acquired multidrug-resistant fungal infections globally. *PLoS Pathog* 2017;13:e1006290.
- [37] Magobo RE, Corcoran C, Seetharam S, Govender NP. *Candida auris*–associated candidemia, South Africa. *Emerg Infect Dis* 2014;20:1250–2.
- [38] Borman AM, Szekely A, Johnson EM. Comparative pathogenicity of United Kingdom isolates of the emerging pathogen *Candida auris* and other key pathogenic *Candida* species. *mSphere* 2016;1. e00189-16.
- [39] Calvo B, Melo AS, Perozo-Mena A, Hernandez M, Francisco EC, Hagen F, et al. First report of *Candida auris* in America: clinical and microbiological aspects of 18 episodes of candidemia. *J Infect* 2016;73:369–74.
- [40] Schwartz IS, Hammond GW. First reported case of multidrug-resistant *Candida auris* in Canada. *Can Commun Dis Rep* 2017;43:150–3.
- [41] Clancy CJ, Nguyen MH. Emergence of *Candida auris*: an international call to arms. *Clin Infect Dis* 2017;64:141–3.
- [42] Low C-Y, Rotstein C. Emerging fungal infections in immunocompromised patients. *F1000 Med Rep* 2011;3:14.
- [43] Osei Sekyere J. *Candida auris*: a systematic review and meta-analysis of current updates on an emerging multidrug-resistant pathogen. *Microbiologyopen* 2018. <https://doi.org/10.1002/mbo.3578> [Epub ahead of print].
- [44] Tsay S, Kallen A, Jackson BR, Chiller TM, Vallabhaneni S. Approach to the investigation and management of patients with *Candida auris*, an emerging multidrug-resistant yeast. *Clin Infect Dis* 2018;66:306–11.
- [45] Lu PL, Liu WL, Lo Hj, Wang FD, Ko WC, Hsueh PR, et al. Are we ready for the global emergence of multidrug-resistant *Candida auris* in Taiwan? *J Formos Med Assoc* 2018;117:462–70.
- [46] Hirschmann JV. The early history of coccidioidomycosis: 1892–1945. *Clin Infect Dis* 2007;44:1202–7.
- [47] Smith CE. Epidemiology of acute coccidioidomycosis with erythema nodosum (“San Joaquin” or “Valley Fever”). *Am J Public Health Natl Health* 1940;30:600–11.
- [48] Brown J, Benedict K, Park BJ, Thompson GR. Coccidioidomycosis: epidemiology. *Clin Epidemiol* 2013;5:185–97.
- [49] Canteros CE, Toranzo A, Ibarra-Camou B, David V, Carrizo SG, Santillan-Iturres A, et al. La coccidioidomicosis en Argentina, 1892–2009. *Rev Argent Microbiol* 2010;42:261–8.
- [50] Badley JW, Winthrop KL, Patkar NM, Delzell E, Beukelman T, Xie F, et al. Geographic distribution of endemic fungal infections among older persons, United States. *Emerg Infect Dis* 2011;17:1664–9.
- [51] Petersen LR, Marshall SL, Barton-Dickson C, Hajjeh RA, Lindsley MD, Warnock DW, et al. Coccidioidomycosis among workers at an archeological site, northeastern Utah. *Emerg Infect Dis* 2004;10(4):637–42.
- [52] Park BJ, Sigel K, Vaz V, Komatsu K, McRill C, Phelan M, et al. An epidemic of coccidioidomycosis in Arizona associated with climatic changes, 1998–2001. *J Infect Dis* 2005;191:1981–7.
- [53] Teel KW, Yow MD, Williams Jr TW. A localized outbreak of coccidioidomycosis in southern Texas. *J Pediatr* 1970;77:65–73.
- [54] Prasad E, Diedi P, Fernandes D, Hodge L, Ower K, Rennie R. Serological evidence of increased *Coccidioides immitis* infections in western Canada in 1996. *Can J Infect Dis* 1998;9:377–81.
- [55] Nicolle L, Rotstein C, Bourgault A, St-Germain G, Garber G. Invasive fungal infections in Canada from 1992 to 1994. *Can J Infect Dis* 1998;9:347–52.
- [56] Freedman M, Jackson BR, McCotter O, Benedict K. Coccidioidomycosis outbreaks, United States and worldwide, 1940–2015. *Emerg Infect Dis* 2018;24: 417–24.
- [57] Liang G, Shen Y, Lv G, Zheng H, Mei H, Zheng X, et al. Coccidioidomycosis: Imported and possible domestic cases in China: a case report and review, 1958–2017. *O Mycoses* 2018. <https://doi.org/10.1111/myc.12750> [Epub ahead of print].
- [58] Huang JY, Bristow B, Shafir S, Sorvillo F. Coccidioidomycosis-associated deaths, United States, 1990–2008. *Emerg Infect Dis* 2012;18:1723–8.
- [59] Lauer A. Coccidioidomycosis: increasing incidence of an “orphan” disease in response to environmental changes. In: Hurst C, editor. Modeling the transmission and prevention of infectious disease. Advances in environmental microbiology, vol. 4. Cham: Springer; 2017. p. 151–85.
- [60] Gallo RC. The discovery of the first human retrovirus: HTLV-1 and HTLV-2. *Retrovirology* 2005;2:17.
- [61] Gonçalves DU, Projetti FA, Ribas JGR, Araújo MG, Pinheiro SR, Guedes AC, et al. Epidemiology, treatment, and prevention of human T-Cell leukemia virus type 1-associated diseases. *Clin Microbiol Rev* 2010;23:577–89.
- [62] Gessain A, Cassar O. Epidemiological aspects and world distribution of HTLV-1 infection. *Front Microbiol* 2012;3:388.
- [63] Eshima N, Tabata M, Kikuchi H, Karukaya S, Taguchi T. Analysis of the infection system of human T-cell leukaemia virus type I based on a mathematical epidemic model. *Stat Med* 2001;20:3891–900.
- [64] Eshima N, Tabata M, Okada T, Karukaya S. Population dynamics of HTLV-I infection: a discrete-time mathematical epidemic model approach. *Math Med Biol* 2003;20:29–45.
- [65] Eshima N, Tabata M, Okada T. Why is the distribution of HTLV-I carriers geographically biased? An answer through a mathematical epidemic model. *Math Med Biol* 2007;24:149–67.
- [66] Paiva A, Casseb J. Origin and prevalence of human T-lymphotropic virus type 1 (HTLV-1) and type 2 (HTLV-2) among indigenous populations in the Americas. *Rev Inst Med Trop Sao Paulo* 2015;57:1–13.
- [67] Yanagihara R, Saitou N, Nerurkar VR, Song KJ, Bastian I, Franchini G, et al. Molecular phylogeny and dissemination of human T-cell lymphotropic virus type I viewed within the context of primate evolution and human migration. *Cell Mol Biol (Noisy-le-grand)* 1995;41(Suppl 1):S145–61.
- [68] Treviño A, Aguilera A, Caballero E, Benito R, Parra P, Eiros JM, et al. Trends in the prevalence and distribution of HTLV-1 and HTLV-2 infections in Spain. *Virol J* 2012;9:71.
- [69] Kaser M, Rondini S, Naegeli M, Stinear T, Portaels F, Certa U, et al. Evolution of two distinct phylogenetic lineages of the emerging human pathogen *Mycobacterium ulcerans*. *BMC Evol Biol* 2007;7:177.
- [70] Clancey J, Dodge R, Lunn HF. Study of a mycobacterium causing skin ulceration in Uganda. *Ann Soc Belg Med Trop* 1920;1962(42):585–90.
- [71] Kibadi K, Panda M, Tamfum JJ, Fraga AG, Longatto Filho A, Anyo G, et al. New foci of Buruli ulcer, Angola and democratic republic of Congo. *Emerg Infect Dis* 2008;14:1790–2.
- [72] Debacker M, Aguiar J, Steunou C, Zinsou C, Meyers WM, Guedenon A, et al. *Mycobacterium ulcerans* disease (Buruli ulcer) in rural hospital, Southern Benin, 1997–2001. *Emerg Infect Dis* 2004;10:1391–8.
- [73] Tano MB, Dassi C, Mosi L, Koussemon M, Bonfoh B. Molecular characterization of mycolactone producing mycobacteria from aquatic environments in Buruli ulcer non-endemic areas in Côte d'Ivoire. *Int J Environ Res Publ Health* 2017;14:E178Pubmed.
- [74] Aboagye SY, Ampah KA, Ross A, Asare P, Otchere ID, Fyfe J, et al. Seasonal pattern of *Mycobacterium ulcerans*, the causative agent of Buruli ulcer, in the environment in Ghana. *Microb Ecol* 2017;74:350–61.
- [75] Chukwuekezie O, Ampadu E, Sopoh G, Dossou A, Tiendrebeogo A, Sadiq L, et al. Buruli ulcer, Nigeria. *Emerg Infect Dis* 2007;13:782–3.

- [76] Bretzel G, Huber KL, Kobara B, Beissner M, Piten E, Herbiger KH, et al. Laboratory confirmation of Buruli ulcer disease in Togo, 2007–2010. *PLoS Neglected Trop Dis* 2011;5:e1228.
- [77] Tai AYC, Athan E, Friedman ND, Hughes A, Walton A, O'Brien DP. Increased severity and spread of *Mycobacterium ulcerans*, southeastern Australia. *Emerg Infect Dis* 2018;24:58–64.
- [78] Al Ramahi JW, Annab H, Al Karmi M, Kirresh B, Wreikat M, Batarseh R, et al. Chronic cutaneous mycobacterial ulcers due to *Mycobacterium ulcerans* (Buruli ulcer): the first indigenous case report from Jordan and a literature review. *Int J Infect Dis* 2017;58:77–81.
- [79] Duker AA, Portaels F, Hale M. Pathways of *Mycobacterium ulcerans* infection: a review. *Environ Int* 2006;32:567–73.
- [80] Maman I, Tchacondo T, Kere AB, Beissner M, Kobara Y, et al. Molecular detection of *Mycobacterium ulcerans* in the environment and its relationship with Buruli ulcer occurrence in Zio and Yoto districts of maritime region in Togo. *PLoS Neglected Trop Dis* 2018;12. e0006455.
- [81] Maman I, Tchacondo T, Kere AB, Piten E, Beissner M, Kobara Y, et al. Risk factors for *Mycobacterium ulcerans* infection (Buruli Ulcer) in Togo – a case-control study in Zio and Yoto districts of the maritime region. *BMC Infect Dis* 2018;18:48.
- [82] Pileggi SM, Jordan H, Clennon JA, Whitney E, Benbow ME, Merritt R, et al. Landscape and environmental influences on *Mycobacterium ulcerans* distribution among aquatic sites in Ghana. *PLoS One* 2017;12. e0176375.
- [83] Aboagye SY, Asare P, Otchere ID, Koka E, Mensah GE, Yirenya-Tawiah D, et al. Environmental and behavioral drivers of Buruli ulcer disease in selected communities along the Densu River basin of Ghana: a case-control study. *Am J Trop Med Hyg* 2017;96:1076–83.
- [84] Zogo B, Djenontin A, Carolan K, Babonneau J, Guegan JF, Eyangoh S, et al. A field study in Benin to investigate the role of mosquitoes and other flying insects in the ecology of *Mycobacterium ulcerans*. *PLoS Neglected Trop Dis* 2015;9. e0003941.
- [85] Marion E, Eyangoh S, Yeramian E, Doannio J, Landier J, Aubry J, et al. Seasonal and regional dynamics of *M. ulcerans* transmission in environmental context: deciphering the role of water bugs as hosts and vectors. *PLoS Neglected Trop Dis* 2010;4(7):e731.
- [86] O'Brien DP, Wynne JW, Buultjens AH, Michalski WP, Stinear TP, Friedman ND, et al. Exposure risk for infection and lack of human-to-human transmission of *Mycobacterium ulcerans* disease, Australia. *Emerg Infect Dis* 2017;23:837–40.
- [87] Zingue D, Bouam A, Militello M, Drancourt M. High-throughput carbon substrate profiling of *Mycobacterium ulcerans* suggests potential environmental reservoirs. *PLoS Neglected Trop Dis* 2017;11. e0005303. pii: e00045–17.
- [88] Zingue D, Bouam A, Tian RBD, Drancourt M. Buruli ulcer, a prototype for ecosystem-related infection, caused by *Mycobacterium ulcerans*. *Clin Microbiol Rev* 2018;31. pii: e00045–17.
- [89] Vandelannoote K, Meehan CJ, Eddyani M, Affolabi D, Phanza DM, Eyangoh S, et al. Multiple introductions and recent spread of the emerging human pathogen *Mycobacterium ulcerans* across Africa. *Genome Biol Evol* 2017;9: 414–26.
- [90] Rolleston JD. The history of scarlet fever. *Br Med J* 1928;2:926–9.
- [91] Memish ZA, Gravel-Tropper D, Oxley C, Toye B, Garber GE. Group A streptococcal endometritis: report of an outbreak and review of the literature. *Can J Infect Dis* 1994;5:276–81.
- [92] Weiss KA, Laverdière M. Group A *Streptococcus* invasive infections: a review. *Can J Surg* 1997;40:18–25.
- [93] Terao Y. The virulence factors and pathogenic mechanisms of *Streptococcus pyogenes*. *J Oral Biosci* 2012;54:96–100.
- [94] Stevens DL, Bryant AE. Severe Group A streptococcal infections. In: Ferretti JJ, Stevens DL, Fischetti VA, editors. *Streptococcus pyogenes : basic biology to clinical manifestations*. Oklahoma City (OK): University of Oklahoma Health Sciences Center(c) The University of Oklahoma Health Sciences Center; 2016.
- [95] Gherardi G, Vitali LA, Creti R. Prevalent emm types among invasive GAS in Europe and North America since year 2000. *Front Public Health* 2018;6:59.
- [96] Nelson GE, Pondo T, Toews KA, Farley MM, Lindgren ML, Lynfield R, et al. Epidemiology of invasive Group A streptococcal infections in the United States, 2005–2012. *Clin Infect Dis* 2016;63:478–86.
- [97] Tyrrell GJ, Lovgren M, St Jean T, Hoang L, Patrick DM, Horsman G, et al. Epidemic of group A *Streptococcus* Memm59 causing invasive disease in Canada. *Clin Infect Dis* 2010;51:1290–7.
- [98] Turner CE, Abbott J, Lamagni T, Holden MT, David S, Jones MD, et al. Emergence of a new highly successful capsular group a *Streptococcus* clade of genotype emm89 in the United Kingdom. *mBio* 2015;6. e000622.
- [99] Johansson MA, Vasconcelos PFC, Staples JE. The whole iceberg: estimating the incidence of yellow fever virus infection from the number of severe cases. *Trans R Soc Trop Med Hyg* 2014;108:482–7.
- [100] Shearer FM, Longbottom J, Browne AJ, Pigott DM, Brady OJ, Kraemer MUG, et al. Existing and potential infection risk zones of yellow fever worldwide: a modelling analysis. *Lancet Glob Health* 2018;6:e270–8.
- [101] Ibanez-Justicia A, Gloria-Soria A, den Hartog W, Dik M, Jacobs F, Stroo A. The first detected airline introductions of yellow fever mosquitoes (*Aedes aegypti*) to Europe, at Schiphol International airport, The Netherlands. *Parasites Vectors* 2017;10:603.
- [102] Couto-Lima D, Madec Y, Bersot MI, Campos SS, Motta MA, Santos FBD, et al. Potential risk of re-emergence of urban transmission of Yellow Fever virus in Brazil facilitated by competent *Aedes* populations. *Sci Rep* 2017;7:4848.
- [103] Hamrick PN, Aldighieri S, Machado G, Leonel DG, Vilca LM, Uriona S, et al. Geographic patterns and environmental factors associated with human yellow fever presence in the Americas. *PLoS Neglected Trop Dis* 2017;11. e0005897.
- [104] Fujita DM, da Silva Nali LH, Salvador FS, de Andrade Júnior HF. Recommendations for travellers during the yellow fever outbreaks in Brazil—2018. *J Trav Med* 2018;25. tay016.
- [105] TdSS Chaves, Orduna T, Lepetic A, Macchi A, Verbanaz S, Risquez A, et al. Yellow fever in Brazil: epidemiological aspects and implications for travelers. *S1477–8939(18)30091-7 Trav Med Infect Dis* 2018. <https://doi.org/10.1016/j.tmaid.2018.05.001> [Epub ahead of print].
- [106] Hamer DH, Angelo K, Caumes E, van Genderen PJ, Florescu SA, Popescu CP, et al. Fatal yellow fever in travelers to Brazil, 2018. *MMWR Morb Mortal Wkly Rep* 2018;67:340–1.
- [107] Paules CI, Fauci AS. Yellow fever - once again on the radar screen in the americas. *N Engl J Med* 2017;376:1397–9.
- [108] Lucey DR, Donaldson H. Yellow fever vaccine shortages in the United States and abroad: a critical issue. *Ann Intern Med* 2017;167:664–5.
- [109] Alessa A, Faezipour M. A review of influenza detection and prediction through social networking sites. *Theor Biol Med Model* 2018;15:2.
- [110] Tang L, Bie B, Park SE, Zhi D. Social media and outbreaks of emerging infectious diseases: a systematic review of literature. pii: S0196–6553(18)30105–6 Am J Infect Contr 2018. <https://doi.org/10.1016/j.ajic.2018.02.010> [Epub ahead of print].
- [111] Charles-Smith LE, Reynolds TL, Cameron MA, Conway M, Lau EH, Olsen JM, et al. Using social media for actionable disease surveillance and outbreak management: a systematic literature review. *PLoS One* 2015;10. e0139701.
- [112] Santillana M, Nguyen AT, Dredze M, Paul MJ, Nsoesie EO, Brownstein JS. Combining search, social media, and traditional data sources to improve influenza surveillance. *PLoS Comput Biol* 2015;11. e1004513.