

RESEARCH

Open Access

# Sex-specific induction of CYP6 cytochrome P450 genes in cadmium and lead tolerant *Anopheles gambiae*

Fauzia K Musasia<sup>1,2</sup>, Alfred O Isaac<sup>1</sup>, Daniel K Masiga<sup>2</sup>, Irene A Omedo<sup>3</sup>, Ramadhan Mwakubambanya<sup>1</sup>, Richard Ochieng<sup>2</sup> and Paul O Mireji<sup>1,2\*</sup>

## Abstract

**Background:** *Anopheles gambiae*, one of the main Afro-tropical mosquito vector of malaria, has adapted to heavy metals in its natural habitat, and developed resistance to most conventional insecticides. Investigations were conducted to establish an association between tolerance to cadmium or lead-heavy metals, and expression of specific genes for cytochrome p450 enzymes associated with pyrethroid resistance in the mosquito.

**Methods:** Juvenile aquatic stages of the mosquito were selected for tolerance to cadmium or lead through chronic exposure of the stages to maximum acceptable toxicant concentrations (MATCs) of the metals. Using real-time quantitative polymerase chain reaction (qPCR), three replicates each of male or female cadmium or lead-tolerant individuals and relevant controls were separately screened for expression of *CYP6M2*, *CYP6P3* and *CYP6Z1* genes. The variance in expression levels of the genes amongst the treatments was compared by ANOVA statistical tool.

**Results:** Expressions of all the genes were significantly lower ( $P < 0.05$ ) in females than in males. Within gender, there 1.3 - 2.3 or 3.1-4.2-fold reduction in expression of the genes in cadmium or lead selected than respective control populations. Expression of all the classes of gene was elevated in cadmium selected female populations relative to their respective controls.

**Conclusion:** These findings suggest that tolerance to cadmium or lead in the mosquito can influence response in cytochrome p450 genes associated with metabolism of pyrethroids in the mosquito in a sex-specific manner. This can, in turn, affect sensitivity of the mosquito to pyrethroids and other xenobiotics associated with these genes, with potential implications in mosquito vector control operations.

**Keywords:** *Anopheles gambiae*, Cadmium, Lead, Cytochrome p450

## Background

*Anopheles gambiae*, one of the main African vector of malaria, is expanding its ecological niche into urban areas where larvae of the mosquito have often been observed proliferating in polluted aquatic habitats [1,2]. The pollutants include domestic and/or industrial sewage [1,3], and heavy metals in excess of natural loads [2]. The mosquito has also developed resistance to conventional insecticides, including pyrethroids, following

extensive application of the insecticides in control programmes of the vector in rural and urban Africa [4,5]. The adaptation to the metals and resistance to the insecticides has in turn facilitated colonization by the mosquitoes of polluted habitats that are potentially unfavourable to susceptible predators and competitors of the mosquitoes [2,6-9]. Resistance has been facilitated by metabolism of the pyrethroids by *CYP6M2*, *CYP6P3* and *CYP6Z1* genes of CYP6 family of cytochrome p450 enzymes in the mosquito [10-12]. Evolutionary pressures have also driven catalytic plasticity in these enzymes, facilitating rapid expansion and diversification of functions of the genes, to potentially handle additional xenobiotics, including heavy metals [13]. The roles of these genes in

\* Correspondence: mireji@gmail.com

<sup>1</sup>Department of Biochemistry and Molecular Biology, Egerton University, P.O. Box 536, Njoro, Kenya

<sup>2</sup>Molecular Biology and Bioinformatics Unit, International Centre of Insect Physiology and Ecology, P.O. Box 30772-00100, Nairobi, Kenya  
Full list of author information is available at the end of the article

mediating resistance to pyrethroids in *Anopheles gambiae* mosquito tolerant to cadmium, copper or lead heavy metal have not been elucidated.

However, tolerance to heavy metals in the mosquito has been associated with loss in biological fitness of the mosquito with potential implication in their ecological performance [14-16]. Therefore, understanding the influence of heavy metal tolerance on CYP6 gene expression responses in the mosquito can potentially elucidate impact of adaptation of the mosquitoes to the metals in nature and on the susceptibility of the mosquitoes to the pyrethroid insecticides routinely used in malaria control interventions against the vector [17]. In this study, the effect of tolerance in *An. gambiae* s.s. to cadmium and lead-heavy metals on expression of the *CYP6M2*, *CYP6P3* and *CYP6Z1* genes of CYP6 family of cytochrome p450 in the species was assessed.

## Methods

### Selection for tolerance to cadmium or lead in *Anopheles gambiae* s.s.

*Anopheles gambiae* s. s. mosquitoes were obtained from a colony kept by the Animal Rearing and Containment Unit (ARCU) of International Centre of Insect Physiology and Ecology (ICIPE), Nairobi, Kenya. This colony was originally collected from Mbita field station (00.025'S, 34.013'E), Homa Bay County, Kenya in December 2000. Cadmium or lead-heavy, metal-tolerant, and control strains of the mosquito were separately generated and maintained through exposures of successive generations of the mosquito to chronic concentrations of the metals in three independent replicates for each strain and treatment since 2006 as described in [15]. Briefly, larval stages of *An. gambiae* s. s. were selected for tolerance to cadmium or lead through chronic exposures to empirically determined maximum acceptable toxicant concentration (MATCs) of the metals. At the time of this work, the colony had been selected for tolerance to respective heavy metals through at least 143 filial generations. Overall, maintenance of the colony followed standard operating procedure for rearing *Anopheles*, where female mosquitoes were blood-fed on anaesthetized mice, and larvae on pulverized Tetramin fish food (Tetra GmbH, Melle, Germany) [18]. Approval for feeding mosquitoes on mice was obtained from the Kenya National Ethical Review Board (protocol number KEMRI/RES/7/3/1), and the protocol reviewed by the KEMRI IACUC. Cadmium and lead used in this study were applied as cadmium chloride ( $\text{CdCl}_2$ ) 99.99% pure (Fisher Scientific LLC, Fair Lawn, NJ, USA) and lead II nitrate [ $\text{Pb}(\text{NO}_3)_2$ ] 99.5% pure analytical salts (Prolabo, Fontenay-sous-Bois, France).

### Extraction of ribonucleic acid from adult *Anopheles gambiae* s.s.

Total ribonucleic acid (RNA) was separately isolated from pools of the three independent replicates, each pool consisting of five three-day-old male or female adult mosquitoes of cadmium or lead-tolerant strain, or their respective untreated controls. RNA was isolated using TRIzol reagent (Invitrogen, Carlsbad, USA) following the manufacturer's protocol. The resultant RNA from each pool was separately suspended in 12  $\mu\text{L}$  of nuclease-free water and stored at  $-80^\circ\text{C}$  until required. Integrity of extracted RNA was validated by electrophoresis in 0.3% agarose (Sigma-Aldrich hemie, GmbH, Steinheim, Germany) RNA denaturing gel in 1.4% sodium phosphate, with 1 mg/mL ethidium bromide staining for visualization. The yields and quality of RNA were determined spectroscopically [19], and the RNA subsequently used for cDNA synthesis.

### Reverse transcriptions of genes for cytochrome p450

Two-steps reverse transcriptions of RNA, and Dnase treatment to remove potential genomic DNA carryover/contaminants were conducted using RevertAid™ H Minus First Strand Synthesis Kit (Fermentas, Lithuania) following manufacturer's instructions. The DNase treatment reaction consisted of 1  $\mu\text{g}$  RNA extract, 1  $\mu\text{L}$  10X reaction buffer with  $\text{MgCl}_2$ , 1  $\mu\text{L}$  of RNase-free DNase I and nuclease-free water in total reaction volumes of 10  $\mu\text{L}$ . The reactions were incubated at  $37^\circ\text{C}$  for 30 min, followed by the addition of 1  $\mu\text{L}$  50mM EDTA, and a further incubation for 10 min at  $65^\circ\text{C}$ . Resultant RNA from each strain was separately denatured by incubation at  $65^\circ\text{C}$  for 5 min in 12  $\mu\text{L}$  reactions mix consisting of 1  $\mu\text{g}$  RNA, 1  $\mu\text{L}$  oligo (dT)<sub>18</sub> primer and 1  $\mu\text{L}$  nuclease-free water. Subsequent cDNA syntheses were separately conducted by the addition of 4  $\mu\text{L}$  5X reaction buffer, 20  $\mu\text{L}$  RiboLock RNase inhibitor, 2  $\mu\text{L}$  10 mM dNTP Mix and 200 $\mu\text{L}$  RevertAid H Minus M-MuLV reverse transcriptase in total reaction volumes of 20  $\mu\text{L}$ . The reactions were incubated at  $45^\circ\text{C}$  for 1 hour, and reverse transcriptase subsequently inactivated at  $70^\circ\text{C}$  for 5 min. The cDNA generated was stored at  $-20^\circ\text{C}$ .

### Real-time qRT-PCR of genes for cytochrome P450

Expression profiles of the *CYP6M2*, *CYP6P3* and *CYP6Z1* genes for cytochrome P450 enzymes in the mosquito strains tolerant to cadmium or lead, and their respective controls, were separately assessed. These genes have been associated with resistance to pyrethroids [10-12], and are potentially responsive to xenobiotics such as heavy metals [13]. Reaction volumes of 15  $\mu\text{L}$  for each strain consisting of 3  $\mu\text{g}$  cDNA template were amplified in three independent replicates with 7.5  $\mu\text{L}$  of Fast SYBR® Green Master Mix (Applied Biosystems,

**Table 1 Cytochrome p450 and RPL19-specific primers for qPCR**

Gene	Accession number	Sequence (5'-3')	Expected size of product (bp)
<i>CYP6M2</i>	AGAP008212-RA	F 5' TTCGTCGACTCTCTCACCT 3' R 5' GAAATGTACCGGGACTGGTG 3'	299
<i>CYP6P3</i>	AGAP002865-RA	F 5' AGCTAATTAACGCGGTGCTG 3' R 5' AAGTGTGGATTCGGAGCGTA 3'	121
<i>CYP6Z1</i>	AF487535	F 5' TTACATTCACACTGCACGAG 3' R 5' CTTACAGCACAAATCCAGAT 3'	146
<i>RPL19</i>	XM_001864829.1	F 5' CCAACTCGCGACAAAACATTC 3' R 5' ACCGGCTTCTTGATGATCAGA 3'	75

*CYP6M2*, *CYP6P3* are *CYP6Z1* are classes M2, P3 and Z1 respectively in family 6 of cytochrome p450 genes; *RPL19* is ribosomal protein L19 internal control; F or R superscript are forward or reverse primers respectively.

Carlsbad, CA, USA) in the presence of 0.5 picomoles of specific primers for *CYP6M2*, *CYP6P3* and *CYP6Z1* and Ribosomal protein L19 (*RPL19*) internal control (Table 1). The reactions were carried out in a real-time qPCR machine (Stratagene MX3005P, Aligent Technologies, CA, USA) according to the manufacturer's instructions. The thermo-cycling conditions involved an initial step of 95°C for 30 sec, 40 cycles of 95°C for 3 sec, 60°C for 30 sec, followed by one cycle of 95°C for 30 sec, 60°C for 1 min and 95°C for 30 sec for all the genes.

#### Data analysis

Relative expressions of *CYP6M2*, *CYP6P3* and *CYP6Z1* genes were calculated using GenEx 5.3.6 [20]. Briefly, relative quantities of expression of *CYP6M2*, *CYP6P3*, *CYP6Z1* and *RPL19* genes were determined in relation to a control sample in the female condition, being the sample with the lowest expression in all gene categories. The expressions of *CYP6M2*, *CYP6P3* and *CYP6Z1* were subsequently normalized with *RPL19* reference gene using the geNorm function in GenEx. For the qRT-PCR statistical analysis, the expression data were log transformed (Log10) to normalize the data and homogenize the variance prior to two-way analysis of variance where effect of gene, sex and interaction were evaluated. Means were separated by Tukey's honestly significant difference (HSD) test and

detransformed for reporting. All data analyses were implemented using R-Development-Core-Team [21].

#### Result and Discussion

Relative expression profiles of *CYP6M2*, *CYP6P3* and *CYP6Z1* genes of CYP6 family of cytochrome p450 genes in cadmium and lead-tolerant *An. gambiae s. s.* and their respective controls are summarized in Table 2. Expressions were significantly lower in females than males in *CYP6M2* ( $F_{1,17} = 21.35$ ,  $P < 0.001$ ), *CYP6P3* ( $F_{1,17} = 7.35$ ,  $P = 0.019$ ) and *CYP6Z1* ( $F_{1,17} = 11.54$ ,  $P = 0.005$ ) genes. Within gender, there was between 3.1 and 4.2-fold reduction in expression of the genes in lead-selected than control among male populations. Similarly, there was between 1.2 and 2.2-fold reduction in expression of the genes in cadmium-selected than respective male control populations. The reductions were not statistically significant. In both cases, most pronounced reductions were observed in expression of *CYP6P3*. However, the expression of all the classes of genes was marginally elevated in cadmium-selected female populations relative to their respective controls. Expressions of all the genes of CYP6 family of cytochrome450 genes assessed in this study were generally depressed by cadmium or lead tolerance in the mosquito in a sex-dependent manner.

**Table 2 Relative mean (± SE) expression levels of various cytochrome p450 genes in heavy metal-tolerant adult *Anopheles gambiae s.s* mosquito**

Gender	Treatment	<i>CYP6M2</i>	<i>CYP6P3</i>	<i>CYP6Z1</i>
Males	Cont	48.386 ± 15.082	25.894 ± 12.015	28.220 ± 11.883
	Cd	38.052 ± 8.370	11.196 ± 3.280	18.287 ± 6.771
	Pb	15.820 ± 4.858	6.110 ± 1.663	8.999 ± 1.959
Females	Cont	6.347 ± 4.241	5.853 ± 2.456	5.737 ± 3.351
	Cd	9.694 ± 3.826	6.006 ± 1.727	6.998 ± 1.535
	Pb	5.145 ± 1.235	3.831 ± 0.800	4.678 ± 1.267

*CYP6M2*, *CYP6P3* are *CYP6Z1* are classes M2, P3 and Z1 respectively in family 6 of cytochrome p450 genes. Cont, Cd and Pb represent control, cadmium and lead-tolerant populations of *Anopheles gambiae s.s.* respectively.

Factors underlying the observed differential responses to the metals in the mosquito are not apparent. Although xenobiotics-associated induction of cytochrome p450 genes in *An. gambiae* inhabiting polluted water bodies has putatively been linked to induction of resistance to pyrethroids in the mosquito, [1,3,22], The findings point to the contrary. Heavy metal exposure down-regulated expression of cytochrome p450 gene [23,24]. Published data suggest that the depressed expression observed in this study could be linked to cadmium induction of haem oxygenase associated with haem degradation [23] or lead inhibition of gamma aminolevulinic acid dehydratase associated with synthesis of haem [25]. Either case would deplete haem, a major component of the CYP6 family of cytochrome p450 gene, precipitating observed reduced expression of the gene. Similarly, sex-specific responses in cadmium-tolerant populations may be linked to female-specific, cadmium-binding protein in the mosquitoes which might have reduced bioavailability of cadmium to cytochrome p450, and subsequently reduced cadmium induced heme-degradation and hence depletion of CYP6 family of cytochrome 450 gene [23]. Further bioassay studies on metal tolerant populations will shed light on influence of the observed genes expression dynamics on susceptibility of the mosquito to classes of insecticides, including pyrethroids, metabolised by CYP6 family of cytochrome 450 enzyme.

## Conclusion

Tolerance to cadmium or lead in *An. gambiae* s.s significantly influences expression of *CYP6M2*, *CYP6P3* and *CYP6Z1* genes of CYP6 family of cytochrome p450 gene in a metal and sex-specific manner. These changes in gene expression profiles can potentially affect susceptibility of the mosquito to pyrethroid insecticides, and influence vector control efforts.

## Competing interests

The authors declare that they have no competing interests

## Authors' contributions

FKM, DKM and POM conceived the study and participated in the implementation, data analysis, interpretation and manuscript preparation. IAO, AOI and RM guided the study from the conception to manuscript finalization. RO participated in the design of the study. All authors read and approved the final manuscript.

## Acknowledgements

We thank Dr Fathiya Khamis for her critical review of the manuscript and Mr Fidelis Ombura for his technical assistance with real-time qPCR. We also thank Dr. Daisy Salifu for assistance with statistical analysis. This study was funded by an intramural grant from the Research and Extension Division of Egerton University, Njoro, Kenya.

## Author details

<sup>1</sup>Department of Biochemistry and Molecular Biology, Egerton University, P.O. Box 536, Njoro, Kenya. <sup>2</sup>Molecular Biology and Bioinformatics Unit, International Centre of Insect Physiology and Ecology, P.O. Box 30772-00100, Nairobi, Kenya. <sup>3</sup>Centre for Biotechnology and Bioinformatics, University of Nairobi, P.O. Box 29053-00625, Nairobi, Kenya.

Received: 9 February 2013 Accepted: 11 March 2013

Published: 15 March 2013

## References

- Awolola TS, Oduola AO, Obansa JB, Chukwurar NJ, Unyimadu JP: *Anopheles gambiae* s. s. breeding in polluted water bodies in urban Lagos, southwestern Nigeria. *J Vector Borne Dis* 2007, **44**:241–244.
- Mireji PO, Keating J, Hassanali A, Mbogo CM, Nyambaka H, Kahindi S, Beier JC: Heavy metals in mosquito larval habitats in urban Kisumu and Malindi, Kenya, and their impact. *Ecotoxicol Environ Saf* 2008, **70**:147–53.
- Djouaka RF, Bakare AA, Bankole HS, Doannio JM, Coulibaly ON, Kossou H, Tamo M, Basene HI, Popoola OK, Akogbeto MC: Does the spillage of petroleum products in *Anopheles* breeding sites have an impact on the pyrethroid resistance? *Malar J* 2007, **6**:159.
- Elissa N, Mouchet J, Riviere F, Meunier JY, Yao K: Status of insecticide susceptibility in *Anopheles gambiae* s.l. from malaria surveillance sites in The Gambia. *Ann Soc Belg Med Trop* 1993, **73**:291–294.
- Vulule JM, Beach RF, Atieli FK, Roberts JM, Mount DL, Mwangi RW: Malaria Mosquito resistance to agricultural insecticides: risk area mapping in Thailand. *Med Vet Entomol* 1994, **48**:71–75.
- Chandre F, Manguin S, Brengues C, Dossou Yovo J, Darriet F, Diabate A, Carnevale P, Guillet P: Distribution of knock-down resistance mutations in *Anopheles gambiae* molecular forms in west and west-central Africa. *Parasitologia* 1999, **41**:319–322.
- Diabaté A, Baldet T, Chandre F, Akoobeto M, Guiguemde TR, Darriet F, Brengues C, Guillet P, Hemingway J, Small GJ, Hougard JM: The role of agricultural use of insecticides in resistance to pyrethroids in *Anopheles gambiae* s.l. in Burkina Faso. *Am J Trop Med Hyg* 2002, **67**:617–622.
- Diabaté A, Brengues C, Baldet T, Dabire KR, Hougard JM, Akogbeto M, Kengne P, Simard F, Guillet P, Hemingway J, Chandre F: The spread of the Leu-Phe kdr mutation through *Anopheles gambiae* complex in Burkina Faso: genetic introgression and de novo phenomena. *Trop Med Int Health* 2004, **9**:1267–1273.
- Chouaibou M, Etang J, Brévault T, Nwane P, Hinzoumbé CK, Mimpfoundi R, Simard F: Dynamics of insecticide resistance in the malaria vector *Anopheles gambiae* s.l. from an area of extensive cotton cultivation in Northern Cameroon. *Trop Med Int Health* 2008, **13**:476–86.
- Djouaka RF, Bakare AA, Coulibaly ON, Akogbeto MC, Ranson H, Hemingway J, Strode C: Expression of the cytochrome P450s, CYP6P3 and CYP6M2 are significantly elevated in multiple pyrethroid resistant populations of *Anopheles gambiae* s.s. from Southern Benin and Nigeria. *BMC Genomics* 2008, **9**:538.
- Muller P, Warr E, Stevenson BJ, Pignatelli PM, Morgan JC, Steven A, Yawson AE, Ranson H, Hemingway J, Paine MJ, Donnelly MJ: Field-caught permethrin-resistant *Anopheles gambiae* over express CYP6P3, a P450 that metabolises pyrethroids. *Genetics* 2008, **4**:11.
- Nikou D, Ranson H, Hemingway J: An adult-specific CYP6 P450 gene is overexpressed in a pyrethroid-resistant strain of the malaria vector, *Anopheles gambiae*. *Gene* 2003, **318**:91–102.
- Poupardin R, Reynaud S, Strode C, Ranson H, Vontas J, David JP: Cross-induction of detoxification genes by environmental xenobiotics and insecticides in the mosquito *Aedes aegypti*: Impact on larval tolerance to chemical insecticides. *Insect Biochem Mol Biol* 2008, **38**:540–551.
- Mireji PO, Keating J, Kenya E, Mbogo C, Nyambaka H, Osir E, Githure J, Beier JC: Differential induction of proteins in *Anopheles gambiae sensu stricto* (Diptera: Culicidae) larvae in response to heavy metal selection. *Int J Trop Insect Sci* 2006, **26**:214–226.
- Mireji PO, Keating J, Hassanali A, Impoinvil DE, Mbogo CM, Muturi MN, Nyambaka H, Kenya EU, Githure JI, Beier JC: Expression of metallothionein and  $\alpha$ -tubulin in heavy metal-tolerant *Anopheles gambiae sensu stricto* (Diptera: Culicidae). *Ecotoxicol Environ Saf* 2010, **73**:46–50.
- Mireji PO, Keating J, Hassanali A, Mbogo CM, Muturi MN, Githure JI, Beier JC: Biological cost of tolerance to heavy metals in the mosquito *Anopheles gambiae*. *Med Vet Entomol* 2010, **24**:101–107.
- Etang J, Chandre F, Guillet P, Manga L: Reduced bio-efficacy of permethrin EC impregnated bed nets against an *Anopheles gambiae* strain with oxidase-based pyrethroid tolerance. *Malar J* 2004, **3**:46.
- Ford HR, Green E: Laboratory rearing of *Anopheles albimanus*. *Mosquito News* 1972, **32**:509–513.
- Sambrook J, Fritsch EF, Maniatis T: *Molecular Cloning: A Laboratory Manual second ed.* New York: Cold Spring Harbour Laboratory Press; 1989.

20. Mangalam H, Stewart J, Zhou J, Schlauch K, Waugh M, Chen G, Farmer AD, Colello G, Weller JW: **GeneX: An open source gene expression database and integrated tool set.** *IBM Systems Journal* 2001, **40**:552–569.
21. R-Development-Core-Team: *R: A language and environment for statistical computing.* R Foundation for Statistical Computing [WWW document]; 2011. <http://www.R-project.org>.
22. Yadouleton AW, Padonou G, Asidi A, Moiroux N, Bio-Banganna S, Corbel V, N'guessan R, Gbenou D, Yacoubou I, Gazard K, Akogbeto MC: **Insecticide resistance status in *Anopheles gambiae* in southern Benin.** *Malar J* 2010, **9**:83.
23. Gordon GG, Skett P: *Introduction to drug metabolism.* 3rd edition. United Kingdom: Nelson Thornes publishers; 2001:161–163.
24. Henczova M, Filla A, Komlosi V, Abraham K, Deer KA: *The role of environmental proteomics and genomics.* European Society for Comparative Physiology and Biochemistry 22nd Conference Biological Effect of Pollutants; 2003.
25. Henczova M, Deer KA, Filla A, Komlosi V, Mink J: **Effects of Cu<sup>2+</sup> and Pb<sup>2+</sup> on different fish species: Liver cytochrome P450-dependent monooxygenase activities and FTIR spectra.** *Comp Biochem Physiol C Toxicol Pharmacol* 2008, **148**:53–60.

doi:10.1186/1475-2875-12-97

**Cite this article as:** Musasia et al.: Sex-specific induction of CYP6 cytochrome P450 genes in cadmium and lead tolerant *Anopheles gambiae*. *Malaria Journal* 2013 **12**:97.

**Submit your next manuscript to BioMed Central  
and take full advantage of:**

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at  
[www.biomedcentral.com/submit](http://www.biomedcentral.com/submit)

