

# Optogenetic control of morphogenesis goes 3D

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**The generation of form in living embryos, a process termed “morphogenesis” from the Greek word *μορφογένεση*, is one of the most fascinating unsolved problems in biology. In embryonic epithelia, most attention has been paid to events occurring at the apical surface of epithelia, particularly the regulation of actomyosin contractility during morphogenetic change. In a new report, De Renzis and colleagues demonstrate a key role for regulated actomyosin contractility at the basal surface of the epithelium during formation of the first epithelial fold in *Drosophila* (the “ventral furrow”) (Krueger *et al*, 2018).**

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See also: D Krueger *et al* (December 2018)

Epithelia are the most ancient type of animal tissue, being present in all animals, including those derived from the very base of the metazoan evolutionary tree. Epithelia are also typically the first type of tissue to arise in animal development, appearing during or immediately after the blastocyst stage of embryogenesis. In many species, establishment of the first epithelium is followed by formation of a single epithelial fold, namely the “primitive streak” in vertebrates or the “ventral furrow” in *Drosophila*. Genetic screens in *Drosophila* identified the *twist* and *snail* genes as being required for epithelial folding and the subsequent process of epithelial-to-mesenchymal transition (EMT) that produces mesodermal tissues inside the protective outer epithelium (Simpson, 1983; Nusslein-Volhard *et al*, 1984).

The *Drosophila* ventral furrow has since become an excellent model system for understanding how epithelial folding can occur, with numerous studies demonstrating the importance of apical actomyosin-driven constriction in driving indentation of the epithelium (Leptin & Grunewald, 1990; Sweeton *et al*, 1991; Dawes-Hoang *et al*, 2005; Martin *et al*, 2009). Thanks to these impressive studies, it is often thought that the problem of ventral furrow formation is now solved. However, computer simulations indicate that apical constriction is not actually sufficient to explain the complete invagination of the epithelium. Instead, the models suggest that apical constriction must be accompanied by basal relaxation to achieve the pyramidal cell shape observed in folds. Just such a disassembly of basal contractile actomyosin has been visualised during ventral furrow formation (Sweeton *et al*, 1991; Dawes-Hoang *et al*, 2005). However, whether basal relaxation is truly necessary for folding remained unclear, as some models point to basal relaxation as a passive consequence of apical constriction (Polyakov *et al*, 2014).

In a new study, De Renzis and colleagues deploy optogenetic control of actomyosin contractility to prevent basal relaxation and consequently disrupt ventral furrow formation. The findings unequivocally demonstrate the essential role of basal relaxation in allowing pyramidal cell shape changes necessary for full folding of the epithelium. The authors voice their support for the model that apical constriction, which recruits myosin-II, may ultimately cause basal relaxation through passive depletion of myosin-II. Nevertheless, it will be

interesting to determine whether specific molecular mechanisms exist to antagonise basal myosin-II recruitment and thus promote basal relaxation during ventral furrow formation.

The findings in the *Drosophila* embryo are likely to be relevant to epithelial morphogenesis in other developmental stages and other species. The basal surfaces of many different epithelia experience dynamic changes in locally generated actomyosin contractile forces (He *et al*, 2010; Sherrard *et al*, 2010; Sun *et al*, 2017; Huebner & Wallingford, 2018) as well as in the constraining force of the basal extracellular matrix (Haigo & Bilder, 2011; Diaz-de-la-Loza *et al*, 2018). Thus, a full understanding of epithelial morphogenesis will require moving from 2D analysis of the apical surface towards 3D analysis of tissue mechanics at both the apical and basal surface of epithelia.

## References

- Dawes-Hoang RE, Parmar KM, Christiansen AE, Phelps CB, Brand AH, Wieschaus EF (2005) Folded gastrulation, cell shape change and the control of myosin localization. *Development* 132: 4165–4178
- Diaz-de-la-Loza MD, Ray RP, Ganguly PS, Alt S, Davis JR, Hoppe A, Tapon N, Salbreux G, Thompson BJ (2018) Apical and basal matrix remodeling control epithelial morphogenesis. *Dev Cell* 46: 23–39 e5
- Haigo SL, Bilder D (2011) Global tissue revolutions in a morphogenetic movement controlling elongation. *Science* 331: 1071–1074
- He L, Wang X, Tang HL, Montell DJ (2010) Tissue elongation requires oscillating contractions of a basal actomyosin network. *Nat Cell Biol* 12: 1133–1142

- Huebner RJ, Wallingford JB (2018) Coming to consensus: a unifying model emerges for convergent extension. *Dev Cell* 46: 389–396
- Krueger D, Tardivo P, Nguyen C (2018) Downregulation of basal myosin-II is required for cell shape changes and tissue invagination. *EMBO J* 37: e100170
- Leptin M, Grunewald B (1990) Cell shape changes during gastrulation in *Drosophila*. *Development* 110: 73–84
- Martin AC, Kaschube M, Wieschaus EF (2009) Pulsed contractions of an actin-myosin network drive apical constriction. *Nature* 457: 495–499
- Nusslein-Volhard C, Wieschaus E, Kluding H (1984) Mutations affecting the pattern of the larval cuticle in *Drosophila melanogaster*: I. Zygotic loci on the second chromosome. *Wilehm Roux Arch Dev Biol* 193: 267–282
- Polyakov O, He B, Swan M, Shaevitz JW, Kaschube M, Wieschaus E (2014) Passive mechanical forces control cell-shape change during *Drosophila* ventral furrow formation. *Biophys J* 107: 998–1010
- Sherrard K, Robin F, Lemaire P, Munro E (2010) Sequential activation of apical and basolateral contractility drives ascidian endoderm invagination. *Curr Biol* 20: 1499–1510
- Simpson P (1983) Maternal-zygotic gene interactions during formation of the dorsoventral pattern in *Drosophila* embryos. *Genetics* 105: 615–632
- Sun Z, Amourda C, Shagirov M, Hara Y, Saunders TE, Toyama Y (2017) Basolateral protrusion and apical contraction cooperatively drive *Drosophila* germ-band extension. *Nat Cell Biol* 19: 375–383
- Sweeton D, Parks S, Costa M, Wieschaus E (1991) Gastrulation in *Drosophila*: the formation of the ventral furrow and posterior midgut invaginations. *Development* 112: 775–789



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