Early Exposure is Necessary for the Lifespan Extension Effects of Cocoa in C. elegans

Mihiri Munasinghe^D, Abdullah Almotayri, Jency Thomas, Deniz Heydarian and Markandeya Jois

Department of Physiology, Anatomy and Microbiology, School of Life Sciences, La Trobe University, Bundoora, VIC, Australia.

Nutrition and Metabolic Insights Volume 14: 1-8 © The Author(s) 2021 Article reuse guidelines: sagepub.com/journals-permissions DOI: 10.1177/11786388211029443



ABSTRACT

BACKGROUND: We previously showed that cocoa, a rich source of polyphenols improved the age-associated health and extended the lifespan in C. elegans when supplemented starting from L1 stage.

AIM: In this study, we aimed to find out the effects of timing of cocoa exposure on longevity improving effects and the mechanisms and pathways involved in lifespan extension in C. elegans.

METHODS: The standard E. coli OP50 diet of wild type C. elegans was supplemented with cocoa powder starting from different larval stages (L1, L2, L3, and L4) till the death, from L1 to adult day 1 and from adult day 1 till the death. For mechanistic studies, different mutant strains of C. elegans were supplemented with cocoa starting from L1 stage till the death. Survival curves were plotted, and mean lifespan was reported.

RESULTS: Cocoa exposure starting from L1 stage till the death and till adult day 1 significantly extended the lifespan of worms. However, cocoa supplementation at other larval stages as well as at adulthood could not extend the lifespan, instead the lifespan was significantly reduced. Cocoa could not extend the lifespan of daf-16, daf-2, sir-2.1, and clk-1 mutants.

CONCLUSION: Early-start supplementation is essential for cocoa-mediated lifespan extension which is dependent on insulin/IGF-1 signaling pathway and mitochondrial respiration.

KEYWORDS: Cocoa, polyphenols, lifespan, antioxidants, mechanisms

RECEIVED: May 3, 2021. ACCEPTED: June 11, 2021.

TYPE: Original Research

FUNDING: The author(s) disclosed receipt of the following financial support for the research, authorship, and/or publication of this article: This study was supported by La Trobe University Postgraduate Research Scholarship (LTUPRS) and La Trobe University Full Fee Research Scholarship (LTUFFRS).

Introduction

As a result of combined effects of demographic and epidemiological transitions and "modernization" over the past few decades, well-recognized non-communicable diseases (NCDs) such as cardiovascular diseases, cancers, diabetes and neurodegenerative diseases have become the leading public health challenges globally.¹ Aging has been identified as the main risk factor for these prevalent NCDs.² A growing list of genetical, behavioural and pharmacological interventions that have shown to improve longevity also have proven effective in delaying the onset of age-related diseases and preserving healthspan.3 The well-defined pathways that regulate longevity including insulin/insulin-like growth factor-1 signalling (IIS), mitochondrial respiration, calorie restriction (CR) are also found to be involved in the onset of aging-related diseases.⁴⁻⁸ Therefore, the therapies that modulate these pathways may be beneficial in delaying/preventing the onset of age-related diseases.

In recent years, dietary interventions have been proposed as alternative approaches to drugs for the prevention of agingrelated diseases, particularly the consumption of foods rich in DECLARATION OF CONFLICTING INTERESTS: The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article

CORRESPONDING AUTHOR: Mihiri Munasinghe, Department of Physiology, Anatomy and Microbiology, School of Life Sciences, College of Science Health and Engineering, La Trobe University, Kingsbury Drive, Bundoora, VIC 3086, Australia. Email: M.Munasinghe@latrobe.edu.au

polyphenols.9 Polyphenols have been shown to alleviate ageassociated phenomena such as oxidative stress, chronic inflammation and toxin accumulation.¹⁰ Resveratrol, curcumin, quercetin, catechin are some of the polyphenols well-studied for their antioxidant activity as potent compounds to mitigate the age-associated oxidative stress and damage induced by metabolic production of reactive oxygen species (ROS).11-14 Cocoa, one of the widely consumed foods is known to be rich in polyphenols comparatively in much higher levels of total phenolics and flavonoids than tea and wine.¹⁵ Moreover, the cocoa-based product chocolate has been identified as a significant contributor to the total antioxidant capacity of European and American diets.¹⁶

Previously, we reported the effects of long-term cocoa supplementation (starting from the first larval stage till die) at varying doses (1, 2, 3, 4, 5 mg/ml) on lifespan of Caenorhabditis elegans (C. elegans), with 3 mg/ml dose achieving the maximum mean lifespan extension.¹⁷ In addition, the same dose extended the median lifespan as well, but not the maximum lifespan. Based on these findings, in this study, we wanted to investigate the influence of timing of cocoa exposure on the longevity

 $(\mathbf{\hat{n}})$

Creative Commons Non Commercial CC BY-NC: This article is distributed under the terms of the Creative Commons Attribution-NonCommercial 4.0 License (https://creativecommons.org/licenses/by-nc/4.0/) which permits non-commercial use, reproduction and distribution of the work without further permission provided the original work is attributed as specified on the SAGE and Open Access pages (https://us.sagepub.com/en-us/nam/open-access-at-sage).

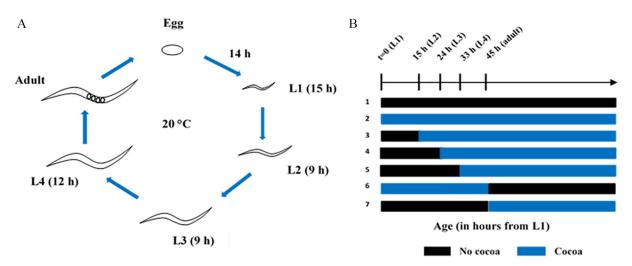


Figure 1. Cocoa-supplementation at different growth stages of *C. elegans*: (A) lifespan of wild-type *C. elegans* at 20°C and (B) schematic representation of cocoa-supplementation at different stages of life (treatment 1 = control; treatment 2 = cocoa from L1 stage; treatment 3 = cocoa from L2 stage; treatment 4 = cocoa from L3 stage; treatment 5 = cocoa from L4 stage; treatment 6 = cocoa from L1-adult day 1; treatment 7 = cocoa from adult day 1 till die).

improving effects with the dose which previously reported the maximum lifespan extension. Additionally, we determined the pathways and mechanisms involved in the lifespan extension effect of cocoa.

Materials and Methods

Strains and culture conditions

All *C. elegans* strains and *Escherichia coli* (*E. coli*) OP50 were acquired from *Caenorhabditis* Genetics Center (CGC) and *C. elegans* were maintained at appropriate temperature (15°C or 20°C) as per instructions. *C. elegans* strains used in this study were as follows: N2 (Bristol, wild-type), DA1116 *eat-2* (*ad1116*) *II.*, VC199 *sir-2.1* (*ok434*) *IV.*, CB4876 *clk-1* (*e2519*) *III.*, EU1 *skn-1* (*zu67*) *IV.*, CB1370 *daf-2* (*e1370*) *III.*, ZG31 *hif-1* (*ia4*) *V.*, TJ356 *daf-16*(*zls356*) *IV.*, and GR1307 *daf-16* (*mgDf50*) *I.* All strains were maintained on nematode growth medium (NGM) plates seeded with *E. coli* OP50.

Cocoa treatment

Specifications and the composition of cocoa powder used in this study as well as the preparation of *E. coli* OP50 food source, and the addition of cocoa to NGM plates were described previously.¹⁷ The major bioactive phytochemicals in cocoa are catechins including mainly monomeric (-) epicatechin and (+) catechin as well as oligomeric and polymeric procyanidins.¹⁸ The total phenolics in cocoa powder used for the study were 27.01 mg GAE/g which was about 56% compared to natural cocoa powder. Total flavonoids were 10.13 mg CE/g which was about 40% compared to the levels in natural cocoa powder. For all the experiments, 3 mg/ml cocoa dose was used. Influence of timing on lifespan extending effects was determined by exposing N2 worms to cocoa at different larval stages (L1, L2, L3, and L4) till they die, L1 to adult day 1 and from adult day 1 till

worms die (Figure 1A and B). *C. elegans* comprises of 4 larval stages (L1, L2, L3, and L4) which are followed by adulthood. The larva emerges from the eggshell has the nervous system and musculature but lacks the reproduction ability. Over four larval stages which ends with a molt, gonad and reproductive system are formed finally making the sexually mature adult worm. The larva is known to grows roughly continuously in size throughout these 4 stages with little change in overall morphology. The larval stages are mainly discriminated by their size. The L1 larvae is about 250 μ m in length while the L2, L3, L4, and adult are 360 to 380 μ m, 490 to 510 μ m, 620 to 650 μ m, and 1110 to 1150 μ m respectively.¹⁹ We supplemented worm diet with cocoa at different larval stages as well as adulthood to represent the different stages of growth and development.

To determine the pathways and mechanisms involved, mutant strains were supplemented with cocoa at a dose of 3 mg/ml starting from L1 stage till the death.

Obtaining synchronous cultures

Age-synchronized worms were obtained by bleaching gravid adults. Briefly, gravid hermaphrodites were washed off from an NGM plate with M9 buffer and made the final volume to 3.5 ml. Bleaching solution (bleach: 5N NaOH = 2:1) was added to the worm suspension at a volume of 1.5 ml. The suspension was mixed by vortexing the tube 5 seconds and keeping further 30 seconds on rest until all the worm bodies got dissolved. The suspension was centrifuged (2 min at 1300g) to pellet the released eggs and the supernatant was poured off. M9 buffer was added to a final volume of 5 ml, centrifuged again and the supernatant was removed. This step was repeated at least 3 times to completely get rid of the bleaching solution. The egg pellet was resuspended in 3 ml of M9 and kept for 48 hours on a shaker for hatching.

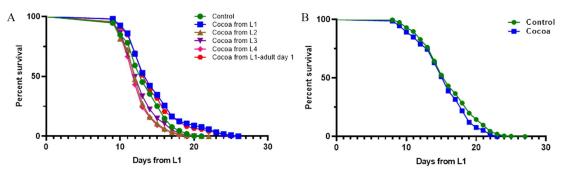


Figure 2. Survival curves for control and cocoa-supplemented wild type (N2) *C. elegans*. Experiments were performed in triplicate. Differences between groups for mean lifespan were calculated using log rank (Mantel-Cox) test. (A) Survival curves for worms supplemented with cocoa at different larval stages (L1, L2, L3, and L4) till die and from L1 to adult day 1. Cocoa significantly increased (P < .05) the mean lifespan when worms were supplemented starting from L1 stage. Cocoa could not extend the lifespan of worms when supplemented at other larval stages (L2, L3, and L4). (B) Survival curves for cocoa-supplemented worms (treatment 7)]. Cocoa could not extend the mean lifespan of worms when supplemented from adulthood.

Lifespan assay

Lifespan assay was performed as described in Sutphin and Kaeberlein.²⁰ Floxuridine (FUdR) was not used to avoid progeny production and instead worms were transferred everyday onto fresh plates until they stop laying eggs (until day 9). Thereafter, worms were transferred every other day. Worms that were crawled off from the plates were excluded from the analysis. Survival curves were plotted. Mean and median lifespan were reported.

DAF-16::GFP localization assay

Control and cocoa-supplemented (3 mg/ml) TJ356 *daf-16(zls356) IV.* worms were scored at L4 stage for cytoplasmic, intermediate or nuclear Green fluorescent protein (GFP) localization as described by Wang et al.²¹ About 100 control and cocoa-supplemented (from L1 stage) worms were collected at L4 stage, washed with M9 buffer twice and placed on a glass slide carrying 2 μ l of 1 M NaN₃. The sub-cellular DAF-16::GFP distribution was observed under a fluorescence microscope at 20-fold magnification. The number of worms in above mentioned three categories were counted and expressed as percentages. Three independent experiments were performed.

Statistical analysis

All statistical analyses were performed in IBM SPSS[®] statistics software (version 24). Data were expressed as mean ± standard error of mean (SEM). Survival function was estimated using Kaplan-Meier curves. Survival curves were compared using log rank (Mantel-Cox) test. The differences between groups for DAF-16 localization was determined using t-Test (two-sample assuming equal variances).

Results

Cocoa extends the lifespan of wild-type C. elegans when supplemented starting from L1 stage

We supplemented the worm diet with cocoa at different larval stages (L1, L2, L3, and L4) till the death, from L1 till adult day

1 and from adult day 1 till the death. Worms showed a significantly extended mean lifespan when supplemented with cocoa starting from L1 stage till they die (P < .05, 8.9% increase, Figure 2A, Table 1). Similarly, cocoa extended the lifespan of worms when supplemented at L1 till only day 1 of adulthood (P < .05, 6.7% increase, Figure 2A, Table 1). However, supplementation of cocoa at L2, L3, or L4 stages could not increase the mean lifespan of worms (Figure 2A, Table 1). Cocoa supplementation at adult day 1 till the death (treatment 7) also could not increase (P < .05) the mean lifespan of worms (Figure 2B, -3.7%, control = 16.2 ± 0.20 , and cocoa = $15.6 \pm$ 0.19 days).

Cocoa extends the lifespan of C. elegans via DAF-16

DAF-16, the *C. elegans* homolog of the forkhead box transcription factors class O (FoxO) is a central regulator of aging, development, stress, metabolism, and immunity.²² Therefore, we tested if DAF-16 can play a role in lifespan extension by cocoa in *C. elegans*. We found that cocoa could not extend the lifespan (P > .05) of *daf-16* mutants (Figure 4A, Table 2). As nuclear localization of DAF-16 is an essential prerequisite for its transcriptional activation, we used the transgenic strain TJ356 (DAF-16::GFP) to explore whether cocoa can activate the nuclear localization of DAF-16. Cocoa treated worms showed a significantly higher (P < .05) nuclear DAF-16 localization compared to control worms as indicated by * (Figure 3A and B).

Cocoa may regulate insulin/insulin-like growth factor-1 signalling (IIS) pathway

In *C. elegans*, IIS pathway is known to regulate longevity via DAF-16.²³ Therefore, we explored whether cocoa could interact with molecules in IIS pathway to regulate the longevity. Single mutations of DAF-2 inhibit IIS pathway and increases longevity in *C. elegans*.²⁴ Therefore, we used *daf-2 (e1370) III*. mutants to study the involvement of IIS pathway in cocoamediated lifespan extension. Cocoa-supplementation could

TREATMENT	SAMPLE SIZE (N)	MEAN LIFESPAN	% EXTENSION COMPARED TO CONTROL	MEDIAN LIFESPAN
1 (control)	344	13.5 ± 0.15		13 ± 0.23
2 (cocoa from L1)	326	$14.7 \pm 0.20^{\star}$	8.9	14 ± 0.21
3 (cocoa from L2)	283	12.7 ± 0.13*	-5.9	12 ± 0.13
4 (cocoa from L3)	209	12.9 ± 0.17*	-4.4	12 ± 0.19
5 (cocoa from L4)	312	$12.5 \pm 0.12^{*}$	-7.4	12 ± 0.12
6 (cocoa from L1-adult day 1)	444	$14.4\pm0.16^{\star}$	6.7	14 ± 0.15

Table 1. Mean and median lifespans of C. elegans when supplemented with cocoa at different stages of life.

*Statistically significant (P < .05) compared to the control group.

not extend the mean lifespan of long-lived insulin-like receptor mutant *daf-2* (Figure 4B, Table 2).

Cocoa could not extend the lifespan of sir-2.1 and eat-2 mutants

The silent information regulator 2 (SIR2) can bind to DAF-16 and extend the lifespan of *C. elegans.*²⁵ We investigated if cocoa could act on SIR-2.1 to extend the lifespan of *C. elegans.* Cocoa could not extend the mean lifespan of *sir-2.1* mutant, indicating that SIR-2.1 is required for cocoa-mediated lifespan extension (Figure 4C, Table 2). In addition, as SIR-2.1 is considered as a key mediator of lifespan extension by CR,²⁶ we used the pharyngeal pumping defective mutant *eat-2* (*ad1116*) *II.* to see if cocoa-induced lifespan extension is dependent on a CR mechanism. According to our results, cocoa could not extend the mean lifespan of *eat-2* mutants (Figure 4D, Table 2).

Lifespan extension by cocoa was not dependent on stress response factors

In *C. elegans skn-1* promotes resistance to oxidative stress and extends lifespan.²⁷ In this study, cocoa-supplementation significantly increased the mean lifespan of *skn-1* mutants (P < .05, Figure 4E, Table 2). Additionally, we determined the effects of cocoa-supplementation on the lifespan of *hif-1* mutants of *C. elegans*. Hypoxia-inducible factor (HIF) enables animals to adapt to the stress caused by hypoxic (low oxygen) conditions that they experience during normal development or during disease. Both HIF-1 over-expression and *hif-1* loss-of-function mutations promote longevity by different pathways in *C. elegans.*²⁸ Our results showed that cocoa supplementation significantly increased the mean lifespan of *hif-1* mutants (P < .05, Figure 4F, Table 2).

Cocoa restores the mitochondrial function to extend the lifespan of C. elegans

The role of mitochondrial function in aging is welldescribed.²⁹⁻³¹Therefore, we used electron transportation chain (ETC) mutant *clk-1* which shows an extended lifespan compared to wild type *C. elegans*³² to see the involvement of mitochondria in cocoa-induced lifespan extension. Cocoa supplementation could not extend the mean lifespan of *clk-1* mutants (Figure 4G, Table 2).

Discussion

In humans, early life nutrition has been reported to significantly influence the risk of developing NCDs in late life,33 emphasizing the importance of early nutrition on healthspan. Earlier initiation of nutritional interventions in humans has been shown to confer greater benefits in long-term cardiometabolic outputs.³⁴ Moreover, studies suggest that early life nutrition can affect intestinal maturation and gut health in later life.³⁵ Enhancing the nutrition at early life is known to regulate the gut microbiota composition and improve the infant immunity development, shaping the life-long health.³⁶ Maternal intake of resveratrol, a polyphenol in Red wine has been shown to fight against the adverse effects of the high-fat diet or lowprotein diet on offspring, such as glucose intolerance, obesity, cholesterol metabolic disorders, non-alcoholic fatty liver disease, or even hypothalamic leptin signaling dysregulation in mice.37

We previously reported that long-term cocoa supplementation extended the lifespan of *C. elegans* when supplemented starting from L1 stage.¹⁷ Based on aforementioned findings on early-life nutrition on health of individuals in later life, we wanted to investigate whether early exposure to cocoa is critical for this longevity improving effect of cocoa.

Our results supported the idea that cocoa supplementation starting from first larval stage (L1) is critical and essential for the longevity extending effects of cocoa. In addition, cocoa intervention starting from L1 stage which continued till day 1 of adulthood increased the lifespan of worms to a similar extent as life-long exposure (L1 to the death). Therefore, these results suggest that the longevity extending effects of cocoa require early-start exposure, but do not need to be continued in a longterm manner. Previous studies support our findings where supplementation of a cranberry extract rich in polyphenols has been prominently promoted the longevity in *C. elegans* when

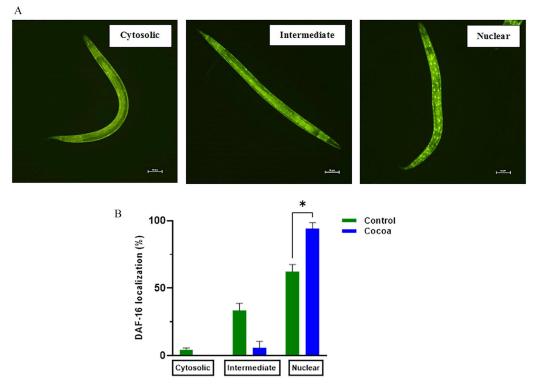


Figure 3. Cocoa-induced nuclear DAF-16 localization in *C. elegans*: (A) representative images of TJ356 *C. elegans* for cytosolic, intermediate and nuclear DAF-16::GFP localization (20-fold magnification, scale bar = 50μ m) and (B) percentage of control and cocoa-treated worms grouped for cytosolic, intermediate and nuclear DAF-16::GFP localization. About 100 worms (at L4 stage) per treatment were used for each replicate. Bar chart represents mean \pm SE of 3 independent experiments. Statistical significance was determined using *t*-Test (two-sample assuming equal variances). Cocoa-treated worms showed a higher nuclear DAF-16 localization compared to control worms (P < .05).

Table 2. Mean and median lifespan of control and cocoa-supplemented C. elegans.	ol and cocoa-supplemented C. elegans.
---	---------------------------------------

STRAIN AND TREATMENT	SAMPLE SIZE (N)	MEAN LIFESPAN	<i>P</i> VALUE	% EXTENSION OF MEAN LIFESPAN COMPARED TO THE CONTROL	MEDIAN LIFESPAN
GR1307 daf-16 (mgDf50) Icontrol	339	14.8 ± 0.11	.12		15 ± 0.13
GR1307 daf-16 (mgDf50) Icocoa	346	15.0 ± 0.11		1.4	15 ± 0.10
CB1370 daf-2 (e1370) IIIcontrol	303	36.1 ± 0.54	.02		38 ± 0.47
CB1370 daf-2 (e1370) IIIcocoa	164	33.7 ± 0.78		-6.6*	35 ± 0.77
VC199 sir-2.1 (ok434) IVcontrol	260	15.7 ± 0.16	.03		16 ± 0.21
VC199 sir-2.1 (ok434) IVcocoa	313	15.0 ± 0.17		-4.5*	15 ± 0.20
DA1116 eat-2 (ad1116) IIcontrol	268	15.9 ± 0.22	.00		15 ± 0.26
DA1116 eat-2 (ad1116) IIcocoa	309	15.2 ± 0.15		-4.4*	15 ± 0.18
EU1 skn-1 (zu67) IVcontrol	239	12.0 ± 0.14	.00		11 ± 0.16
EU1 skn-1 (zu67) IVcocoa	217	13.0 ± 0.17		8.3*	13 ± 0.15
ZG31 hif-1 (ia4) Vcontrol	310	16.8 ± 0.27	.00		17 ± 0.38
ZG31 hif-1 (ia4) Vcocoa	383	19.0 ± 0.22		13.0*	19 ± 0.22
CB4876 clk-1 (e2519) IIIcontrol	353	18.3 ± 0.20	.00		18 ± 0.24
CB4876 clk-1 (e2519) IIIcontrol	360	17.3 ± 0.17		-5.5*	17 ± 0.20

*Statistically significant extension compared to the control group (P < .05).

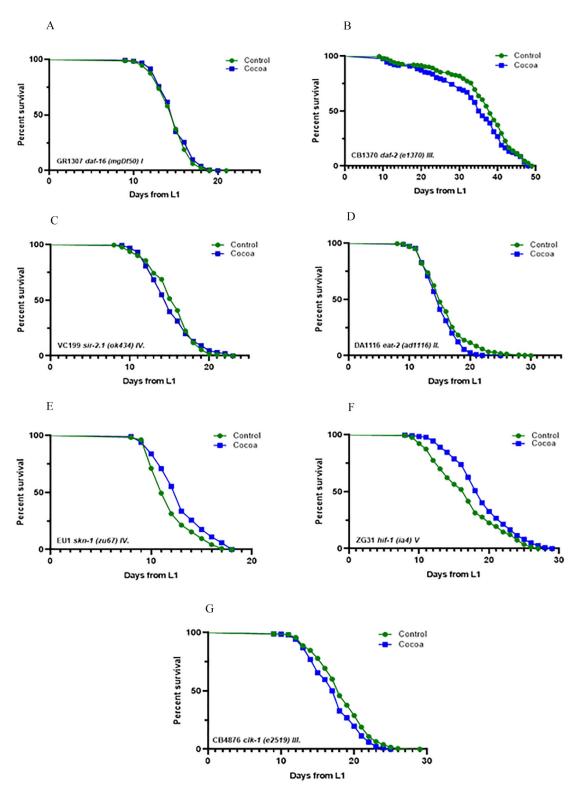


Figure 4. Different pathways and mechanisms involved in cocoa-mediated lifespan extension. Experiments were performed in triplicate. Differences between groups for mean and median lifespans were calculated using log rank (Mantel-Cox) test. (A) Survival curves for control and cocoa-supplemented GR1307 *daf-16 (mgDf50) l.* worms. Cocoa could not extend the lifespan of *daf-16* mutants (P > .05). (B) Survival curves for control and cocoa-supplemented CB1370 *daf-2 (e1370) III*. Worms. Cocoa could not extend the lifespan of *daf-2* mutants. (C) Survival curves for control and cocoa-supplemented VC199 *sir-2.1 (ok434) IV*. worms. Cocoa could not extend the lifespan of *sir-2.1* mutants. (D) Survival curves for control and cocoa-supplemented DA1116 *eat-2 (ad1116) II*. worms. Cocoa could not extend the lifespan of *eat-2* mutants. (E) Survival curves for control and cocoa-supplemented EU1 *skn-1 (zu67) IV*. worms. Cocoa significantly extended the lifespan of *skn-1* mutants (P < .05). (G) Survival curves for control and cocoa-supplemented ZG31 *hif-1 (ia4) V*. worms. Cocoa could not extend the lifespan of *hif-1* mutants (P < .05). (G) Survival curves for control and cocoa-supplemented CB4876 *clk-1 (e2519) III*. worms. Cocoa could not extend the lifespan of *clk-1* mutants.

the supplementation was started at early developmental stage compared to the late-start.³⁸ Moreover, Zuckerman and Geist³⁹ reported that the antioxidant α -tocopherol supplementation at early in the pre-reproductive stage significantly extended both mean and maximum lifespan in *C. elegans*. These effects arose from early supplementation were prominent compared to the supplementation at day 4, indicating α -tocopherol was most effective if present during larval stages.

However, cocoa supplementation at late larval stages (L2, L3, L4) and adulthood did not seem to play a role in lifespan extension. Additionally, supplementation of cocoa at these stages significantly reduced the lifespan. Some of the previous studies have been reported the adverse effects of polyphenols and antioxidant supplementation in animal models. Resveratrol, one of the most researched polyphenols has been reported to act as a pro-oxidant at high concentrations, promoting DNA damage while increasing oxidative stress in vitro and in animal models.⁴⁰ Supplementation of the growth medium with alphatocopherol increased oxidative stress and decreased cell lifespan in Saccharomyces cerevisiae, highlighting the pro-oxidant action of antioxidants.⁴¹ Another study reported that dietary supplementation of both vitamin E and C dramatically shortened the lifespan in voles with no exact explanation for the effect on lifespan but may be due to pro-oxidant effects.⁴² In humans, supplementation of antioxidants (vitamin C and E) has been reported to suppress the health-promoting effects (insulin sensitivity, reactive oxygen species defense) of physical exercise.43 In addition, the consumption of the green tea derived polyphenol (-)-epigallocatechin gallate or its metabolites has been reported to associate with hepatotoxicity in humans.⁴⁴ However, in this study, we were not able to establish the exact mechanism for lifespan reduction effects of cocoa when supplemented at late larval stages and adulthood.

Molecular mechanisms underlying aging have recently gained much attention as aging is the most significant risk factor for many chronic disease conditions.⁴⁵ Most of these mechanisms influencing aging are conserved across organisms.46 Among different genetic factors that regulate aging, IIS, mitochondrial metabolism and CR pathways have been extensively studied.47 To find out which pathways are involved in cocoaassociated lifespan extension in C. elegans, we used several mutant strains related to these well-known pathways. IIS pathway, the first pathway implicated in the aging of animals has a well-established role in aging where the reduced IIS leads to lifespan extension in C. elegans.48 IIS pathway is a signal transduction cascades that consists of insulin-like peptides (ILPs), an insulin/IGF-1 receptor (DAF-2), a phosphoinositide 3-kinase (AGE-1/AAP-1/PI3K), serine/threonine kinases (PDK-1, AKT-1, and AKT-2) and the pivotal downstream FoxO (DAF-16) in C. elegans.²³ In this study, we investigated whether DAF-16 was involved in the lifespan extension by cocoa. Our results showed that cocoa could not increase the lifespan of *daf-16* mutants, indicating that cocoa may regulate lifespan through DAF-16. Moreover, cocoa-treated worms

showed a higher nuclear DAF-16 localization compared to their counterparts, further confirming the involvement of DAF-16. As IIS pathway is a central regulator of DAF-16 activity,²² we investigated whether cocoa can interact with the components in IIS pathway to regulate the longevity of worms. According to the results, cocoa could not improve the lifespan of long-lived insulin like receptor daf-2 (e1370) III. Therefore, the cocoa induced longevity via DAF-16 may be dependent on IIS pathway. In C. elegans, SIR-2.1 is a member of the SIR2 family of NAD+-dependent protein deacetylases and has been shown to regulate nematode aging via the insulin/IGF pathway transcription factor DAF-16.49 Besides, SIR-2.1 is required for lifespan extension by CR, independent of the IIS pathway.⁵⁰ In our study, cocoa treatment could not extend the lifespan in sir-2.1 mutants, suggesting that lifespan extension effect of cocoa is dependent on sir-2.1. Further, cocoa supplementation could not extend the lifespan of pharyngeal pumping defective mutant *eat-2* which mimics the functional effect of CR through the reduced food intake. This suggests that a CR mechanism is involved with the lifespan extension effects of cocoa. However, as we did not determine how cocoa affects the pharyngeal pumping rate of worms, we are unable to establish the exact mechanism for this. Moreover, we previously found that cocoa-treated worms were significantly longer and thicker than the control worms,¹⁷ a result which is inconsistent with the CR effect as caloric restricted worms are thinner than the normally-fed worms.⁵¹ SKN-1 (Skinhead-1) transcription factor in C. elegans which is the homolog of Nrf1/Nrf2 (NF-E2 related factor 1/2) in vertebrates regulates stress resistance and extends lifespan.²⁷ HIF-1 is another stress response molecule that regulates the lifespan of C. elegans.28 Cocoa extended the lifespan of both skn-1 and hif-1 mutants, indicating that lifespan extension by cocoa is not dependent on these stress response molecules.

Mitochondrial respiration is another significant contributor to the aging process.⁵² Impairments in ETC is known to extend the lifespan of *C. elegans.*⁵³ We tested the effects of cocoa supplementation on long-lived *clk-1* mutants. This gene encodes a hydroxylase in the ETC that is required for ubiquinone biosynthesis.⁵⁴ We found that *clk-1* mutants displayed a shorter-lifespan following the cocoa-supplementation, indicating that cocoa's lifespan extension is dependent on mitochondria and cocoa restores mitochondrial function.

Conclusion

In summary, we demonstrated that early-start supplementation is essential for cocoa-mediated lifespan extension in *C. elegans*. In addition, longevity improving effects of cocoa were mediated through IIS pathway and mitochondrial respiration.

Acknowledgements

We would like to thank Dr. Ebony Monson and A/Prof. Karla Helbig from Helbig lab, Department of Physiology, Anatomy and Microbiology, School of Life Sciences, La Trobe University, Bundoora, Australia for the help with the fluorescence microscope for GFP images.

Author Contributions

Study conception and design: MM and MJ; conducting experiments and data collection: MM; data analysis and drafting the manuscript: MM; reviewing of the manuscript; MJ, AA, JT and DH.

ORCID iD

Mihiri Munasinghe D https://orcid.org/0000-0001-5840-740X

REFERENCES

- Gyasi RM, Phillips DR. Aging and the rising burden of noncommunicable diseases in Sub-Saharan Africa and other low- and middle-income countries: a call for holistic action. *Gerontologist*. 2020;60:806-811.
- Zheng S-Q, Huang X-B, Xing T-K, Ding A-J, Wu G-S, Luo H-R. Chlorogenic acid extends the lifespan of *Caenorhabditis elegans* via insulin/IGF-1 signaling pathway. J Gerontol A Biol Sci Med Sci. 2017;72:464-472.
- Barzilai N, Huffman DM, Muzumdar RH, Bartke A. The critical role of metabolic pathways in aging. *Diabetes*. 2012;61:1315-1322.
- Chistiakov DA, Sobenin IA, Revin VV, Orekhov AN, Bobryshev YV. Mitochondrial aging and age-related dysfunction of mitochondria. *Biomed Res Int.* 2014;2014:238463.
- Dillin A, Crawford DK, Kenyon C. Timing requirements for insulin/IGF-1 signaling in C. elegans. Science. 2002;298:830-834.
- Hsu AL, Murphy CT, Kenyon C. Regulation of aging and age-related disease by DAF-16 and heat-shock factor. *Science*. 2003;300:1142-1145.
- 7. Kenyon CJ. The genetics of ageing. Nature. 2010;464:504-512.
- van Exel E, Eikelenboom P, Comijs H, Deeg DJH, Stek ML, Westendorp RGJ. Insulin-like growth factor-1 and risk of late-onset Alzheimer's disease: findings from a family study. *Neurobiol Aging*. 2014;35:725.e7-725.e710.
- Rossi L, Mazzitelli S, Arciello M, Capo CR, Rotilio G. Benefits from dietary polyphenols for brain aging and Alzheimer's disease. *Neurochem Res.* 2008;33: 2390-2400.
- Queen BL, Tollefsbol TO. Polyphenols and aging. Curr Aging Sci. 2010;3: 34-42.
- Grzesik M, Naparło K, Bartosz G, Sadowska-Bartosz I. Antioxidant properties of catechins: comparison with other antioxidants. *Food Chem.* 2018;241:480-492.
- Gülçin İ. Antioxidant properties of resveratrol: a structure-activity insight. Innov Food Sci Emerg Technol. 2010;11:210-218.
- 13. Sökmen M, Akram Khan M. The antioxidant activity of some curcuminoids and chalcones. *Inflammopharmacology*. 2016;24:81-86.
- Zhang M, Swarts SG, Yin L, et al. Antioxidant properties of quercetin. Adv Exp Med Biol. 2011;701:283-289.
- Lee KW, Kim YJ, Lee HJ, Lee CY. Cocoa has more phenolic phytochemicals and a higher antioxidant capacity than teas and red wine. J Agric Food Chem. 2003;51:7292-7295.
- Vinson JA, Proch J, Bose P, et al. Chocolate is a powerful ex vivo and in vivo antioxidant, an antiatherosclerotic agent in an animal model, and a significant contributor to antioxidants in the European and American Diets. *J Agric Food Chem.* 2006;54:8071-8076.
- Munasinghe M, Almotayri A, Thomas J, Heydarian D, Weerasinghe M, Jois M. Cocoa improves age-associated health and extends lifespan in *C. elegans. Nutr Healthy Aging.* 2021;6:73-86.
- Ellam S, Williamson G. Cocoa and human health. Annu Rev Nutr. 2013;33: 105-128.
- Meneely PM, Dahlberg CL, Rose JK. Working with worms: Caenorhabditis elegans as a model organism. Curr Protoc Essent Lab Tech. 2019;19:e35.
- Sutphin GL, Kaeberlein M. Measuring *Caenorhabditis elegans* life span on solid media. J Vis Exp. 2009;27:1152.
- Wang X, Zhang J, Lu L, Zhou L. The longevity effect of echinacoside in *Caenorhabditis elegans* mediated through daf-16. *Biosci Biotechnol Biochem*. 2015;79:1676-1683.
- 22. Zečić A, Braeckman BP. DAF-16/FoxO in *Caenorhabditis elegans* and its role in metabolic remodeling. *Cells*. 2020;9:109.
- Sun X, Chen W-D, Wang Y-D. DAF-16/FOXO transcription factor in aging and longevity. Mini Review. Front Pharmacol. 2017;8:548.
- Zhao X, Lu L, Qi Y, Li M, Zhou L. Emodin extends lifespan of *Caenorhabditis* elegans through insulin/IGF-1 signaling pathway depending on DAF-16 and SIR-2.1. *Biosci Biotechnol Biochem.* 2017;81:1908-1916.

- Berdichevsky A, Viswanathan M, Horvitz HR, Guarente L. C. elegans SIR-2.1 interacts with 14-3-3 proteins to activate DAF-16 and extend life span. Cell. 2006;125:1165-1177.
- Anderson RM, Bitterman KJ, Wood JG, Medvedik O, Sinclair DA. Nicotinamide and PNC1 govern lifespan extension by calorie restriction in Saccharomyces cerevisiae. *Nature*. 2003;423:181-185.
- Tullet JMA, Green JW, Au C, et al. The SKN-1/Nrf2 transcription factor can protect against oxidative stress and increase lifespan in *C. elegans* by distinct mechanisms. *Aging Cell*. 2017;16:1191-1194.
- Zhang Y, Shao Z, Zhai Z, Shen C, Powell-Coffman JA. The HIF-1 hypoxiainducible factor modulates lifespan in *C. elegans. PLoS One.* 2009;4:e6348.
- Harman D. The biologic clock: the mitochondria? J Am Geriatr Soc. 1972;20:145-147.
- Linnane AW, Marzuki S, Ozawa T, Tanaka M. Mitochondrial DNA mutations as an important contributor to ageing and degenerative diseases. *Lancet*. 1989;1:642-645.
- Miquel J, Economos AC, Fleming J, Johnson JE Jr. Mitochondrial role in cell aging. *Exp Gerontol.* 1980;15:575-591.
- Felkai S, Ewbank JJ, Lemieux J, Labbé JC, Brown GG, Hekimi S. CLK-1 controls respiration, behavior and aging in the nematode *Caenorhabditis elegans*. *EMBO J*. 1999;18:1783-1792.
- Kelishadi R, Farajian S. The protective effects of breastfeeding on chronic noncommunicable diseases in adulthood: a review of evidence. *Adv Biomed Res.* 2014;3:3.
- He S, Stein AD. Early-life nutrition interventions and associated long-term cardiometabolic outcomes: a systematic review and meta-analysis of randomized controlled trials. *Adv Nutr.* 2020;12:461-489.
- Ley D, Desseyn J-L, Gouyer V, et al. Early life nutrition influences susceptibility to chronic inflammatory colitis in later life. *Sci Rep.* 2019;9:18111.
- Zhou X, Du L, Shi R, Chen Z, Zhou Y, Li Z. Early-life food nutrition, microbiota maturation and immune development shape life-long health. *Crit Rev Food Sci Nutr.* 2019;59:S30-S38.
- Zhou L-Y, Deng M-Q, Zhang Q, Xiao X-H. Early-life nutrition and metabolic disorders in later life: a new perspective on energy metabolism. *Chin Med J.* 2020;133:1961-1970.
- Guha S, Natarajan O, Murbach CG, et al. Supplement timing of cranberry extract plays a key role in promoting *Caenorhabditis elegans* healthspan. *Nutrients*. 2014;6:911-921.
- Zuckerman BM, Geist MA. Effects of vitamin E on the nematode *Caenorhabdi*tis elegans. Age. 1983;6:1-4.
- Shaito A, Posadino AM, Younes N, et al. Potential adverse effects of resveratrol: a literature review. Int J Mol Sci. 2020;21:2084.
- Lam YT, Stocker R, Dawes IW. The lipophilic antioxidants alpha-tocopherol and coenzyme Q10 reduce the replicative lifespan of Saccharomyces cerevisiae. *Free Radic Biol Med.* 2010;49:237-244.
- Selman C, McLaren JS, Collins AR, et al. Deleterious consequences of antioxidant supplementation on lifespan in a wild-derived mammal. *Biol Lett.* 2013;9:20130432.
- 43. Ristow M, Zarse K, Oberbach A, et al. Antioxidants prevent health-promoting effects of physical exercise in humans. *Proc Natl Acad Sci.* 2009;106:8665-8670.
- Mazzanti G, Menniti-Ippolito F, Moro PA, et al. Hepatotoxicity from green tea: a review of the literature and two unpublished cases. *Eur J Clin Pharmacol.* 2009;65:331-341.
- Uno M, Nishida E. Lifespan-regulating genes in C. elegans. NPJ Aging Mech Dis. 2016;2:16010.
- Curran SP, Ruvkun G. Lifespan regulation by evolutionarily conserved genes essential for viability. *PLoS Genet*. 2007;3:e56.
- 47. Stein G, Murphy C. The intersection of aging, longevity pathways, and learning and memory in *C. elegans*. Review. *Front Genet*. 2012;3:259.
- Kenyon C, Chang J, Gensch E, Rudner A, Tabtiang R. A C. elegans mutant that lives twice as long as wild type. Nature. 1993;366:461-464.
- Viswanathan M, Kim SK, Berdichevsky A, Guarente L. A role for SIR-2.1 regulation of ER stress response genes in determining *C. elegans* life span. *Dev Cell.* 2005;9:605-615.
- Wang Y, Tissenbaum HA. Overlapping and distinct functions for a *Caenorhabditis elegans* SIR2 and DAF-16/FOXO. *Mech Ageing Dev.* 2006;127:48-56.
- Lenaerts I, Walker GA, Van Hoorebeke L, Gems D, Vanfleteren JR. Dietary restriction of *Caenorhabditis elegans* by axenic culture reflects nutritional requirement for constituents provided by metabolically active microbes. *J Gerontol A Biol Sci Med Sci.* 2008;63:242-252.
- 52. Lee HC, Wei YH. Mitochondrial alterations, cellular response to oxidative stress and defective degradation of proteins in aging. *Biogerontology*. 2001;2: 231-244.
- Butler JA, Ventura N, Johnson TE, Rea SL. Long-lived mitochondrial (Mit) mutants of *Caenorhabditis elegans* utilize a novel metabolism. *FASEB J*. 2010;24: 4977-4988.
- Wong A, Boutis P, Hekimi S. Mutations in the clk-1 gene of *Caenorhabditis elegans* affect developmental and behavioral timing. *Genetics*. 1995;139:1247-1259.