Natural products as potential cancer therapy enhancers: A preclinical update

SAGE Open Medicine 2: 2050312114546924 © The Author(s) 2014 Reprints and permissions: sagepub.co.uk/journalsPermissions.nav DOI: 10.1177/2050312114546924 smo.sagepub.com



Abed Agbarya¹, Nili Ruimi², Ron Epelbaum¹, Eran Ben-Arye^{3,4} and Jamal Mahajna^{2,5}

Abstract

Cancer is a multifactorial disease that arises as a consequence of alterations in many physiological processes. Recently, hallmarks of cancer were suggested that include sustaining proliferative signaling, evading growth suppressors, resisting cell death, enabling replicative immortality, inducing angiogenesis, and activating invasion and metastasis, along with two emerging hallmarks including reprogramming energy metabolism and escaping immune destruction. Treating multifactorial diseases, such as cancer with agents targeting a single target, might provide partial treatment and, in many cases, disappointing cure rates. Epidemiological studies have consistently shown that the regular consumption of fruits and vegetables is strongly associated with a reduced risk of developing chronic diseases, such as cardiovascular diseases and cancer. Since ancient times, plants, herbs, and other natural products have been used as healing agents. Moreover, the majority of the medicinal substances available today have their origin in natural compounds. Traditionally, pharmaceuticals are used to cure diseases, and nutrition and herbs are used to prevent disease and to provide an optimal balance of macro- and micro-nutrients needed for good health. We explored the combination of natural products, dietary nutrition, and cancer chemotherapeutics for improving the efficacy of cancer chemotherapeutics and negating side effects.

Keywords

Cancer, cancer therapy, natural products, herbs, nutrition

Date received: 11 February 2014; accepted: 18 July 2014

Introduction

A long history exists of natural products originating from plants, fungi, and microorganisms that have been used for the treatment and prevention of human diseases. In recent years, there has been an emerging focus on the exploration of natural products, including dietary phytoconstituents, in cancer prevention and treatment. An analysis of the origin of drugs developed between 1981 and 2002 showed that natural products or natural-product-derived drugs comprise 28% of all novel chemical entities (NCEs) launched into the market.¹ Examples of anti-cancer agents originating from natural sources include vinblastine from *Vinca rosea*, one of the earliest examples, and paclitaxel, the most recent example, which originates from a Chinese pacific yew plant.² Other plant-derived anti-cancer agents include etoposide, teniposide, homoharringtonine, and camptothecin derivatives.

Natural products as inhibitors of cancer cell proliferation and as inducers of cancer cell cycle arrests and apoptosis

Proliferation is the multiplication or reproduction of cells resulting in the rapid expansion of a cell population. Mammalian cell growth and proliferation are mediated via cell cycle progression. In each cell division cycle, chromosomes are replicated once (DNA synthesis or S-phase) and segregated to create two genetically identical daughter cells (mitosis or M-phase). These events are spaced by intervals of growth and reorganization (gap phases G1 and G2). Progression through the G1 phase of the cell division cycle is

Corresponding author:

Jamal Mahajna, Cancer Drug Discovery Program, MIGAL-Galilee Research Institute, Kiryat Shmona 11016, Israel. Email: jamalm@migal.org.il

Creative Commons CC-BY-NC: This article is distributed under the terms of the Creative Commons Attribution-NonCommercial 3.0 License (http://www.creativecommons.org/licenses/by-nc/3.0/) which permits non-commercial use, reproduction and distribution of the work without further permission provided the original work is attributed as specified on the SAGE and Open Access page (http://www.uk.sagepub.com/aboutus/openaccess.htm).

¹Thoracic Oncology Clinic, Division of Oncology, Rambam Health Care Campus, Haifa, Israel

²Cancer Drug Discovery Program, MIGAL-Galilee Research Institute, Kiryat Shmona, Israel

³Complementary and Traditional Medicine Unit, Department of Family Medicine, Faculty of Medicine, Technion-Israel Institute of Technology, Haifa, Israel

⁴Integrative Oncology Program, The Oncology Service, Lin Medical center, Clalit Health Services, Haifa and Western Galilee District, Israel ⁵Department of Nutritional Sciences, Tel-Hai College, Kiryat Shmona, Israel

Table 1. Major anti-cancer natural products and their principal target genes.

Major natural products	Principal target genes	
Curcumin	Wnt/β-catenin pathway; ⁶⁵ MMP2; ^{40,63} MMP14; ⁴⁰ TIMP-1; ⁶³ Gelatinase; ⁴⁰ EGF; ^{51–54} MMP9; ^{41,} ^{39,43} VEGF; ^{39,43,63} bFGF; ⁶³ NF-κB; ⁵ STAT3; ⁵ PI3K/AKT; ⁵ mTOR; ^{6,7} JNK; ⁵¹ ERK1/2; ⁵¹ uPA; ⁵¹ VEGF; ⁴² KDR; ⁴² Angiopoietin1/2 ⁴²	
Sulforaphane (SFN)	Wnt/β-catenin pathway; ⁷² IL-6; ⁶⁰ IL-1β; ⁶⁰ TNFα; ⁶⁰ PDGF; ⁶⁰ VEGF; ⁶⁰ NF-κB; ⁷⁴ GATA6 ⁷⁴	
Resveratrol	β-catenin; ⁷¹ VEGF; ⁴⁶ Src; ^{15,46} NF-κB; ^{14,15} AP1; ^{14,15} Egr1; ^{14,15} MAPKs; ^{14,15} AR; ¹⁶ AKT; ^{14,16} Caspase- 9; ¹⁴ COX; ¹⁵ NOS; ¹⁵ IL-1β; ⁷⁵ PI3K ⁷⁵	
Caffeic acid phenethyl ester (CAPE)	Wnt/ β -catenin signaling; ⁶⁵ NF- κ B; ³⁰ HER2; ³¹ AKT; ³² ERK; ³² ER- α ; ³² MMP2 ⁶⁶	
Quercetin	Wnt/β-catenin signaling; ⁶⁵ EGF; ^{51,53} VEGF; ⁵⁴ HER2/neu; ^{51,53} HER3; ^{51,53} VEGF-R2; ³⁶ COX- 2; ⁵² iNOS; ⁵² TGF-α; ⁵³ c-Raf; ⁵³ MEK1/2; ⁵³ Elk-1; ⁵³ AKT; ^{36,53} mTOR ³⁶	
EGCG	Wnt/ β -catenin signaling; ⁶⁵ NF- κ B; ¹⁷ DNMT ^{76–78}	
Lycopene	NF-κB; ⁶⁴ MMP9; ⁶⁴ IGF-1; ²⁵ AKT; ^{23–25} β-catenin; ²⁵ cyclin D1; ²⁴ Bad; ²⁴ AR; ²⁵ PSA; ²⁶ pRb; ²³ ICAM- 1; ²² TNFα; ²² SP-1; ⁶⁴ IGF-1R ⁶⁴	
Genistein	EGF; ^{18,51} FOXO3; ¹⁸ NF-кB; ¹⁸ Notch-1; ¹⁸ иPA; ⁵¹ JNK; ⁵¹ ERK1/2 ⁵¹	
13-cis-retinoic acid	IL-2; ⁴⁴ TIMP-1; ⁴⁴ NF- <i>k</i> B; ⁴⁴ ATF-2; ⁴⁴ c-fos ⁴⁴	
Indol-3-carbinol (I3C)	IGF1R;8IRS-1;8ER $lpha$ 8	
Urolithin, ellagitannins and punicalagin	PSA; ²⁰ Aromatase; ²¹ NF-κB ²⁰	
Amentoflavone	COX-2; ⁵² iNOS ⁵²	
Indirubin	VEGF-R2; ³⁸ JAK/STAT3 ³⁸	
Salvianolic Acid B (Sal B)	TGF-β1; ⁶² Smad2/3; ⁶² Smad7; ⁶² MMP2 ⁶²	

13C: indol-3-carbinol; SAL B: salvianolic acid B; *MMP*: matrix metalloproteinase; *TIMP-1*: tissue inhibitor of metalloproteinase-1; *NF*- κ B: nuclear factor kappa-light-chain-enhancer of activated B cells; *EGF*: epidermal growth factor; *FOXO3*: forkhead box O3; *VEGF*: vascular endothelial growth factor; *HER2*: human epidermal growth factor receptor 2; *bFGF*: basic fibroblast growth factor; *STAT3*: signal transducer and activator of transcription 3; *mTOR*: mamma-lian target of rapamycin; *JNK*: c-Jun N-terminal kinases; *ERK*: extracellular-signal-regulated kinase; *uPA*: urokinase-type plasminogen activator; *KDR*: kinase insert domain receptor; *IL*: interleukin; *TNF*: tumor necrosis factor; *PGDF*: platelet-derived growth factor receptor; *MAPK*: mitogen-activated protein kinase; *AR*: androgen receptor; *COX*: cyclooxygenase; NOS: nitric oxide synthase; *ER-* α : estrogen receptor-alpha; *iNOS*: inducible nitric oxide synthase; *DNMT*: DNA cytosine methyltransferase; *IGF*: insulin-like growth factor; *PSA*: prostate specific antigen; ICAM: intercellular adhesion molecule; *SP-1*: specificity protein 1; *ATF-2*: activating transcription factor 2; *GATA6*: GATA-binding factor 6.

a rate-limiting step in mammalian cell proliferation and is governed by numerous mitogenic pathways until the restriction point is passed. Cyclin-dependent kinases (CDK), *CDK4* and *CDK6*, complexed with cyclin D1 are responsible for cell cycle progression through the G1 phase, and the *CDK2/cyclin E* complex functions in the progression of the cell from the late G1 to the early S-phase.

Apoptosis is the predominant mechanism by which cancer cells die when subjected to chemotherapy or irradiation. However, cancer cells develop resistance to these therapies that may be due, at least in part, to the development of effective anti-apoptotic mechanisms.³

More than 600 natural products are reported to possess pharmaceutical activity and many of them exhibit anti-cancer activity. Among the 600 natural products, curcumin (diferuloylmethane), a yellow spice and phenolic compound derived from the plant *Curcuma longa*, is one of the most powerful and promising chemo-preventive and anti-cancer agents.⁴ Curcumin has been found to exert preventive and therapeutic effects in various cancers, in part, due to its ability to influence a diverse range of molecular targets and signaling pathways, including nuclear factor kappa-light-chain-enhancer of activated B cells (*NF-κB*), signal transducer and activator of transcription 3 (*STAT3*), *PI3K* and *AKT* pathways. Moreover, the number of its proposed cellular targets grows as research continues. Recently, curcumin was implicated in modulating cancer cell proliferation by targeting the mammalian target of rapamycin (mTOR) signaling pathways^{6,7} (Table 1).

Another natural product which exhibited anti-cancer activity is indol-3-carbinol (I3C), a natural hydrolysis product of glucobrassicin in cruciferous vegetables. 13C was reported to block proliferation of cancer cells by modulating the expression of insulin-like growth factor receptor-1 (IGF1R) and insulin receptor substrate-1 (IRS1), and to induce protein degradation of estrogen receptor-alpha (ER- α).⁸ Moreover, cruciferous vegetables contain sulforaphane (SFN), a naturally occurring organosulfur compound formed by the hydrolysis of glucosinolates that possess anti-cancer and anti-oxidant activities. Epidemiologic studies suggest that cruciferous vegetable intake may lower overall cancer risk, including colon and prostate cancers.9 SFN was shown to block proliferation and induce cell survival and cell cycle arrest in both in vitro and in vivo systems. SFN also exhibited anti-cancer activity in cancer animal models, as evident from the significant reduction in tumor volume in treated animals.10 Another indole compound derived from cruciferous vegetables is brassinin, which was reported to exhibit anti-proliferative effects against cancer in both in vitro and in vivo models¹¹ (Table 1).

Resveratrol (3,4',5-trihydroxy-trans-stilbene), another widely recognized natural product, is a polyphenolic found in grapes, showing chemo-preventive properties against

several cancers, heart diseases, inflammation, and viral infections. Resveratrol was reported to block proliferation, promote cell cycle arrest, and induce apoptosis in cancer cells, mediated by the suppression of extracellular-signalregulated kinase (ERK)1/2 signaling pathway, p53, Rb/E2F, cyclins, and CDKs. Furthermore, resveratrol affects the activity of transcriptional factors involved in proliferation and stress responses, such as $NF - \kappa B$, activator protein 1 (AP1) and Egr1, mitogen-activated protein kinases (MAPKs) and tyrosine kinases (e.g. Src), leading to apoptosis induction.¹²⁻¹⁵ In addition, resveratrol also inhibits cellular proliferation of prostate cancer cells in both androgen receptor (AR)-dependent and independent mechanisms.Resveratrol inhibits AR transcriptional activity and stimulates phosphatase and tensin homolog (PTEN) expression and decreased AKT phosphorylation¹⁶ (Table 1).

Drinking green tea is also associated with a decreased frequency of cancer development, mainly due to the presence of epigallocatechin gallate (EGCG) and other polyphenols. EGCG suppresses androgen receptor expression and signaling via several growth factor receptors. Moreover, EGCG blocks nuclear translocation of the transcription factor NF- κB .¹⁷

Genistein is an isoflavone found in soy. Soy consumption is associated with a lower incidence of a number of cancers, including colon cancer which is believed to be mediated by genistein. Genistein was reported to inhibit cancer progression and block proliferation, in part by attenuating the negative effect of epidermal growth factor (*EGF*) on forkheadbox O3 (*FOXO3*) activity.¹⁸ However, its therapeutic actions in vivo has been questioned due to contradictory reports from animal studies. Recent in vivo data argue that genistein exhibited a cancer promoting effect¹⁹ (Table 1).

Ellagitannins are bioactive polyphenols found in berries and pomegranate fruit which have attracted recent attention due to their anti-cancer and anti-atherosclerotic, anti-oxidant, and anti-inflammatory bioactivities. Ellagitannins are not absorbed intact into the bloodstream but are hydrolyzed to ellagic acid. They are also metabolized by gut flora into urolithins which are conjugated in the liver and excreted in the urine. These urolithins are also bioactive and inhibit cancer cell proliferation, mediated in part by interfering with the activity of the NF-kB pathway. In clinical studies, pomegranate juice administration led to a decreased rate of prostate specific antigen (PSA) rise after primary treatment with surgery or radiation.²⁰ Moreover, urolithin, an ellagitannins derivative, significantly inhibited testosterone-induced MCF-7aro (MCF-7 that over-expresses aromatase protein) cell proliferation, probably by exhibiting anti-aromatase activity²¹ (Table 1).

Epidemiological studies have shown that the consumption of lycopene is inversely related to human prostate cancer. Moreover, experimental studies have shown that lycopene inhibits the growth of breast, prostate, and endometrial cancer cells with regulation of cell-cycle-related genes, mediated by interfering with $NF \kappa B$ activity.²² In colon cancer cells, lycopene was reported to inhibit the activity of AKT signaling²³ and consequently induced apoptosis.²⁴ In addition, it has been reported that lycopene inhibited insulin-like growth factor-1 (*IGF-1*) mediated *AKT* and *AR* signaling in rat prostate cancer.²⁵ Clinical trials have revealed that lycopene supplements could reduce tumor size and *PSA* level in localized prostate cancers,²⁶ consistent with the down-regulation of *AR* nuclear translocation found during in vitro studies (Table 1).

Moreover, a widely recognized nutritional dietary supplement called Propolis made by honeybees and containing flavonoids, phenolic acids and esters, and caffeic acid phenethyl ester (CAPE) exerted a variety of anti-cancer activities by modulating cell proliferation; induction of cell cycle arrest and apoptosis²⁷ mediated by inhibition of *NF-κB*, *PI3K*, and *p53* signaling pathways;^{28–30} and reduction in phosphorylated human epidermal growth factor receptor 2 (*HER2*) protein in breast cancer cell lines.³¹ Furthermore, the presence of CAPE augmented activity of docetaxel and paclitaxel in prostate cancer cells that was mediated by interfering with *AKT*, *ERK*, and *ER-α* activity³² (Table 1).

Natural products interfering with cancer angiogenesis

Angiogenesis is a physiological process involving the growth of new blood vessels from pre-existing vessels. It is a normal process in growth and development as well as in wound healing. However, this is also a fundamental step in the transition of tumors from a dormant state to a malignant state. Tumors induce angiogenesis by secreting various growth factors, such as vascular endothelial growth factor (VEGF) and basic fibroblast growth factor (bFGF), which induce capillary growth into the tumor and allow it to grow by supplying nutrients and oxygen and removing waste products. In addition, the new vessels allow tumor cells to escape into the circulation and lodge in other organs (i.e. tumor metastases). A vast array of products of natural origin have been shown to have anti-angiogenic potential in preclinical models,33 including Artemisia annua (Chinese wormwood), Viscum album (European mistletoe), Curcuma longa (curcumin), Scutellariabaicalensis (Chinese skullcap), resveratrol and proanthocyanidin (grape seed extract), Magnolia officinalis (Chinese magnolia tree), Camellia sinensis (green tea), Ginkgo biloba, quercetin, Poriacocos, Zingiberofficinalis (ginger), Panax ginseng, Rabdosiarubescenshora(Rabdo sia), and Chinese destagnation herbs^{34,35} (Table 1).

Quercetin, at non-toxic concentrations, was reported to significantly inhibit micro-vessel sprouting, endothelial cell proliferation, migration, invasion, and tube formation, which are key events in the process of angiogenesis. Furthermore, quercetin exhibited anti-angiogenic activity in ex vivo angiogenesis assays, using the chicken egg chorioallantoic membrane (CAM) assay. Moreover, quercetin also exhibited in vivo anti-tumor activity manifested by significant reduction of tumor size in a xenograft mouse model by targeting angiogenesis³⁶ (Table 1).

Green tea polyphenols down-regulated the activity of a number of key enzymes, including *MAPK* and vascular endothelial growth factor receptor (*VEGFR*) signaling, leading to blocking the proliferation of endothelial cells.¹⁷ Also, ellagitannin-rich pomegranate extract was demonstrated to inhibit the proliferation of endothelial cells and to block tumor-associated angiogenesis in animal models.³⁷

Indirubin, the active component of a traditional Chinese herbal medicine, Banlangen, exhibited anti-angiogenic activity when tested in the CAM assay and mouse corneal model. Moreover, indirubin inhibited endothelial cell migration, tube formation, and in vitro cell survival³⁸ (Table 1).

Curcumin was reported to down-regulate the expression of the VEGF and MMP9 genes that are associated with angiogenesis.³⁹ Additionally, curcumin interfered with the activity of both MMP2 and MMP9 and, consequently, reduced degradation of the extracellular matrix (ECM),⁴⁰ leading to a reduction in the levels of released angiogenic factors stored in the ECM. Furthermore, curcumin also inhibits growth factor receptors, such as EGFR and VEGFR, and the other intracellular signaling tyrosine kinases implicated in angiogenesis. Recent reports implicated curcumin in decreasing the gelatinolytic activities of MMP9. In addition, treatment with curcumin inhibited glioma-induced angiogenesis.⁴¹The membrane-bound enzyme CD13 (aminopeptidase N) is found in blood vessels undergoing active angiogenesis. Curcumin binds to CD13 and blocks its activity, thereby inhibiting angiogenesis and invasion of tumor cells^{42,43} (Table 1).

13-cis-retinoic acid significantly inhibited in vitro angiogenesis, as well as micro-vessel sprouting, vascular endothelial (VE) cell proliferation, migration, and tube formation.⁴⁴

Resveratrol inhibits *VEGF*-induced angiogenesis by disruption of reactive oxygen species–dependent *Src* kinase activation and subsequent VE-cadherin tyrosine phosphorylation.^{45,46} Edible berries contain high concentrations of proanthocyanidin which inhibits tumor necrosis factor (*TNF*) α -induced *VEGF* expression. Feeding proanthocyanidins to mice with tumor xenografts resulted in reduced intratumoral microvasculature^{47–49} (Table 1).

In addition to its effect on cell proliferation, *EGFR* is also implicated in cancer angiogenesis.⁵⁰*EGF* stimulates urokinase-type plasminogen activator (*uPA*) expression, which is involved in angiogenesis promotion. Genistein, curcumin, resveratrol, and quercetin were reported to inhibit the effects of *EGF*.^{45,51–54} In in vitro systems, genistein and curcumin inhibit *EGF*-stimulated *uPA* production. Another family member of *EGFR*, the *HER2/neu* gene, is amplified in more than 30% of patients with breast cancer and is correlated with higher levels of angiogenesis.⁵⁵ The activity of herceptin, a drug that inhibits *HER2/neu* expressing breast cancer cells, can be further enhanced by oleic acid.⁵⁶ Interestingly, emodin, a natural constituent of *Polygonummultiflorum* and aloe, inhibits *HER2/neu* expression and exhibits selective cellular toxicity to cancer cells⁵⁷ (Table 1).

Natural products interfering with cancer invasion and metastasis

Metastasis, the spread of cancer cells from the primary tumor to distant organs, is a multi-step process in which cancer cells must invade through the extracellular matrix, intravasate into the bloodstream, survive transport through the circulatory system and, finally, extravasate to distant organs.⁵⁸ Aberrant activation of a developmental program, termed the epithelial-to-mesenchymal transition (EMT), has recently been recognized as an important driver of the metastatic process. EMT is a conserved developmental process in which epithelial cells lose E-cadherin-mediated cell–cell contacts and apical–basal polarity and become motile and invasive. This program is accompanied by expression changes in a variety of genes.

SFN was reported to synergize with the multi-kinase inhibitor, sorafenib, in reducing tumor size of pancreatic cancer in an animal model, due to the blockage of proliferation and angiogenesis, and down-regulation of EMT modulators^{59,60} (Table 1).

In addition, the bioactive component, grape seed proanthocyanidins (GSPs), interfered with the invasion potential of head and neck squamous cell carcinoma (HNSCC). The inhibition of cell invasion by GSPs was associated with the reversal of the EMT process.⁶¹

Salvianolic acid B (Sal B) is a water-soluble component from Danshen (Salvia miltiorrhiza Bunge), a traditional Chinese herb reported to prevent tubular EMT in the fibrotic kidney⁶² (Table 1).

Curcumin is reported to possess anti-invasive activity which is partly mediated by down-regulation of *MMP2* and up-regulation of tissue inhibitor of metalloproteinase-1 *(TIMP1)*,⁶³ enzymes that are involved in the regulation of tumor cell invasion.

Experimental studies have shown that lycopene exhibited anti-cancer activity,²² that mediated, in part, by inhibiting *NF-\kappaB*-mediated expression of *MMP9*, leading to the inhibition of invasion of cancer cells.⁶⁴ In addition, EGCG reduced cancer cell invasiveness through the inhibition of *Wnt* signaling⁶⁵ (Table 1).

Moreover, CAPE found in the nutritional dietary supplement Propolis is reported to interfere with cancer metastasis and invasion by modulating activities of *MMP2*⁶⁶ (Table 1).

Natural products targeting cancer stem cells

The present understanding of cancer biology argues for the existence of a small portion of cells that show stem cell–like characters. These cells constitute a limited subpopulation of

primitive undifferentiated cancer cells that have the ability to self-renew, are tumorigenic and invasive, undergo asymmetrical divisions, and generate all aspects of cancers. Like non-malignant stem cells, putative cancer stem cells (CSC) show remarkable resistance to radiation and chemotherapy.⁶⁷ A number of reports implicate stem-like cells as a potential cause of chemo resistance.⁶⁸ The EMT process that regulates cancer metastasis is also implicated in the generation of CSC and has been associated with resistance to chemotherapy.⁶⁹ In order to cure cancer, it is necessary to eliminate CSC in addition to differentiated cancer cells, to decrease metastasis, reduce recurrence, and improve patient survival.

Diverse dietary constituents, such as vitamins A and D, genistein, EGCG, SFN, piperine, theanine, choline, and curcumin, have been shown to modify self-renewal properties of CSC and influence proliferation, as well as other functions in CSC,⁷⁰ suggesting the potential of using these dietary components in preventing resistance and cancer recurrence. *Wnt* signaling and modulation of β -catenin expression is essential for CSC. A number of phenolic compounds, such as CAPE, curcumin, resveratrol, quercetin, isoflavone, fisetin, EGCG, and isoflavone, were able to inhibit *Wnt* and β -catenin signaling⁶⁵ (Table 1).

Resveratrol has been shown to significantly decrease the level of β -catenin in the nucleus of cancer cells⁷¹ (Table 1). Recent studies in breast cancer cells demonstrated that curcumin inhibited aldehyde dehydrogenases (ALDH)expressing breast CSC self-renewal but did not cause toxicity to differentiated cells by suppressing *Wnt* signaling. Likewise, curcumin has been shown to inhibit CD133 positive medulloblastoma, glioblastoma, pancreatic, and colon CSC proliferation.^{72,73} Moreover, a recent report demonstrated that SFN suppresses the activity of NF-ĸB/GATA6 and thus affects proliferation and migration of vascular smooth muscle cell (VSMC) as well as CSC.74Others have reported activity of SFN against stem cells that is mediated by blocking the *Wnt/\beta-catenin* self-renewal pathway⁷² (Table 1).

Natural products modulate epigenetic modifications

Epigenetics is defined as a heritable modification to the DNA that regulates chromosome architecture and modulates gene expression without changes to the underlying nucleotide sequence, ultimately determining phenotype from genotype. DNA methylation and post-translational histone modifications are classical levels of epigenetic regulation. Epigenetic changes in DNA methylation patterns at CpG sites or deregulated chromatin states of tumor promoting genes and non-coding RNAs emerge as major governing factors in tumor progression and cancer drug sensitivity. DNA methylation in mammals is an enzymatic process primarily mediated by active DNA cytosine methyltransferase (*DNMT*).⁷⁵ During cell division, methylation patterns in the

parental strand of DNA are maintained in the daughter strand by the action of DNMT1, which catalyses the transfer of a methyl group from S-adenosylmethionine (SAM), the methyl donor, to the cytosine residues, restoring the symmetrically methylated CpG dinucleotide pair. Aberrant patterns and dysregulation of DNA methylation cause stable, heritable transcriptional silencing of the associated gene during tumorigenesis.76,77 Epigenetic variability at specific transcription regulation sites appear to be susceptible to modulation by nutritional changes.78 Therefore, dietary components which can affect the process of DNA methylation may influence tumorigenesis by regulation of the expression of certain key genes. Currently, the best evidence to show that nutritional components can modulate epigenetic status of mammal cells comes from studies with mice carrying the agouti viable yellow gene.79,80 Various environmental factors, such as nutrition, remodel our epigenomes lifelong in a beneficial or detrimental way. Since epigenetic marks are reversible in contrast to genetic defects, chemo-preventive nutritional polyphenols are currently evaluated for their ability to reverse adverse epigenetic marks in cancer cells to attenuate tumorigenesis progression, prevent metastasis, or sensitize for drug sensitivity.^{81,82}

Nutrients involved in one-carbon metabolism, namely folate, vitamin B12, vitamin B6, riboflavin, methionine, choline, and betaine, are involved in DNA methylation by regulating the levels of the universal methyl donor SAM and S-adenosylhomocysteine (SAH). Other nutrients and bioactive food components, such as retinoic acid, resveratrol, curcumin, SFN, and tea polyphenols, can modulate epigenetic patterns by altering the levels of SAM and SAH or affecting the catalytic activity of enzymes involved in DNA methylation and histone modifications.^{83,84} Cancer and other agerelated diseases are associated with profound changes in epigenetic patterns, although it is not yet known whether these changes are programmatic or stochastic in nature⁸⁵ (Table 2).

The green tea polyphenol, EGCG, is believed to be a key active ingredient for cancer inhibition through epigenetic control. It has been found that EGCG can reverse CpG island hypermethylation of various methylation-silenced genes and reactivate these gene expressions.⁸⁶ It was also reported that consumption of polyphenols could lead to a decrease in the availability of SAM and an increase in SAH and homocysteine levels. Currently, green tea extracts have been applied in clinical trials, including oral cancer prevention, indicating that tea polyphenols could be used in multiple human cancer preventive and therapeutic purposes due to their bioactivities, such as regulating epigenetic factors⁸⁷ (Table 2).

A methyl donor diet that is used for the synthesis of SAM, including folate and vitamin B12, is expected to affect DNA methylation. Studies on feeding in rats with diets deficient in folate showed a significant genome-wide DNA hypomethylation, as well as gene-specific DNA hypermethylation^{88,89} (Table 2).

 Table 2. Natural products as epigenetics modifiers.

Natural product	Epigenetic activity
EGCG	DNMT inhibitor ^{105,106}
	HAT ¹⁰⁷
	HDAC3 ¹⁰⁷
Parthenolide	DNMT inhibitor ¹⁰⁶
Folate	Methyl group donor ^{89,90,108}
Genistein	DNMT inhibitor ^{91,92,94–96,109}
	HAT inhibitor ^{94–96,109}
	HDAC6110
Caffeic acid phenethyl ester (CAPE)	HDAC inhibitor ³¹
Curcumin	HDAC inhibitor ¹¹¹
	HAT inhibitor ¹¹¹
	DNMT inhibitor ^{106,112,113}
Selenium	Decrease DNMT1 expression ¹¹⁴
	Affect homocysteine availability ¹¹⁴
Methionine	Methyl group donor ^{108,115}
Choline	Methyl group donor ^{108,115}
Betaine	Methyl group donor ^{108,115}
Folate	Methyl group donor ^{115,108}
Vitamin BI2	Methyl group donor ^{115,108}
Resveratrol	Activating SIRT-1 ¹¹⁶
Sulforaphane	HDAC inhibitor ¹¹⁷

EGCG: epigallocatechin gallate; DNMT: DNA cytosine methyltransferase; HAT: histone acetyltransferases; HDAC: histone deacetylases.

The soybean product, genistein, has been shown to be associated with a lower incidence and mortality rate of breast cancer in Asian women who consume soybean products as their daily diet.^{90,91} Genistein is believed to be a chemo-preventive agent against various types of cancer cells.92 It is becoming clear that genistein exerts multiple effects on cancer cell growth, including regulation of gene expression, by modulating epigenetic events such as DNA methylation and/or chromatin modification^{93–95} (Table 2). However, the anti-cancer properties of genistein in breast cancer have raised concerns because of its estrogen-like effect that may be contraindicated for women at high risk of breast cancer. Studies, both in epidemiology and animals, have confirmed that exposure to a soy diet in women in early life greatly impacts breast cancer risk, suggesting exposure time is essential for genistein to exert its effects on breast cancer prevention.

Selenium is an essential trace element with both anti-oxidant and pro-apoptotic properties.^{96,97} Davis et al.⁹⁸ have demonstrated that in the colon and liver, selenium deficiency causes global hypomethylation and in addition promotes methylation of *p53* and *p16* genes, suggesting that impacting DNA methylation may be a crucial mechanism of selenium for cancer prevention. Selenium has been shown to inhibit *DNMT* through direct interaction and indirect action by influencing plasma homocysteine concentrations and the SAM:SAH ratio.^{99,100} Some of the dietary agents, such as butyrate, flavonoids, and curcumin, are capable of altering the epigenetic landscape which can modulate gene/microRNA (miRNA) transcription and subsequently trigger changes in cell proliferation, differentiation, and cell survival^{101,102} (Table 2). Interestingly, several investigators have recently begun to explore how bioactive dietary agents alter the inter-regulatory patterns between promoter regions of miRNAs and several genes.¹⁰³

Natural products' activity mediated by modulation of miRNA expression

miRNAs are small non-coding RNAs (~22 nucleotides long) that play a critical role in basic biological processes, including carcinogenesis. miRNAs are found in both plants and animals and regulate protein expression by acting through complementarity to 3' un-translated regions (UTRs) of their "target" mRNAs, which results in the repression of target gene expression post-transcriptionally.¹⁰⁴ Currently, more than 800 human and mouse miRNAs have been identified that are involved in almost all human malignancies.¹⁰⁵ Furthermore, miRNAs have been correlated to tumor location, mutation status of several tumor suppressor genes/oncogenes, and cancer disease stages. Dietary intake of natural products contributes to disease prevention and therapy, partly due to their capacity to alter the expression of miRNAs and consequently regulate cellular signaling and biological behavior. Curcumin, isoflavone, 3,3'-diinodolylmethane (DIM), I3C, and EGCG are typical examples of natural agents that have been demonstrated to regulate miRNA expression.¹⁰⁶

A growing body of evidence demonstrates that a high intake of n-3 polyunsaturated fatty acids (PUFAs) is protective against tumorigenesis.¹⁰⁷ In contrast, diets rich in n-6 PUFAs (linoleic acid (LA) and arachidonic acid (AA)) enhance both the initiation and promotion of cancer.¹⁰⁸ Recently, miRNA expression of let-7d, miR-15b, miR-107, miR-191, and miR-324-5p were modulated by a n-3 PUFA-enriched diet,¹⁰⁹ arguing that miRNAs may be involved in mediating some of the anti-oncogenic and chemo-protective properties of PUFAs (Table 3).

Butyrate, a short-chain fatty acid produced via fermentation of dietary fiber, exhibited cancer protective effects which are believed to be mediated in part by modulating miRNA expression,¹¹⁰ such as miR-17~92, miR~18b-106a, and miR-106b~25 clusters. The same applies to all-transretinoic acid, the most biologically active metabolites of vitamin A; up-regulated miR-186, miR-215, and miR-223;¹¹¹ and down-regulated miR-17, miR-25, miR-93, miR-193, and miR-181b¹¹² (Table 3).

Polyphenols are ubiquitous secondary metabolites found in dietary nutrition that exhibit chemo-prevention activity against a number of chronic diseases.¹¹³ Some studies have demonstrated that curcumin has protective properties against several

Table 3. Natural products modulate miRNA expression.

Natural product	Up-regulated miRNA	Down- regulated miRNA	Target genes and pathways
EGCG	miR-16, let-7c, miR-18, miR-25, miR-92, ¹³⁷ miR- 210 ¹³⁸	miR-129, miR- 196, miR-200, miR-342, and miR-526 ¹³⁷	HIF-1α
Genistein	miR-200 ¹³⁰		ZEB1, Slug, Vimentin, EMT regulators
Resveratrol	miR-663, miR-21, miR-25, miR-92a, and miR-520h ¹³⁴		EMT, <i>TGF</i> - β, <i>FOXC</i> 2 ¹³⁴
Curcumin	miR-15a, miR-15b, ¹³² miRNA-22, ¹²⁹ miR-200 ¹³⁰	miR-21, ¹³¹ miR-199a ¹²⁹	Bcl2, Cdc25A, EMT
Butyrate	miR-17~92, miR~18b-106a, and miR- 106b~25 ¹²⁴		
All-trans- retinoic acid	miR-186, miR-215, miR-223 ¹²⁵	miR-17, miR- 25, miR-93, miR-193, and miR-181b ¹²⁶	
n-3 PUFA	let-7d, miR-15b, miR-107, miR-191, and miR-324-5 ¹²³		

miRNA: microRNA; PUFA: polyunsaturated fatty acids; EMT: epithelialto-mesenchymal transition; *HIF*: hypoxia-inducible factor; *ZEB1*: zinc finger E-box-binding homeobox 1; *FOXC2*: forkhead box protein C2; *TGF*: transforming growth factor.

types of cancers by the modification of gene expression,¹¹⁴ as well as up-regulation of a subset of miRNAs such as miRNA-22 and down-regulation of another subset of miRNAs such as miR-199a.¹¹⁵ Moreover, DIM and curcumin have been shown to increase the level of the miR-200 family in pancreatic cancer cells, which is involved in the regulation of EMT and invasion behavior, and which was also mechanistically linked to stem cell signatures.¹¹⁶ Curcumin and its synthetic analog, diflourinated curcumin (CDF), down-regulated miR-21 expression¹¹⁷ and reduced the expression of Bcl2 by up-regulating miR-15a and miR-15b.118 DIM was reported to increase the expression of miR-21 and consequently reduced the expression of its target, Cdc25A.119 In addition, resveratrol was also reported to affect the EMT process and transforming growth factor beta (TGF- β) and forkhead box protein C2 (FOXC2) expression by regulating miR-663, miR-21, miR-25, miR-92a, and miR-520h.120 Furthermore, the anti-cancer activity of ellagitannins was shown to be mediated in part by regulating the expression of a number of miRNAs¹²¹ (Table 3).

The EGCG compound exerts its anti-cancer activity by inducing apoptosis, suppressing NF- κB , up- or downregulating tumor suppressor genes/oncogenes, and modulating epigenetic changes of the chromatin.¹²² Interestingly, some EGCG activities are mediated by affecting the expression of miRNAs such as miR-16, let-7c, miR-18, miR-25, and miR-92 which were up-regulated and miR-129, miR-196, miR-200, miR-342, and miR-526 which were down-regulated.¹²³ Moreover, EGCG affects the expression of the hypoxia-inducible factor 1 alpha (*HIF-1a*) pathway, an effect which is mediated by regulation of miR-210¹²⁴ (Table 3).

Soy isoflavones, such as daidzein, genistein, and glycitein, have been reported to have anti-carcinogenic effects mediated by inhibition of cell growth, invasion, and metastasis.¹²⁵ Genistein regulates the expression of miRNAs implicated in controlling cancer cell proliferation,¹²⁶ and also up-regulating miR-200, which was associated with the down-regulation of validated targets zinc finger E-boxbinding homeobox 1 (*ZEB1*), slug, and vimentin, known to play a role in the EMT process¹¹⁶ (Table 3).

Concluding remarks

Many natural products or dietary substances exhibit anticancer activity in in vitro systems against a variety of cancer cell lines, including leukemia, lymphoma, breast, prostate, liver, lung, and myeloma cells. The anti-cancer activity of natural products includes the inhibition of proliferation, induction of apoptosis, induction of cell cycle arrest, inhibition of invasive behavior, and suppression of tumor angiogenesis in many experimental systems. We suggest a need for more in-depth studies that focus on the most promising herbal-derived substances, such as curcumin, genistein, and others. Preliminary clinical data have shown promising efficacies of natural products in cancer treatment as well as in other indications. Yet, few of these natural products have been subjected to randomized clinical trials (RCTs) under the International Conference on Harmonization (ICH) Good Clinical Practice Guidelines to determine their efficacy and/ or safety. The data summarized here show that many nonclinical in vitro and in vivo studies on herbal medicines have commonly supported the traditional therapeutic claims. However, systematic reviews of the study protocols or data interpretation and validation are lacking. We believe that there is a need to explore the full potential of the dietary supplements of natural products, and to assess their safety and efficacy in well-designed, double-blinded, randomized, placebo-controlled clinical trials as stand-alone treatments or in combination with other treatments. To achieve this goal, standardization of pure natural products or active extracts is an important element. Because the composition and amount of biologically active substances depend on sites of production, cultivation conditions, and extraction procedures, standardization will help the acceptance of natural products as suitable for cancer treatment. However, there is a need for the identification and prediction of potential herb-drug interactions.

Declaration of conflicting interests

No author has any conflict of interest to declare.

Funding

This research received no specific grant from any funding agency in the public, commercial, or not-for-profit sectors.

References

- Newman DJ, Cragg GM and Snader KM. Natural products as sources of new drugs over the period 1981–2002. *J Nat Prod* 2003; 66: 1022–1037.
- Butler MS. Natural products to drugs: natural productderived compounds in clinical trials. *Nat Prod Rep* 2008; 25: 475–516.
- 3. Hanahan D and Weinberg RA. Hallmarks of cancer: the next generation. *Cell* 2011; 144: 646–674.
- Bar-Sela G, Epelbaum R and Schaffer M. Curcumin as an anti-cancer agent: review of the gap between basic and clinical applications. *Curr Med Chem* 2010; 17: 190–197.
- Jagtap S, Meganathan K, Wagh V, et al. Chemoprotective mechanism of the natural compounds, epigallocatechin-3-O-gallate, quercetin and curcumin against cancer and cardiovascular diseases. *Curr Med Chem* 2009; 16: 1451–1462.
- Johnson SM, Gulhati P, Arrieta I, et al. Curcumin inhibits proliferation of colorectal carcinoma by modulating Akt/ mTOR signaling. *Anticancer Res* 2009; 29: 3185–3190.
- Beevers CS, Zhou H and Huang S. Hitting the golden TORget: curcumin's effects on mTOR signaling. *Anticancer Agents Med Chem* 2013; 13: 988–994.
- Marconett CN, Singhal AK, Sundar SN, et al. Indole-3carbinol disrupts estrogen receptor-alpha dependent expression of insulin-like growth factor-1 receptor and insulin receptor substrate-1 and proliferation of human breast cancer cells. *Mol Cell Endocrinol* 2012; 363: 74–84.
- Al-Hajj M, Wicha MS, Benito-Hernandez A, et al. Prospective identification of tumorigenic breast cancer cells. *Proc Natl Acad Sci USA* 2003; 100: 3983–3988.
- Qazi A, Pal J, Maitah M, et al. Anticancer activity of a broccoli derivative, sulforaphane, in Barrett adenocarcinoma: potential use in chemoprevention and as adjuvant in chemotherapy. *Transl Oncol* 2010; 3: 389–399.
- 11. Izutani Y, Yogosawa S, Sowa Y, et al. Brassinin induces G1 phase arrest through increase of p21 and p27 by inhibition of the phosphatidylinositol 3-kinase signaling pathway in human colon cancer cells. *Int J Oncol* 2012; 40: 816–824.
- De Leo A, Arena G, Stecca C, et al. Resveratrol inhibits proliferation and survival of Epstein Barr virus-infected Burkitt's lymphoma cells depending on viral latency program. *Mol Cancer Res* 2011; 9: 1346–1355.
- Ding XZ and Adrian TE. Resveratrol inhibits proliferation and induces apoptosis in human pancreatic cancer cells. *Pancreas* 2002; 25: e71–e76.
- Li Y, Liu J, Liu X, et al. Resveratrol-induced cell inhibition of growth and apoptosis in MCF7 human breast cancer cells are associated with modulation of phosphorylated Akt and caspase-9. *Appl Biochem Biotechnol* 2006; 135: 181–192.

- Signorelli P and Ghidoni R. Resveratrol as an anticancer nutrient: molecular basis, open questions and promises. J Nutr Biochem 2005; 16: 449–466.
- Wang Y, Romigh T, He X, et al. Resveratrol regulates the PTEN/AKT pathway through androgen receptor-dependent and -independent mechanisms in prostate cancer cell lines. *Hum Mol Genet* 2010; 19: 4319–4329.
- Beltz LA, Bayer DK, Moss AL, et al. Mechanisms of cancer prevention by green and black tea polyphenols. *Anticancer Agents Med Chem* 2006; 6: 389–406.
- Pan H, Zhou W, He W, et al. Genistein inhibits MDA-MB-231 triple-negative breast cancer cell growth by inhibiting NF-kappaB activity via the Notch-1 pathway. *Int J Mol Med* 2012; 30: 337–343.
- Nakamura H, Wang Y, Kurita T, et al. Genistein increases epidermal growth factor receptor signaling and promotes tumor progression in advanced human prostate cancer. *PLoS One* 2011; 6: e20034.
- 20. Heber D. Multitargeted therapy of cancer by ellagitannins. *Cancer Lett* 2008; 269: 262–268.
- 21. Adams LS, Zhang Y, Seeram NP, et al. Pomegranate ellagitannin-derived compounds exhibit antiproliferative and antiaromatase activity in breast cancer cells in vitro. *Cancer Prev Res (Phila)* 2010; 3: 108–113.
- Hung CF, Huang TF, Chen BH, et al. Lycopene inhibits TNF-alpha-induced endothelial ICAM-1 expression and monocyte-endothelial adhesion. *Eur J Pharmacol* 2008; 586: 275–282.
- 23. Tang FY, Shih CJ, Cheng LH, et al. Lycopene inhibits growth of human colon cancer cells via suppression of the Akt signaling pathway. *Mol Nutr Food Res* 2008; 52: 646–654.
- Palozza P, Sheriff A, Serini S, et al. Lycopene induces apoptosis in immortalized fibroblasts exposed to tobacco smoke condensate through arresting cell cycle and downregulating cyclin D1, pAKT and pBad. *Apoptosis* 2005; 10: 1445–1456.
- Liu X, Allen JD, Arnold JT, et al. Lycopene inhibits IGF-I signal transduction and growth in normal prostate epithelial cells by decreasing DHT-modulated IGF-I production in cocultured reactive stromal cells. *Carcinogenesis* 2008; 29: 816–823.
- Kucuk O, Sarkar FH, Djuric Z, et al. Effects of lycopene supplementation in patients with localized prostate cancer. *Exp Biol Med (Maywood)* 2002; 227: 881–885.
- Chen MJ, Chang WH, Lin CC, et al. Caffeic acid phenethyl ester induces apoptosis of human pancreatic cancer cells involving caspase and mitochondrial dysfunction. *Pancreatology* 2008; 8: 566–576.
- Wu J, Omene C, Karkoszka J, et al. Caffeic acid phenethyl ester (CAPE), derived from a honeybee product propolis, exhibits a diversity of anti-tumor effects in pre-clinical models of human breast cancer. *Cancer Lett* 2011; 308: 43–53.
- Omene CO, Wu J and Frenkel K. Caffeic Acid Phenethyl Ester (CAPE) derived from propolis, a honeybee product, inhibits growth of breast cancer stem cells. *Invest New Drugs* 2012; 30: 1279–1288.
- Onori P, DeMorrow S, Gaudio E, et al. Caffeic acid phenethyl ester decreases cholangiocarcinoma growth by inhibition

of NF-kappaB and induction of apoptosis. *Int J Cancer* 2009; 125: 565–576.

- Omene C, Kalac M, Wu J, et al. Propolis and its active component, Caffeic Acid Phenethyl Ester (CAPE), modulate breast cancer therapeutic targets via an epigenetically mediated mechanism of action. *J Cancer Sci Ther* 2013; 5: 334–342.
- Tolba MF, Esmat A, Al-Abd AM, et al. Caffeic acid phenethyl ester synergistically enhances docetaxel and paclitaxel cytotoxicity in prostate cancer cells. *IUBMB Life* 2013; 65: 716–729.
- Neal CP, Berry DP, Doucas H, et al. Clinical aspects of natural anti-angiogenic drugs. *Curr Drug Targets* 2006; 7: 371–383.
- Sagar SM, Yance D and Wong RK. Natural health products that inhibit angiogenesis: a potential source for investigational new agents to treat cancer-part 1. *Curr Oncol* 2006; 13: 14–26.
- Elluru SR, Duong Van Huyen JP, Delignat S, et al. Antiangiogenic properties of Viscum album extracts are associated with endothelial cytotoxicity. *Anticancer Res* 2009; 29: 2945–2950.
- Pratheeshkumar P, Budhraja A, Son YO, et al. Quercetin inhibits angiogenesis mediated human prostate tumor growth by targeting VEGFR-2 regulated AKT/mTOR/P70S6K signaling pathways. *PLoS One* 2012; 7: e47516.
- Sartippour MR, Seeram NP, Rao JY, et al. Ellagitannin-rich pomegranate extract inhibits angiogenesis in prostate cancer in vitro and in vivo. *Int J Oncol* 2008; 32: 475–480.
- Zhang X, Song Y, Wu Y, et al. Indirubin inhibits tumor growth by antitumor angiogenesis via blocking VEGFR2mediated JAK/STAT3 signaling in endothelial cell. *Int J Cancer* 2011; 129: 2502–2511.
- Kim JH, Shim JS, Lee SK, et al. Microarray-based analysis of anti-angiogenic activity of demethoxycurcumin on human umbilical vein endothelial cells: crucial involvement of the down-regulation of matrix metalloproteinase. *Jpn J Cancer Res* 2002; 93: 1378–1385.
- Chen HW, Yu SL, Chen JJ, et al. Anti-invasive gene expression profile of curcumin in lung adenocarcinoma based on a high throughput microarray analysis. *Mol Pharmacol* 2004; 65: 99–110.
- Perry MC, Demeule M, Regina A, et al. Curcumin inhibits tumor growth and angiogenesis in glioblastoma xenografts. *Mol Nutr Food Res* 2010; 54: 1192–1201.
- Gururaj AE, Belakavadi M, Venkatesh DA, et al. Molecular mechanisms of anti-angiogenic effect of curcumin. *Biochem Biophys Res Commun* 2002; 297: 934–942.
- Hahm ER, Gho YS, Park S, et al. Synthetic curcumin analogs inhibit activator protein-1 transcription and tumorinduced angiogenesis. *Biochem Biophys Res Commun* 2004; 321: 337–344.
- Guruvayoorappan C and Kuttan G. 13 cis-retinoic acid regulates cytokine production and inhibits angiogenesis by disrupting endothelial cell migration and tube formation. *J Exp Ther Oncol* 2008; 7: 173–182.
- Igura K, Ohta T, Kuroda Y, et al. Resveratrol and quercetin inhibit angiogenesis in vitro. *Cancer Lett* 2001; 171: 11–16.

- Lin MT, Yen ML, Lin CY, et al. Inhibition of vascular endothelial growth factor-induced angiogenesis by resveratrol through interruption of Src-dependent vascular endothelial cadherin tyrosine phosphorylation. *Mol Pharmacol* 2003; 64: 1029–1036.
- Roy S, Khanna S, Alessio HM, et al. Anti-angiogenic property of edible berries. *Free Radic Res* 2002; 36: 1023–1031.
- Bagchi D, Bagchi M, Stohs SJ, et al. Free radicals and grape seed proanthocyanidin extract: importance in human health and disease prevention. *Toxicology* 2000; 148: 187–197.
- Singh RP, Tyagi AK, Dhanalakshmi S, et al. Grape seed extract inhibits advanced human prostate tumor growth and angiogenesis and upregulates insulin-like growth factor binding protein-3. *Int J Cancer* 2004; 108: 733–740.
- Casanova ML, Larcher F, Casanova B, et al. A critical role for ras-mediated, epidermal growth factor receptordependent angiogenesis in mouse skin carcinogenesis. *Cancer Res* 2002; 62: 3402–3407.
- Smith PC, Santibanez JF, Morales JP, et al. Epidermal growth factor stimulates urokinase-type plasminogen activator expression in human gingival fibroblasts. Possible modulation by genistein and curcumin. *J Periodontal Res* 2004; 39: 380–387.
- Banerjee T, Van der Vliet A and Ziboh VA. Downregulation of COX-2 and iNOS by amentoflavone and quercetin in A549 human lung adenocarcinoma cell line. *Prostaglandins Leukot Essent Fatty Acids* 2002; 66: 485–492.
- 53. Huynh H, Nguyen TT, Chan E, et al. Inhibition of ErbB-2 and ErbB-3 expression by quercetin prevents transforming growth factor alpha (TGF-alpha)- and epidermal growth factor (EGF)-induced human PC-3 prostate cancer cell proliferation. *Int J Oncol* 2003; 23: 821–829.
- Ma ZS, Huynh TH, Ng CP, et al. Reduction of CWR22 prostate tumor xenograft growth by combined tamoxifenquercetin treatment is associated with inhibition of angiogenesis and cellular proliferation. *Int J Oncol* 2004; 24: 1297–1304.
- Blackwell KL, Dewhirst MW, Liotcheva V, et al. HER-2 gene amplification correlates with higher levels of angiogenesis and lower levels of hypoxia in primary breast tumors. *Clin Cancer Res* 2004; 10: 4083–4088.
- 56. Menendez JA, Vellon L, Colomer R, et al. Oleic acid, the main monounsaturated fatty acid of olive oil, suppresses Her-2/neu (erbB-2) expression and synergistically enhances the growth inhibitory effects of trastuzumab (Herceptin) in breast cancer cells with Her-2/neu oncogene amplification. *Ann Oncol* 2005; 16: 359–371.
- Wasserman L, Avigad S, Beery E, et al. The effect of aloe emodin on the proliferation of a new merkel carcinoma cell line. *Am J Dermatopathol* 2002; 24: 17–22.
- Chambers AF, Groom AC and MacDonald IC. Dissemination and growth of cancer cells in metastatic sites. *Nat Rev Cancer* 2002; 2: 563–572.
- Rausch V, Liu L, Kallifatidis G, et al. Synergistic activity of sorafenib and sulforaphane abolishes pancreatic cancer stem cell characteristics. *Cancer Res* 2010; 70: 5004–5013.
- Hunakova L, Sedlakova O, Cholujova D, et al. Modulation of markers associated with aggressive phenotype in MDA-MB-231 breast carcinoma cells by sulforaphane. *Neoplasma* 2009; 56: 548–556.

- 61. Sun Q, Prasad R, Rosenthal E, et al. Grape seed proanthocyanidins inhibit the invasive potential of head and neck cutaneous squamous cell carcinoma cells by targeting EGFR expression and epithelial-to-mesenchymal transition. *BMC Complement Altern Med* 2011; 11: 134.
- 62. Wang QL, Tao YY, Yuan JL, et al. Salvianolic acid B prevents epithelial-to-mesenchymal transition through the TGFbeta1 signal transduction pathway in vivo and in vitro. *BMC Cell Biol* 2010; 11: 31.
- Shao ZM, Shen ZZ, Liu CH, et al. Curcumin exerts multiple suppressive effects on human breast carcinoma cells. *Int J Cancer* 2002; 98: 234–240.
- Huang CS, Fan YE, Lin CY, et al. Lycopene inhibits matrix metalloproteinase-9 expression and down-regulates the binding activity of nuclear factor-kappa B and stimulatory protein-1. J Nutr Biochem 2007; 18: 449–456.
- Kim J, Zhang X, Rieger-Christ KM, et al. Suppression of Wnt signaling by the green tea compound (-)-epigallocatechin 3-gallate (EGCG) in invasive breast cancer cells. Requirement of the transcriptional repressor HBP1. *J Biol Chem* 2006; 281: 10865–10875.
- 66. Lee KW, Kang NJ, Kim JH, et al. Caffeic acid phenethyl ester inhibits invasion and expression of matrix metalloproteinase in SK-Hep1 human hepatocellular carcinoma cells by targeting nuclear factor kappa B. *Genes Nutr* 2008; 2: 319–322.
- Ribacka C, Pesonen S and Hemminki A. Cancer, stem cells, and oncolytic viruses. *Ann Med* 2008; 40: 496–505.
- Huber M, Bahr I, Kratzschmar JR, et al. Comparison of proteomic and genomic analyses of the human breast cancer cell line T47D and the antiestrogen-resistant derivative T47D-r. *Mol Cell Proteomics* 2004; 3: 43–55.
- Ahmed N, Abubaker K, Findlay J, et al. Epithelial mesenchymal transition and cancer stem cell-like phenotypes facilitate chemoresistance in recurrent ovarian cancer. *Curr Cancer Drug Targets* 2010; 10: 268–278.
- Kim YS, Farrar W, Colburn NH, et al. Cancer stem cells: potential target for bioactive food components. J Nutr Biochem 2012; 23: 691–698.
- Hope C, Planutis K, Planutiene M, et al. Low concentrations of resveratrol inhibit Wnt signal throughput in colon-derived cells: implications for colon cancer prevention. *Mol Nutr Food Res* 2008; 52(Suppl. 1): S52–S61.
- 72. Li Y, Zhang T, Korkaya H, et al. Sulforaphane, a dietary component of broccoli/broccoli sprouts, inhibits breast cancer stem cells. *Clin Cancer Res* 2010; 16: 2580–2590.
- Nautiyal J, Kanwar SS, Yu Y, et al. Combination of dasatinib and curcumin eliminates chemo-resistant colon cancer cells. *J Mol Signal* 2011; 6: 7.
- Kwon JS, Joung H, Kim YS, et al. Sulforaphane inhibits restenosis by suppressing inflammation and the proliferation of vascular smooth muscle cells. *Atherosclerosis* 2012; 225: 41–49.
- 75. Bestor TH. The DNA methyltransferases of mammals. *Hum Mol Genet* 2000; 9: 2395–2402.
- Egger G, Liang G, Aparicio A, et al. Epigenetics in human disease and prospects for epigenetic therapy. *Nature* 2004; 429: 457–463.
- Baylin SB and Ohm JE. Epigenetic gene silencing in cancer—a mechanism for early oncogenic pathway addiction? *Nat Rev Cancer* 2006; 6: 107–116.

- Waterland RA and Jirtle RL. Early nutrition, epigenetic changes at transposons and imprinted genes, and enhanced susceptibility to adult chronic diseases. *Nutrition* 2004; 20: 63–68.
- Yen TT, Gill AM, Frigeri LG, et al. Obesity, diabetes, and neoplasia in yellow A(vy)/- mice: ectopic expression of the agouti gene. *FASEB J* 1994; 8: 479–488.
- Michaud EJ, Van Vugt MJ, Bultman SJ, et al. Differential expression of a new dominant agouti allele (Aiapy) is correlated with methylation state and is influenced by parental lineage. *Genes Dev* 1994; 8: 1463–1472.
- Vanden Berghe W. Epigenetic impact of dietary polyphenols in cancer chemoprevention: lifelong remodeling of our epigenomes. *Pharmacol Res* 2012; 65: 565–576.
- Huang J, Plass C and Gerhauser C. Cancer chemoprevention by targeting the epigenome. *Curr Drug Targets* 2011; 12: 1925–1956.
- Berletch JB, Liu C, Love WK, et al. Epigenetic and genetic mechanisms contribute to telomerase inhibition by EGCG. J *Cell Biochem* 2008; 103: 509–519.
- Mittal A, Piyathilake C, Hara Y, et al. Exceptionally high protection of photocarcinogenesis by topical application of (–)-epigallocatechin-3-gallate in hydrophilic cream in SKH-1 hairless mouse model: relationship to inhibition of UVB-induced global DNA hypomethylation. *Neoplasia* 2003; 5: 555–565.
- Ross SA. Evidence for the relationship between diet and cancer. *Exp Oncol* 2010; 32: 137–142.
- Fang MZ, Wang Y, Ai N, et al. Tea polyphenol (-)-epigallocatechin-3-gallate inhibits DNA methyltransferase and reactivates methylation-silenced genes in cancer cell lines. *Cancer Res* 2003; 63: 7563–7570.
- Yuasa Y, Nagasaki H, Akiyama Y, et al. Relationship between CDX2 gene methylation and dietary factors in gastric cancer patients. *Carcinogenesis* 2005; 26: 193–200.
- Kim YI. Nutritional epigenetics: impact of folate deficiency on DNA methylation and colon cancer susceptibility. *J Nutr* 2005; 135: 2703–2709.
- Stempak JM, Sohn KJ, Chiang EP, et al. Cell and stage of transformation-specific effects of folate deficiency on methionine cycle intermediates and DNA methylation in an in vitro model. *Carcinogenesis* 2005; 26: 981–990.
- Lee HP, Gourley L, Duffy SW, et al. Dietary effects on breast-cancer risk in Singapore. *Lancet* 1991; 337: 1197–1200.
- Fang CY, Tseng M and Daly MB. Correlates of soy food consumption in women at increased risk for breast cancer. J Am Diet Assoc 2005; 105: 1552–1558.
- Barnes S. Effect of genistein on in vitro and in vivo models of cancer. J Nutr 1995; 125: 7778–783S.
- 93. Fang MZ, Chen D, Sun Y, et al. Reversal of hypermethylation and reactivation of p16INK4a, RARbeta, and MGMT genes by genistein and other isoflavones from soy. *Clin Cancer Res* 2005; 11: 7033–7041.
- 94. Li Y, Liu L, Andrews LG, et al. Genistein depletes telomerase activity through cross-talk between genetic and epigenetic mechanisms. *Int J Cancer* 2009; 125: 286–296.
- 95. Majid S, Kikuno N, Nelles J, et al. Genistein induces the p21WAF1/CIP1 and p16INK4a tumor suppressor genes in prostate cancer cells by epigenetic mechanisms involving

active chromatin modification. *Cancer Res* 2008; 68: 2736–2744.

- Clark LC, Cantor KP and Allaway WH. Selenium in forage crops and cancer mortality in U.S. counties. *Arch Environ Health* 1991; 46: 37–42.
- Clark LC, Combs GF Jr, Turnbull BW, et al. Effects of selenium supplementation for cancer prevention in patients with carcinoma of the skin. A randomized controlled trial. Nutritional Prevention of Cancer Study Group. *JAMA* 1996; 276: 1957–1963.
- Davis CD, Uthus EO and Finley JW. Dietary selenium and arsenic affect DNA methylation in vitro in Caco-2 cells and in vivo in rat liver and colon. *J Nutr* 2000; 130: 2903–2909.
- Davis CD and Uthus EO. Dietary selenite and azadeoxycytidine treatments affect dimethylhydrazine-induced aberrant crypt formation in rat colon and DNA methylation in HT-29 cells. *J Nutr* 2002; 132: 292–297.
- Uthus EO and Ross SA. Dietary selenium affects homocysteine metabolism differently in Fisher-344 rats and CD-1 mice. *J Nutr* 2007; 137: 1132–1136.
- Duthie SJ. Epigenetic modifications and human pathologies: cancer and CVD. *Proc Nutr Soc* 2011; 70: 47–56.
- Fu S and Kurzrock R. Development of curcumin as an epigenetic agent. *Cancer* 2010; 116: 4670–4676.
- Tsai KW, Wu CW, Hu LY, et al. Epigenetic regulation of miR-34b and miR-129 expression in gastric cancer. *Int J Cancer* 2011; 129: 2600–2610.
- Esquela-Kerscher A and Slack FJ. Oncomirs—microRNAs with a role in cancer. *Nat Rev Cancer* 2006; 6: 259–269.
- Griffiths-Jones S, Saini HK, van Dongen S, et al. miRBase: tools for microRNA genomics. *Nucleic Acids Res* 2008; 36: D154–D158.
- Shah MS, Davidson LA and Chapkin RS. Mechanistic insights into the role of microRNAs in cancer: influence of nutrient crosstalk. *Front Genet* 2012; 3: 305.
- West SG, Krick AL, Klein LC, et al. Effects of diets high in walnuts and flax oil on hemodynamic responses to stress and vascular endothelial function. *J Am Coll Nutr* 2010; 29: 595–603.
- Whelan J and McEntee MF. Dietary (n-6) PUFA and intestinal tumorigenesis. J Nutr 2004; 134: 3421S–3426S.
- Davidson LA, Wang N, Shah MS, et al. n-3 Polyunsaturated fatty acids modulate carcinogen-directed non-coding micro-RNA signatures in rat colon. *Carcinogenesis* 2009; 30: 2077–2084.
- 110. Hu S, Dong TS, Dalal SR, et al. The microbe-derived short chain fatty acid butyrate targets miRNA-dependent p21 gene expression in human colon cancer. *PLoS One* 2011; 6: e16221.
- 111. Rossi A, D'Urso OF, Gatto G, et al. Non-coding RNAs change their expression profile after Retinoid induced differentiation of the promyelocytic cell line NB4. *BMC Res Notes* 2010; 3: 24.
- 112. Garzon R, Pichiorri F, Palumbo T, et al. MicroRNA gene expression during retinoic acid-induced differentiation of

human acute promyelocytic leukemia. *Oncogene* 2007; 26: 4148–4157.

- 113. Spencer JP, Abd El, Mohsen MM, Minihane AM, et al. Biomarkers of the intake of dietary polyphenols: strengths, limitations and application in nutrition research. *Br J Nutr* 2008; 99: 12–22.
- 114. Lopez-Lazaro M. Anticancer and carcinogenic properties of curcumin: considerations for its clinical development as a cancer chemopreventive and chemotherapeutic agent. *Mol Nutr Food Res* 2008; 52(Suppl. 1): S103–S127.
- 115. Sun M, Estrov Z, Ji Y, et al. Curcumin (diferuloylmethane) alters the expression profiles of microRNAs in human pancreatic cancer cells. *Mol Cancer Ther* 2008; 7: 464–473.
- 116. Li Y, VandenBoom TG 2nd, Kong D, et al. Up-regulation of miR-200 and let-7 by natural agents leads to the reversal of epithelial-to-mesenchymal transition in gemcitabineresistant pancreatic cancer cells. *Cancer Res* 2009; 69: 6704–6712.
- 117. Bao B, Ali S, Banerjee S, et al. Curcumin analogue CDF inhibits pancreatic tumor growth by switching on suppressor microRNAs and attenuating EZH2 expression. *Cancer Res* 2012; 72: 335–345.
- Yang J, Cao Y, Sun J, et al. Curcumin reduces the expression of Bcl-2 by upregulating miR-15a and miR-16 in MCF-7 cells. *Med Oncol* 2010; 27: 1114–1118.
- Jin Y. 3,3'-Diindolylmethane inhibits breast cancer cell growth via miR-21-mediated Cdc25A degradation. *Mol Cell Biochem* 2011; 358: 345–354.
- 120. Hu FW, Tsai LL, Yu CH, et al. Impairment of tumorinitiating stem-like property and reversal of epithelialmesenchymal transdifferentiation in head and neck cancer by resveratrol treatment. *Mol Nutr Food Res* 2012; 56: 1247–1258.
- 121. Wen XY, Wu SY, Li ZQ, et al. Ellagitannin (BJA3121), an anti-proliferative natural polyphenol compound, can regulate the expression of MiRNAs in HepG2 cancer cells. *Phytother Res* 2009; 23: 778–784.
- 122. Surh YJ, Kundu JK, Na HK, et al. Redox-sensitive transcription factors as prime targets for chemoprevention with anti-inflammatory and antioxidative phytochemicals. *J Nutr* 2005; 135: 2993S–3001S.
- 123. Tsang JS, Ebert MS and van Oudenaarden A. Genomewide dissection of microRNA functions and cotargeting networks using gene set signatures. *Mol Cell* 2010; 38: 140–153.
- 124. Wang H, Bian S and Yang CS. Green tea polyphenol EGCG suppresses lung cancer cell growth through upregulating miR-210 expression caused by stabilizing HIF-1alpha. *Carcinogenesis* 2011; 32: 1881–1889.
- 125. Dixon RA and Pasinetti GM. Flavonoids and isoflavonoids: from plant biology to agriculture and neuroscience. *Plant Physiol* 2010; 154: 453–457.
- 126. Parker LP, Taylor DD, Kesterson J, et al. Modulation of microRNA associated with ovarian cancer cells by genistein. *Eur J Gynaecol Oncol* 2009; 30: 616–621