





Aspergillus Species from the Sabkha Marsh: Potential Antimicrobial and Anticancer Agents Revealed Through Molecular and Pharmacological Analysis

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Introduction: This study aimed to investigate the fungal growth and diversity in the Sabkha marsh. The anti-bacterial properties of the isolated fungi were assessed using an agar disk diffusion assay, and the crude extracts were tested for their anticancer activities. Liquid chromatography-mass spectrometry was employed to identify the active compounds of the fungal secondary metabolites. In-silico studies were conducted to predict the toxicity, pharmacokinetic properties, and safety profiles of the identified compounds.

Results: The analysis revealed that the isolated fungi belonged to the *Aspergillus* species, specifically *Aspergillus flavus* and *Aspergillus niger*. The crude extract of *A. flavus* exhibited significant anticancer activity against various cancer cell lines, while the antifungal activities against pathogenic bacteria varied between the two fungi. Liquid chromatography-mass spectrometry analysis identified several compounds in the fungal isolates. In *Aspergillus flavus*, the compounds included Aflavinine, Dihydro-24-hydroxyaflavinine, Phomaligin A, Hydroxysydonic acid, Gregatin B, Pulvinulin A, Chrysogine, Aspergilliacid, Aflatoxin B1, and Aflatoxin G1. In *Aspergillus niger*, the compounds identified were atromentin, fonsecin B, firalenone, rubrofusarin, aurasperone E, aurasperone D, aurasperone C, nigerone, and $\alpha\beta$ -dehydrocurvularin.

Conclusion: This study demonstrated promising fungal growth and diversity in the Sabkha marsh, with *Aspergillus* species being the most prevalent. The fungal crude extract showed anticancer activities against various cancer cell lines, while the antifungal activities against pathogenic bacteria varied between the two fungi. Future research should focus on investigating the antimicrobial activities of these fungi against multidrug-resistant bacteria and exploring the genetic changes in bacteria and cancer cells treated with these fungal extracts. Additionally, it is important to test the anticancer activity of the active compounds separately to determine which one is the active agent against cancer cells. This information can be used in drug development trials.

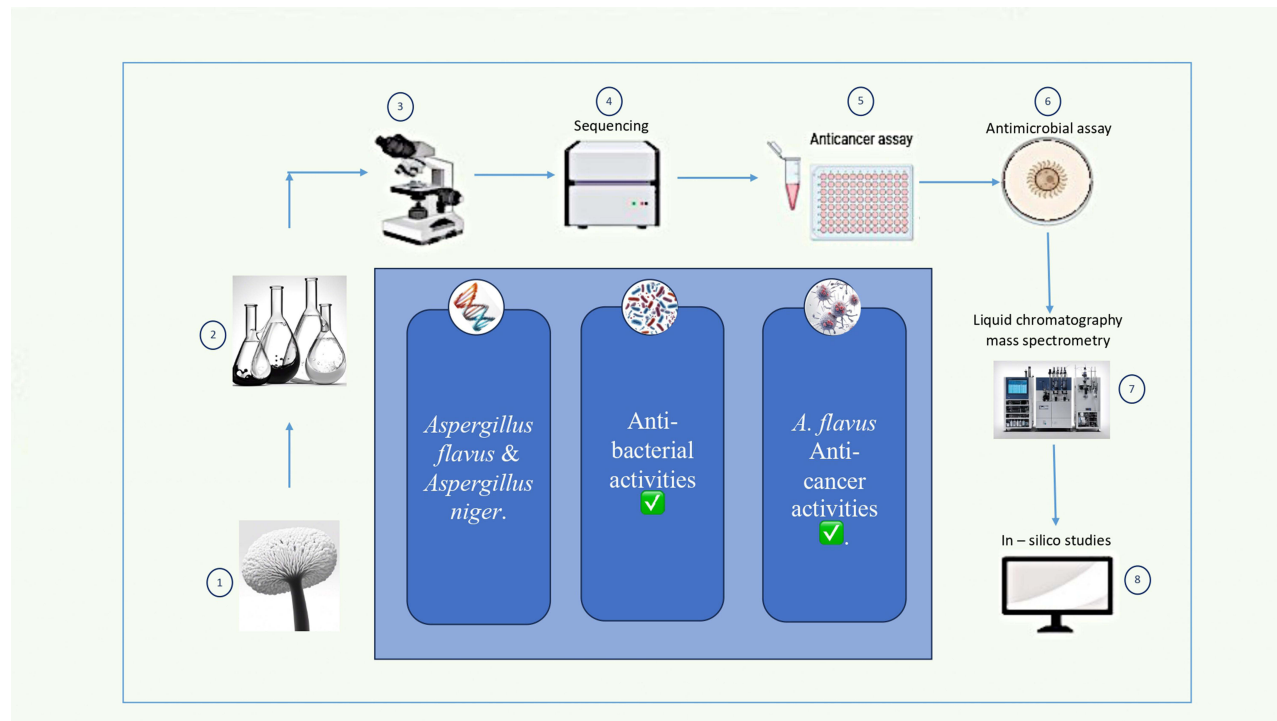
Keywords: saline environment, biological activity, *Aspergillus* genus, microbial isolation, antimicrobial screening

Introduction

The Sabkha desert sites in Saudi Arabia boast extreme environmental conditions, characterized by high salinity, neutral pH, and fluctuating moisture content, rendering them seemingly inhospitable for many life forms.¹ Despite these challenges, recent studies have unveiled the remarkable resilience of a diverse range of fungi in the Sabkha environment, showcasing their ability to thrive under extreme environmental pressures.¹

The presence of fungal species in the Sabkha desert not only underscores their capacity to adapt and flourish in challenging conditions but also sheds light on their pivotal role in maintaining ecological balance within extreme ecosystems.² The diverse microbial communities thriving in these environments, including fungal species such as

Graphical Abstract



Aspergillus, *Penicillium*, *Fusarium*, and *Chaetomium*, demonstrate the fungi's ability to endure harsh conditions and contribute to the ecological dynamics of the Sabkha desert.²

These findings underscore the necessity for further research into the adaptations and ecological significance of fungi in the Sabkha desert.² Fungi in the Sabkha Desert have evolved unique biochemical pathways and specialized structures that enable them to thrive in extreme temperatures and arid conditions.³ They utilize mechanisms such as heat-shock proteins and compatible solutes to safeguard their cellular structures from damage caused by heat or cold. Furthermore, some desert fungi can enter a dormant state during extreme conditions, ensuring their survival until more favorable conditions prevail.⁴

Thermophilic fungi have garnered attention for their potential applications in biotechnology and medicine. Certain compounds produced by these fungi have exhibited promising antimicrobial properties, positioning them as potential candidates for the development of novel antibiotics.⁵

The intriguing anti-cancer properties of these compounds have catalyzed the exploration of novel therapeutic strategies. Researchers are delving into the use of these fungal enzymes as potential targets for drug development, aiming to harness their anti-cancer effects in a more targeted and controlled manner. By comprehending the mechanisms of action of these enzymes, scientists aim to unlock new possibilities for cancer treatment.

For instance, *Aspergillus niger* has been extensively studied, with previous research testing biogenic silver nanoparticles produced from the extract of *Aspergillus niger* as antimicrobial, anti-cancerous, and anti-angiogenic agents.⁶ Similarly, Ethyl Acetate Extracted from *Aspergillus niger* has demonstrated both anticancer and anti-microbial activities in a previous study.⁷ As for *Aspergillus flavus*, a study utilized it to produce selenium nanoparticles and conducted further testing to evaluate their potential for biological applications, including antiviral, anticancer, antibacterial, antioxidant, and antibiofilm properties.⁸ Additionally, a pyranone derivative obtained from *Aspergillus flavus* was previously tested for its antimicrobial and antitumor activities.⁹

The relationship between cancer and microorganisms, particularly fungi, has been extensively explored, with a growing emphasis on the potential of thermophilic fungi in biotechnology and medicine.⁵ However, there is still a research gap in fully exploring the potential of the active compounds derived from these fungi, which possess both anti-cancer and antimicrobial properties.^{10,11} This study marks the commencement of future research endeavors aimed at evaluating and testing these active compounds separately for their pharmaceutical potential. The objective is to highlight the potential of locally sourced natural compounds, derived from our own land, for pharmaceutical applications.

Materials and Methods

Isolation and Morphological Characterization of Fungi

To investigate the microflora in the region of Makkah, specifically Rabigh city in Saudi Arabia, thirty samples of Sabkha soil were collected. Soil samples ($n = 5$) were obtained from depths of 5 to 20 cm, placed in clean plastic bags, and transferred to the laboratory for examination. The collected samples were then prepared for fungi isolation using dilution and blotter techniques. Subsequently, they were incubated at a temperature of 25 ± 2 °C for 7 days.¹²

Pure colonies were stored on potato dextrose agar (PDA) slants at 4 °C for future use. The isolated colonies were examined for species identification using traditional methods. These methods involved characterizing their texture on specific media, including Sabouraud dextrose agar and Czapek-Dox agar, all supplemented with chloramphenicol. Furthermore, surface, reverse colony, and microscopic observations were performed.¹²

Physicochemical Analysis of Soil Samples

The soil samples were examined using a digital camera to classify particle characteristics, including color, shape, and morphology. pH analysis was conducted using a pH meter (model HI98107), and the moisture content (MC) was determined according to.^{13,14} Additionally, the electrical conductivity (EC) of each soil sample was measured using an EC meter (Mettler Toledo-AG).

Microscopic Examination of Isolates

In this investigation, we examined the external characteristics of colonies and assessed their colors to identify the fungi. We used scanning electron microscopy (SEM) to observe fungal biofilms on the crystalline surface of the substrate. Sample preparation included fixing, drying, and gold coating the samples. Finally, we analyzed the samples using a SEM (JEOL 7500FA JEOL, Peabody, MA, USA) at 10 kV.

Characterization at the Molecular Level of the Isolated Samples

Preparation of Mycelium and DNA Extraction

To prepare the mycelium, a 2 μ L aliquot of potato dextrose broth (PDB) from Himedia, Mumbai, India was added to Potato Dextrose Agar (PDA) tubes and vortexed to disperse the spores. The spore-PDB mixtures were then added to flasks containing 100 mL of PDB and kept undisturbed at room temperature for two to three days. The mycelium was harvested by filtration, frozen at -80 °C for 30 minutes, lyophilized, and stored at -80 °C. Subsequently, the mycelium was ground in liquid nitrogen with a sterile mortar to obtain mycelium powder.

For DNA extraction, 20 mg of mycelium powder was used with a DNeasy Plant Mini Kit. The DNA quantity and quality were assessed by electrophoresis on a 0.8% agarose gel and visualized with ethidium bromide under UV transillumination.¹⁵

Amplification of ITS Region and PCR

The internal transcribed spacer (ITS) region of ribosomal DNA was amplified by PCR using the ITS1-F (5'-CTTGGTCATTTAGAGGAAGTAA-3') and ITS4-R (5'-TCCTCCGCTTATTGATATGC-3') primers. PCR amplification was carried out in a final volume of 50 μ L. The amplified product was checked by electrophoresis on a 1% agarose gel and visualized with ethidium bromide under UV transillumination, following the manufacturer's instructions.¹⁶

Sequencing and Analysis

The PCR product was purified using an Exo SAPIT kit, followed by Sanger sequencing at Beijing Genomic Institute (BGI), Hong Kong, China. The sequence was compared to the NCBI database using BLAST software. The obtained sequences were then aligned in Ugene using the T-Coffee algorithm.¹⁷ A phylogenetic tree was created using iTOL, an interactive tree tool, which offers various functionalities such as data upload, sharing projects, help resources, subscription information, and search capabilities.¹⁸

Antimicrobial Assay

The antibacterial properties of the fungal isolates were tested using an agar disk diffusion assay. Each fungal isolate was cultured on Sabouraud dextrose agar with 1% chloramphenicol for seven days at 28 °C. Afterward, 15 mm diameter disks were cut from the agar and placed on media plates inoculated with bacteria (blood agar, nutrient agar, and Sabouraud dextrose agar). The plates were then incubated at 37 °C for seven days. The antimicrobial activity was assessed by visualizing and measuring the inhibition zones (in mm).

Statistical Analysis

The data is presented as the mean plus or minus the standard deviation (SD) of three replicates.

Examining the Anticancer Activity

Preparing the Fungal Crude Extract

The contents of three fully grown fungal plates were transferred into a 500-mL beaker using a spatula under a biohazard cabinet, and 300 mL of brine solution was added to the beaker. The mixture was homogenized for 3–5 minutes at a rate of 7000–9000 rpm using the Ultra-Turrax[®] homogenizer. The homogenized mixture was then transferred into a specially designed 500-mL separating funnel see (Figure S1) in the [Supplementary files](#) and 300 mL of ethyl acetate was added. The mixture was stirred for 5 minutes at around 1100 rpm using an overhead stirrer, and the stirring was then stopped to allow the two layers to separate. The lower aqueous layer was transferred into a 500-mL conical flask, and the organic layer was drawn into another 500-mL conical flask with anhydrous sodium sulphate added to absorb the water. The aqueous phase was transferred back into the funnel, and the entire process was repeated twice. The organic phases were combined and filtered on paper into a 500-mL round bottom flask, then concentrated to a few mL on a rotary evaporator before being transferred into a tared vial. The crude extract was dried using a vacuum concentrator, and the vial was weighed and stored at 4°C until further use.

Cell Culture

In this study, six different cell lines were utilized. The hepatoma cell line HepG2, colorectal cancer cell lines HCT8 and HCT116, and the human drug-resistant metastatic breast cancer cell line MDA-MB-231 were procured from American Type Culture Collection (ATCC), United States. The epithelial-like breast carcinoma cell lines KAIMRC1¹⁹ and triple-negative KAIMRC2²⁰ were obtained and characterized at King Abdullah International Medical Research Center (KAIMRC) in Riyadh, Saudi Arabia. All these cell lines were cultivated in advanced Dulbecco's Modified Eagle Medium (DMEM) supplemented with 10% fetal bovine serum, 1% L-glutamine, and 1% antibiotics (Pen/Strep). The cells were cultured for a minimum of 24 hours before any experimentation, in 96-well plates at a density of 10,000 cells per well.

Cell Proliferation Assay (MTT)

In this study, the extracts from *Aspergillus flavus* isolate, and *Aspergillus niger* were investigated for their potential anticancer effects on the mentioned cell lines. The cells were exposed to varying concentrations of each extract, ranging from 0 to 1000 µg/mL. Doxorubicin, a chemotherapy drug, and mitoxantrone, an antineoplastic agent, were used as positive controls for assessing cell toxicity, while cells treated with Dimethylsulfoxide (DMSO) only were used as a negative control. After 48 hours, the cells were treated with 5 mg/mL MTT solution. Following a 3-hours incubation at 37 °C, the media was removed, and 100 µL of DMSO solvent was added to each well. After shaking the plate on an

orbital shaker for 45 minutes, the absorbance was measured at OD 590 nm. The half-maximal inhibitory concentration (IC₅₀) values for each substance were determined using the dose–response curve in GraphPad Prism 8. Each experiment was repeated three times for accuracy.

Utilizing Liquid Chromatography–Mass Spectrometry (QTOF-LCMS) for Secondary Metabolite Identification *Aspergillus Flavus*

The ethanolic extract of *Aspergillus flavus* was analyzed using total ion current spectra (TIC) raw data and the Mass Hunter qualitative and quantitative analysis software. The mass screening of the spectrum (Figure 1) yielded the following findings. Chemical characteristics were extracted from the LC–MS data using the Molecular Features Extraction (MFE) algorithm and the recursive analysis workflow. These features were screened based on detected nodes at various retention times per minute, with a minimum intensity of 6000 counts, and aligned with previously detected compounds, taking into account adducts ([M+H]⁺ and [M-H]⁻).

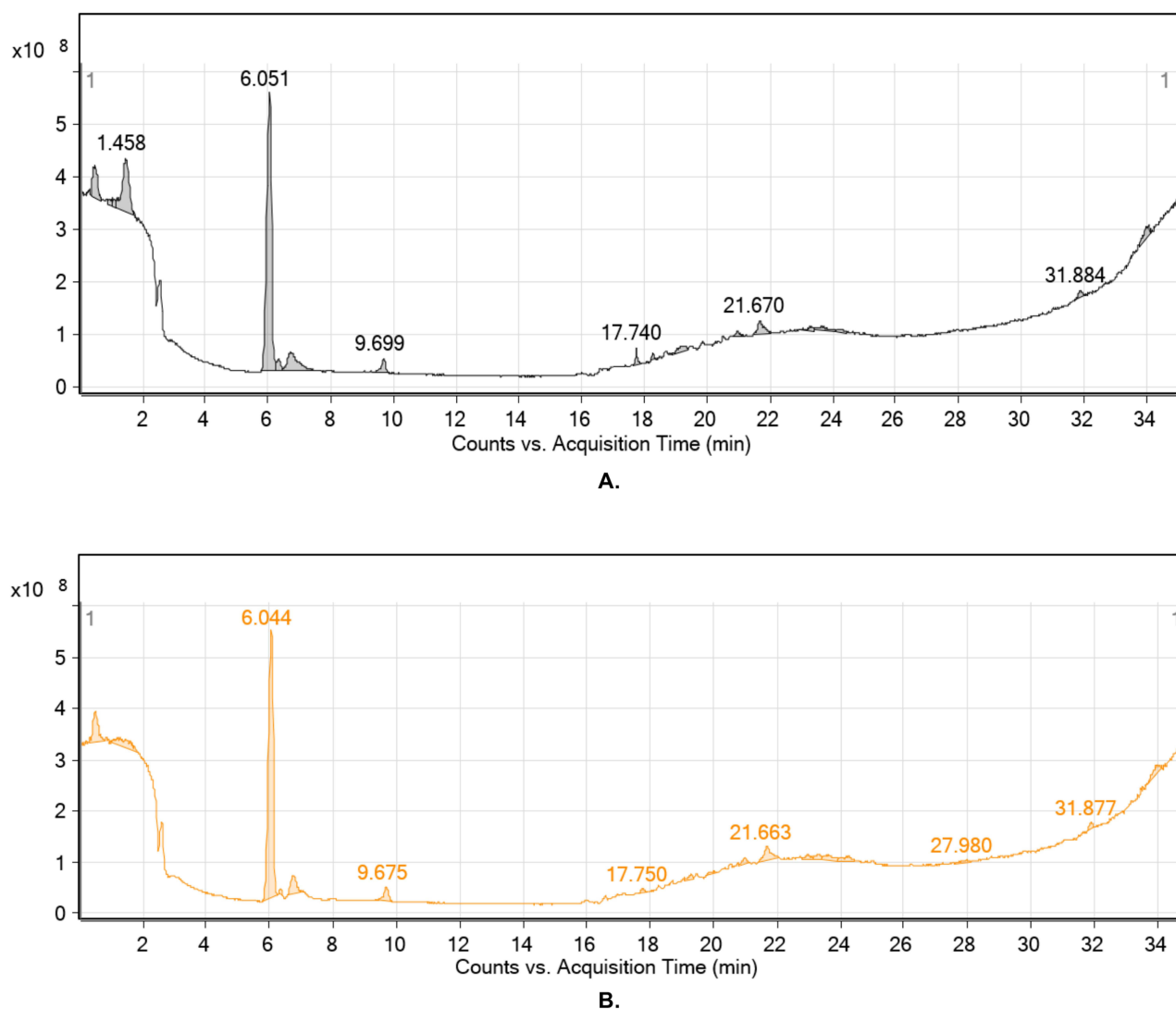


Figure 1 Elector spray ionization total ion chromatogram positive mode. (A) Electro spray ionization total ion chromatogram (Positive Mode) of *Aspergillus flavus* extract) and negative mode. (B) Electro spray ionization total ion chromatogram (Negative Mode) of *Aspergillus flavus* extract) of *Aspergillus flavus* extract.

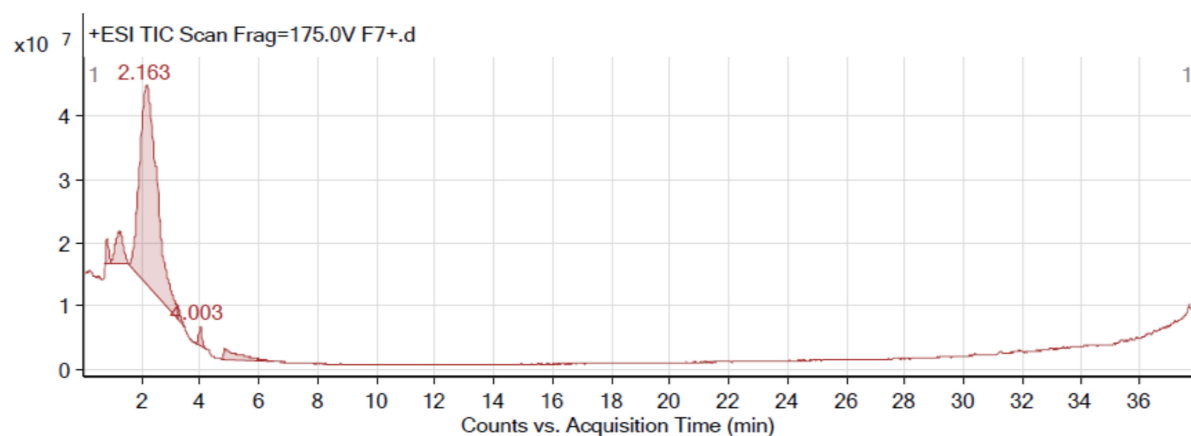
Aspergillus Niger

The ethanolic extract of *Aspergillus niger* was analyzed using the Mass Hunter qualitative and quantitative analysis software, based on the total ion current spectra (TIC) raw data refer to (Figure 2). Following the mass screening of the spectrum (Figure 2), the following observations were made. Chemical characteristics were identified from the LC–MS data using the Molecular Features Extraction (MFE) algorithm and the recursive analysis workflow. These features were obtained by evaluating the detected nodes at different retention times per minute, with a minimum intensity of 6000 counts, and were matched with previously identified compounds, taking into consideration adducts ($[M+H]^+$ and $[M-H]^-$).

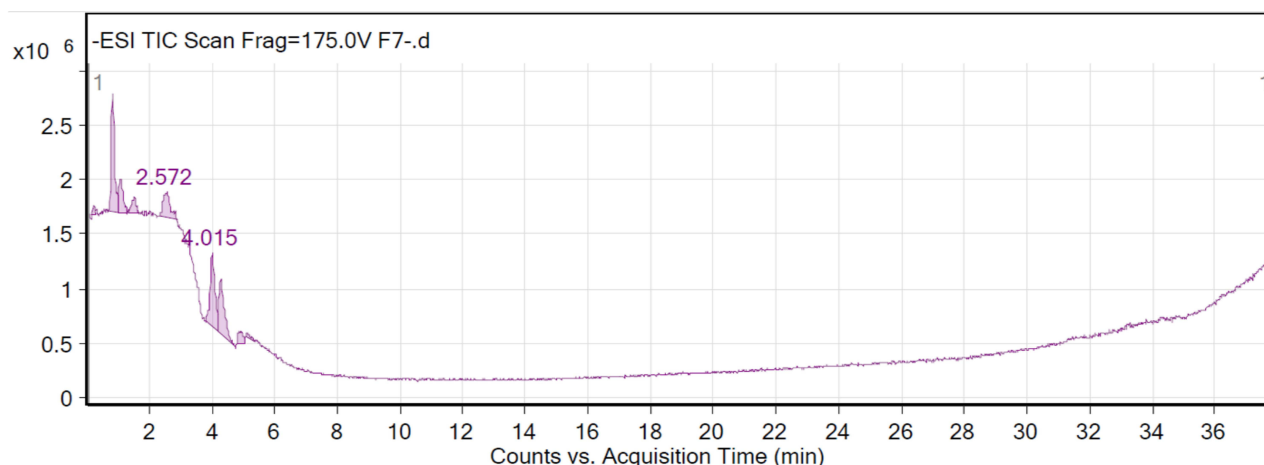
Computational Predictions and Investigations

Computational Predictions for Secondary Metabolites

The secondary metabolites of the fungal samples were identified using a library of 28 chemical structures. These structures were then used in computational predictions to assess the biological anticancer activity, pharmacokinetic



A.



B.

Figure 2 Elector spray ionization total ion chromatogram positive mode. (A) Electro spray ionization total ion chromatogram (Positive Mode) of *Aspergillus Niger* extract) and negative mode. (B) Electro spray ionization total ion chromatogram (Negative Mode) of *Aspergillus Niger* extract) of *Aspergillus Niger* extract).

properties, and safety profiles of each molecule. The in silico predictions were performed using Simplified Molecular Input Line Entry System (SMILES) representations, which are summarized in [Supplementary Table S1](#). The chemical structures of all the metabolites are shown in (Figures 3 and 4) for reference.

Aspergillus flavus

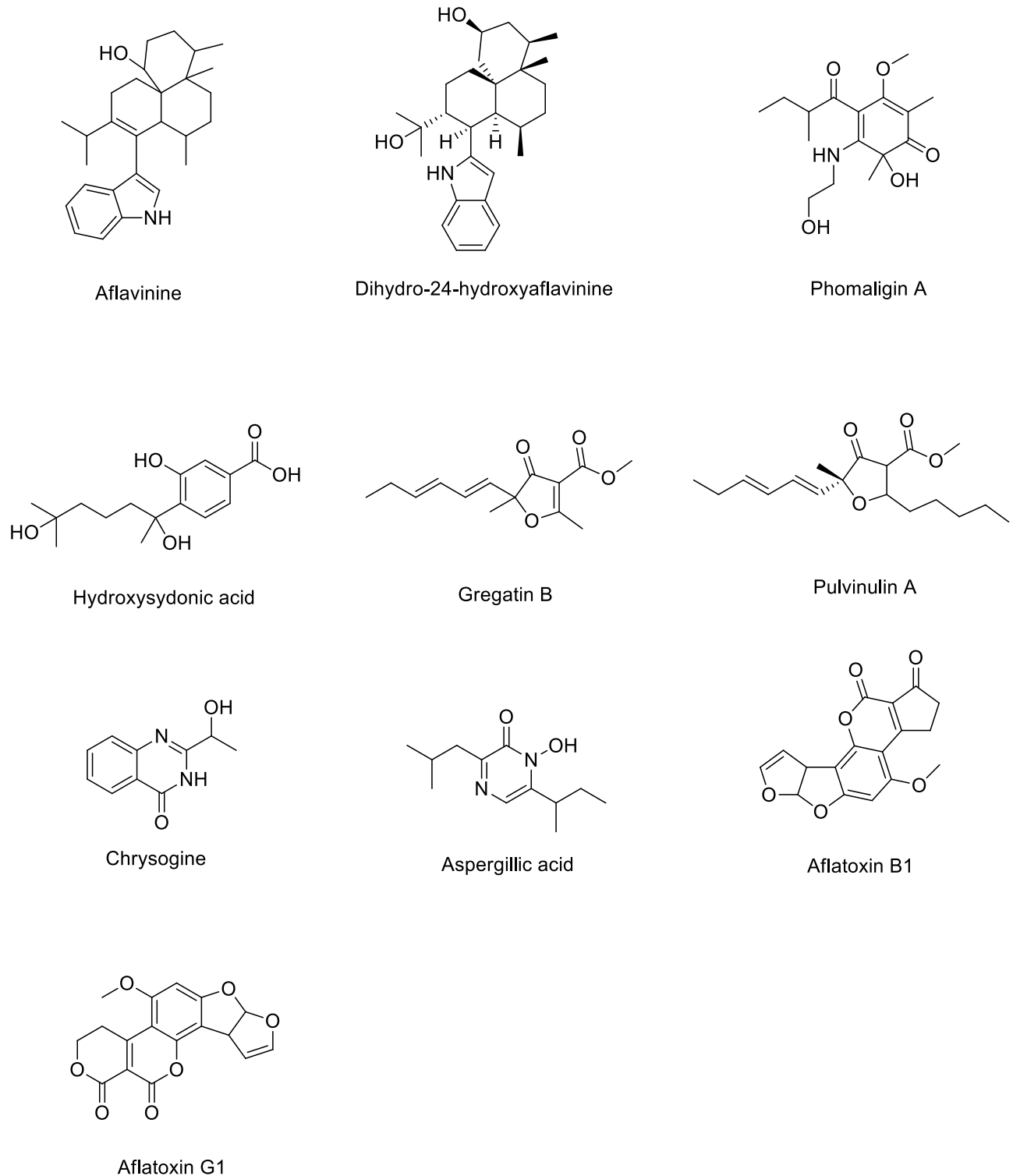


Figure 3 The chemical Structures of Secondary Metabolites Identified in *Aspergillus flavus*.

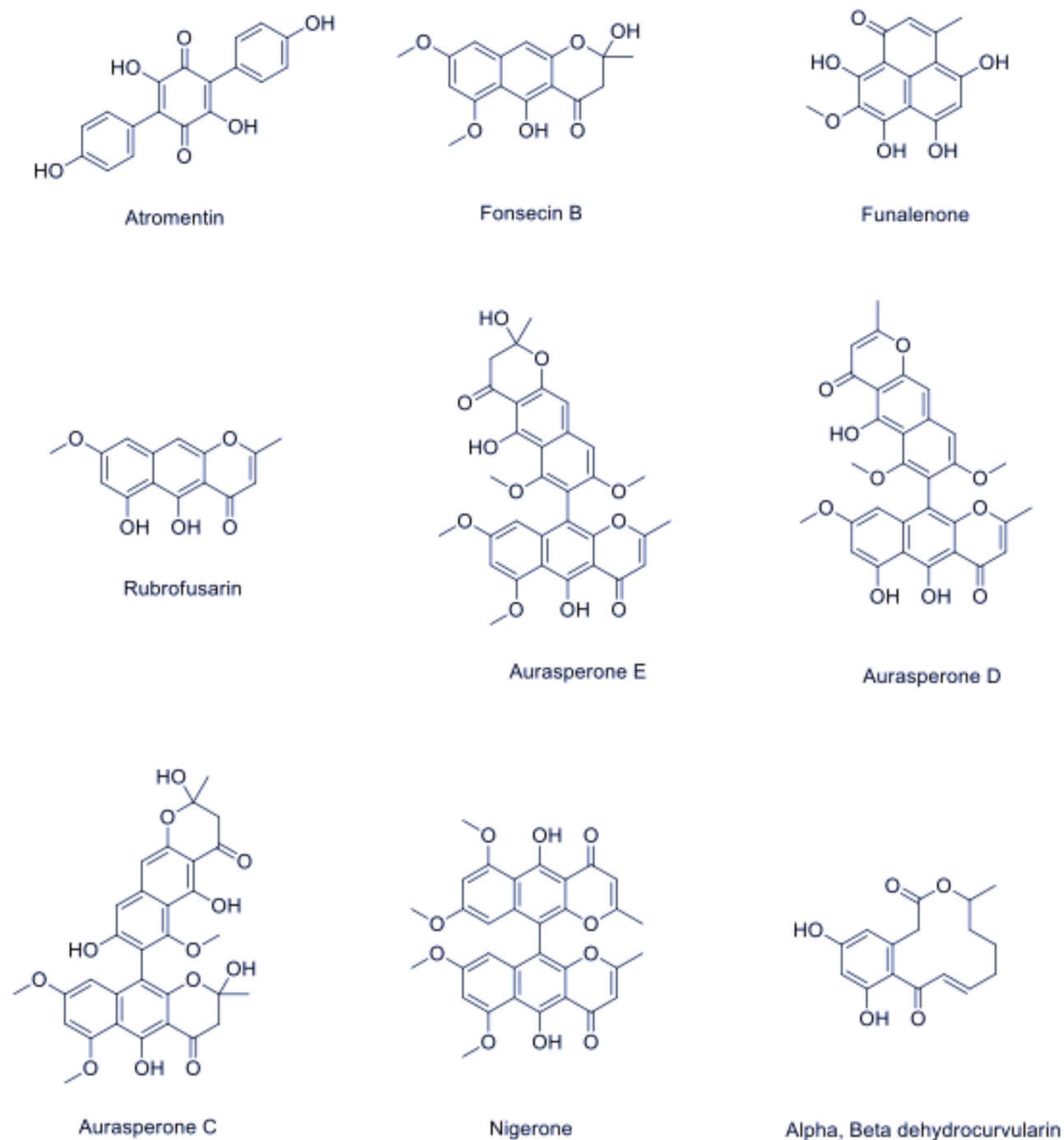
Aspergillus niger

Figure 4 The chemical Structures of Secondary Metabolites Identified in *Aspergillus Niger*.

Utilization of PASS Online Web Server for Anticancer Activity Predictions

To predict the anticancer potential of the identified metabolites, we inputted the 2D chemical structures into the PASS Online web server (Prediction of Activity Spectra for Substances). This web server can predict over 4000 types of biological activity, including pharmacological effects, mechanisms of action, toxic and adverse effects, and more. The

results are presented as a probability score (Pa), where a score exceeding 0.7 indicates strong anti-cancer properties. A score between 0.7 and 0.5 suggests moderate anti-cancer activity if experimentally tested.²¹

Predicting Pharmacokinetic Parameters and Oral Bioavailability

To assess various pharmacokinetic parameters for the identified metabolites, we utilized the SwissADME online web tool. These parameters include lipophilicity, molecular weight, solubility, insolubility, insaturation, and flexibility. The goal was to determine the likelihood of the metabolite being orally active. The results were presented using a radar plot, where the properties falling within the colored zone indicate the acceptable range for orally active drugs.²²

Leveraging the PASS Online Web Server for Anticancer Activity Predictions

To predict the anticancer activity of the identified metabolites, we submitted their 2D chemical structures to the PASS Online web server. The results are presented as a probability score (Pa). A score exceeding 0.7 indicates robust anticancer properties, while a score between 0.7 and 0.5 suggests moderate anticancer activity if experimentally tested. The PASS Online web server predicts over 4000 types of biological activity, including pharmacological effects, mechanisms of action, toxic and adverse effects, and more. This provides valuable insights into the potential anticancer properties of the compounds.²³

Assessment of Blockage of hERG K⁺ Channels

The potential cardiotoxicity of the identified metabolites was evaluated using the Pred-hERG 5.0 web server. This web server predicts the blockage of hERG K⁺ channels and provides probability predictions for each molecule, distinguishing between blocker and non-blocker. The Pred-hERG 5.0 web server utilizes advanced machine learning models and an extensive database to enhance accuracy and reliability in predicting hERG blockage. This assessment contributes to evaluating the cardiotoxicity of the compounds (https://tox-new.charite.de/protox_II/).²⁴

Results

Soil Physicochemical Investigations

The soil samples collected from all locations showed a slightly alkaline pH. The electrical conductivity (EC) of the soil samples was highest at 82400 $\mu\text{S}/\text{CM}$. The soil texture was clay. Upon closer examination of the soil granules, variations in particle size and shape were observed among the different sites studied (see Table 1 and Figure 5).

Macroscopic Results of Isolates

Conidial morphology-based characterization is a fundamental approach in fungal taxonomy.²⁵ Scanning Electron Microscopy (SEM) offers a precise method for examining conidial ornamentation, allowing for a more distinct differentiation between various types of ornamentation. Given that both isolates belonged to the *Aspergillus* genus, the resemblance of their conidia was clearly evident under SEM (Figure 6).

Molecular Identification of Fungal Strains

The strain was identified as *Aspergillus flavus* with a 100% match in both identity and coverage through the BLAST analysis. This strain has been assigned the accession number OR262346.1 and has been designated as WIS. The phylogenetic tree revealed the presence of three distinct clusters (Figure 7). Within one of these clusters, the *Aspergillus flavus* strain WIS was grouped alongside forty-two other *A. flavus* strains. Furthermore, it is noteworthy that the closest eight clusters also included strains of *A. flavus*.

Table 1 Soil Characteristics and Microbial Abundance

Location	PH	EC $\mu\text{S}/\text{CM}$	Sand %	Silt %	Clay %	Texture	Fungal Isolates (%)	Bacterial Isolates (%)
Makkah region, Rabigh city	8.11	82,400	1.5	1.6	8.3	Clay soil	9.29%	9.09%

Abbreviation: EC, electrical conductivity.



Figure 5 Images of the study area of the Makkah region, Rabigh city (Sabkha).

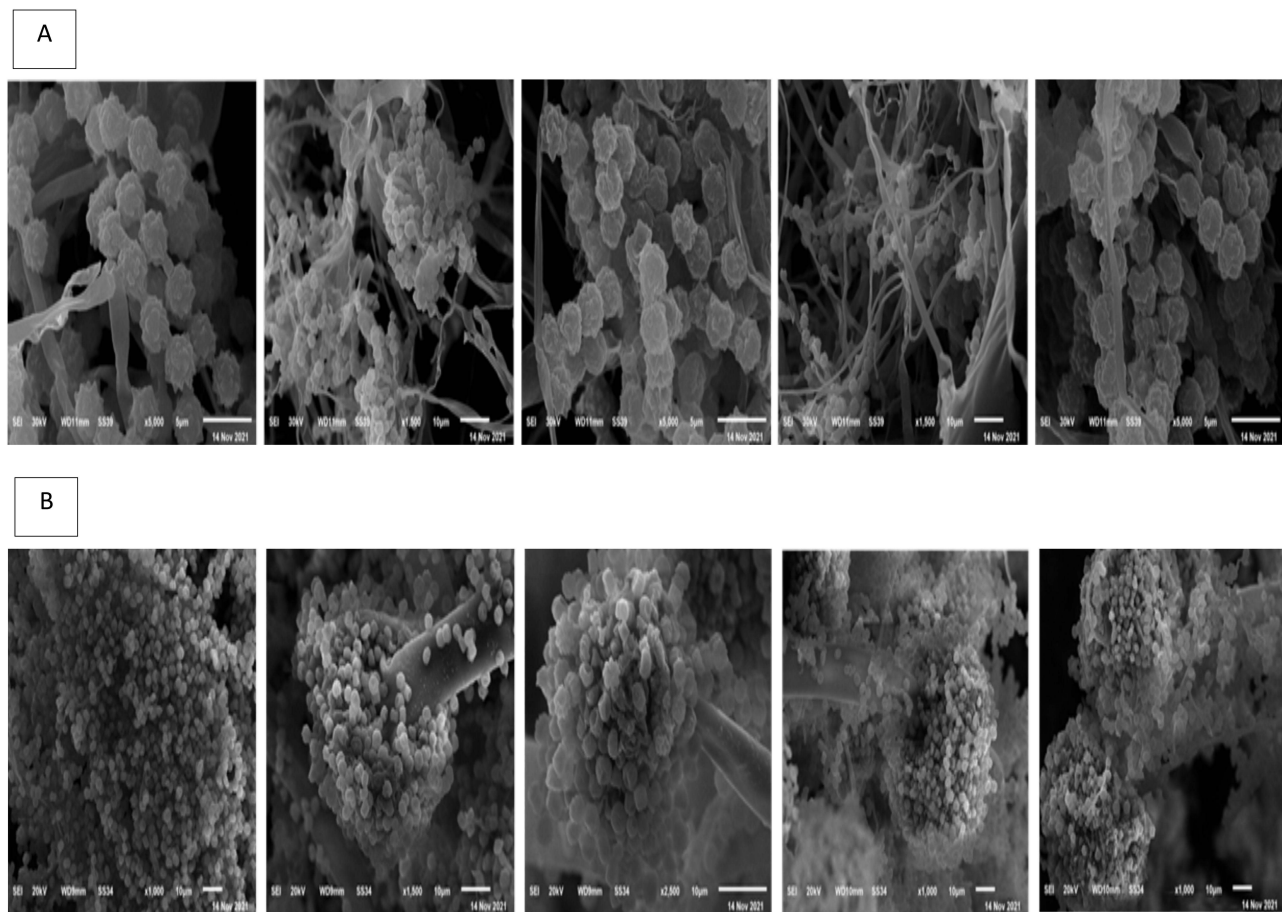


Figure 6 SEM images showing characteristic features of *Aspergillus* conidia: line (A) *Aspergillus flavus*, line (B) *Aspergillus Niger*.

The final WIS isolate was confirmed as *Aspergillus niger* with a 100% match in both identity and coverage through BLAST analysis. This specific strain was labeled WIS and allocated the accession number OR262345.1. In the phylogenetic tree, three distinct clusters were evident, as shown in (Figure 8). Within one of these clusters, the *Aspergillus niger* strain WIS was grouped alongside thirty-six other *A. niger* strains. Notably, the closest cluster also encompassed several *A. niger* strains.

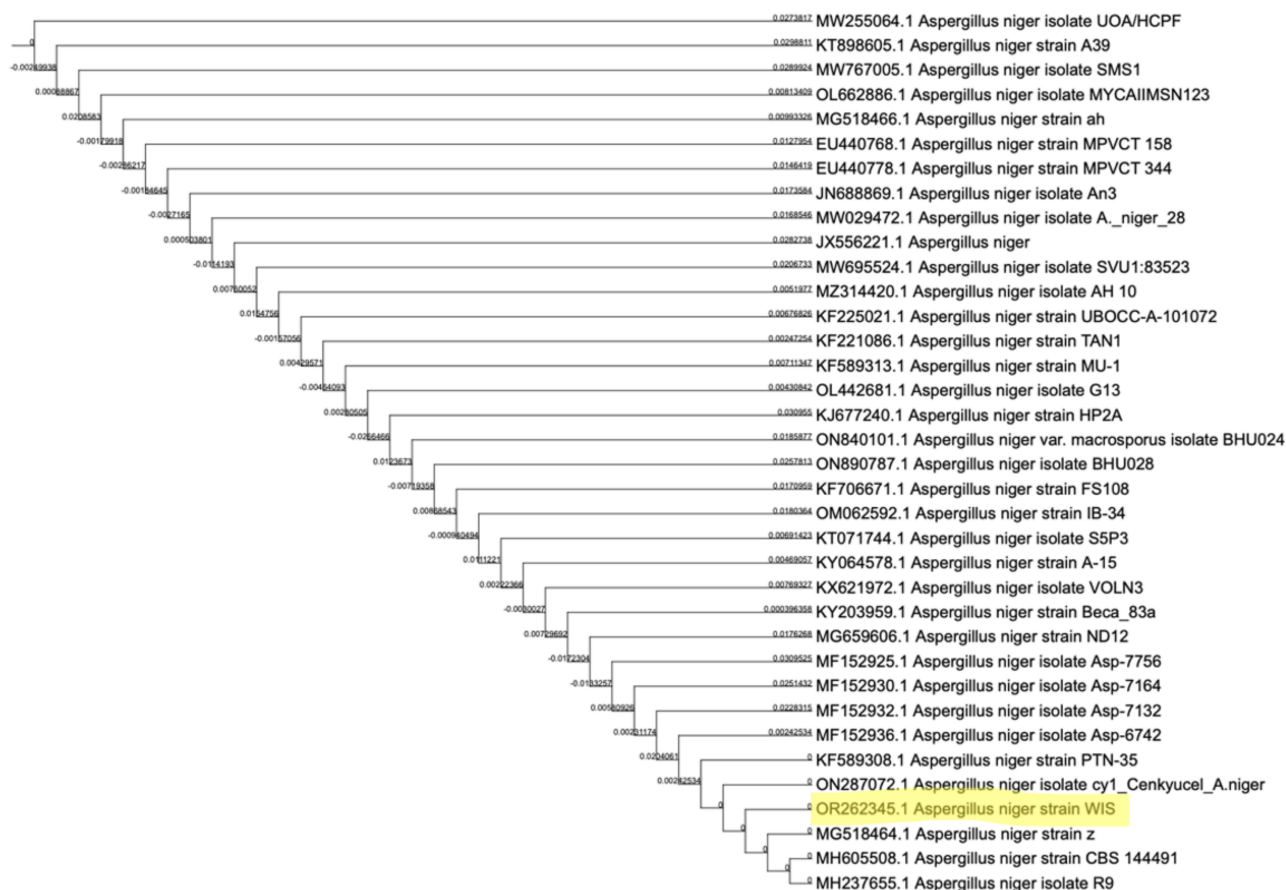


Figure 8 Phylogenetic tree showing the position of *Aspergillus Niger* strain WIS (acc. no. OR262345.1) and the closest related strains based on (ITS) region sequences.

growth inhibition, mitoxantrone and doxorubicin were employed as positive controls. We utilized six different cell lines, including breast cancer, hepatic cancer, and colorectal cancer, to diversify our findings. These cell lines have distinct genomic and proteomic profiles. Additionally, we used newly developed Saudi-specific KAIMRC1, ER+/PR+/HER2-, and KAIMRC2 triple-negative cell lines to target the local population.

The extracts from *Aspergillus flavus* isolate demonstrated significant anticancer effects on all cell lines, with IC50 values below 50 µg/mL. The IC50 results are summarized in (Table 3), and visual representations are provided in (Figure 11).

Table 2 Antibacterial Activity Using Fungi *Aspergillus Flavus* and *Aspergillus niger* Against Pathogenic Bacteria Evaluated by the Inhibition Zone Diameter in Mm

Growth Condition	Fungal Isolations	Gram-negative <i>E. coli</i>	Gram-positive <i>S. aureus</i>
Sabouraud Dextrose Agar	<i>Aspergillus flavus</i>	NE	NE
	<i>Aspergillus niger</i>	NE	NE
Nutrient Agar	<i>Aspergillus flavus</i>	14.8 ± 0.1	NE
	<i>Aspergillus niger</i>	14.8 ± 0.2	10.6 ± 0.4
Blood Agar	<i>Aspergillus flavus</i>	13.6 ± 0.4	13.7 ± 0.3
	<i>Aspergillus niger</i>	14.7 ± 0.3	11 ± 0.2

Notes: Mean significant at $p < 0.05$. *No effect (NE).

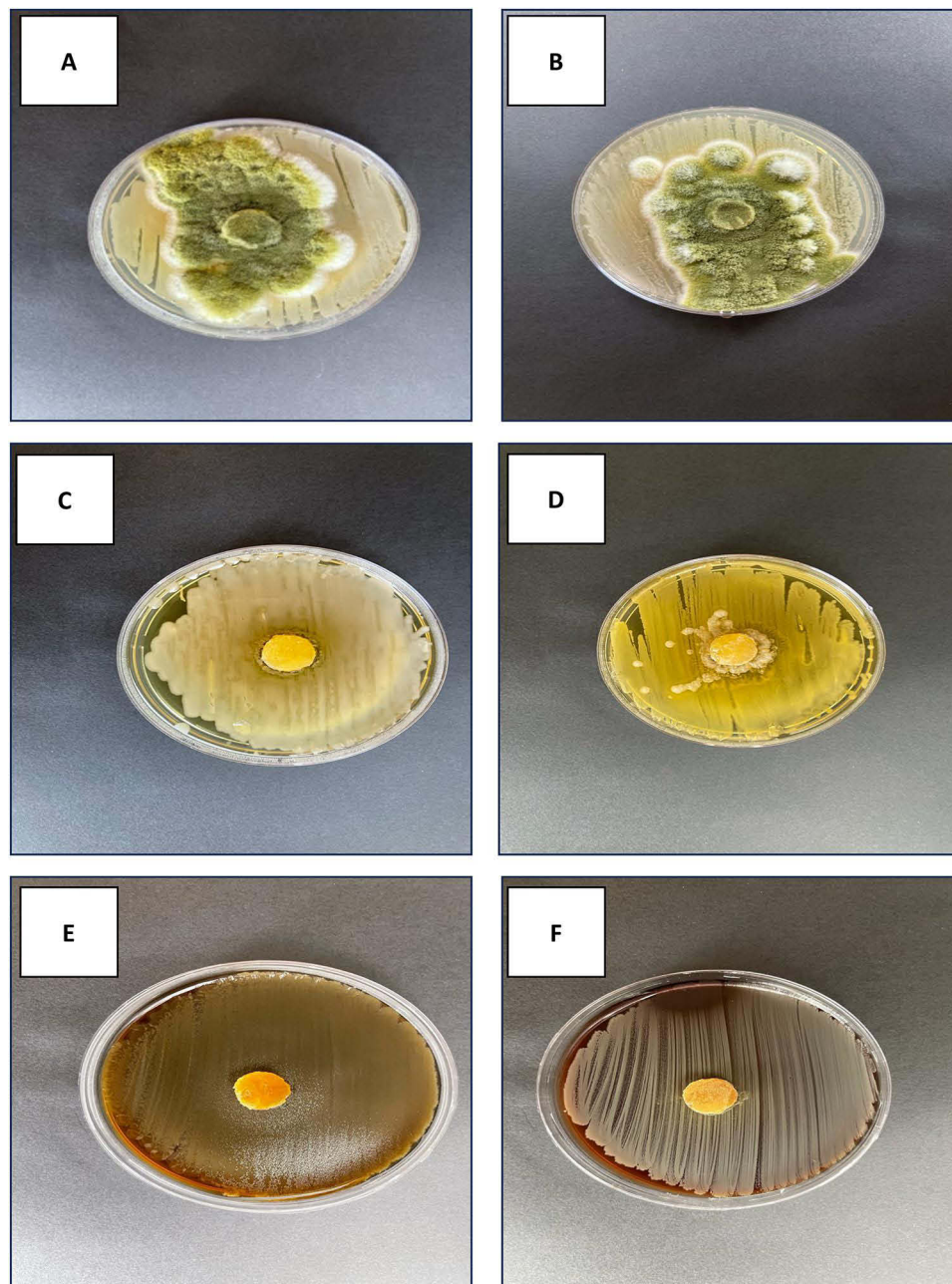


Figure 9 Antimicrobial activity against gram-positive bacteria (+ve) *Staphylococcus aureus* (*S. aureus*) and gram-negative bacteria (-ve) *Escherichia coli* (*E. coli*) of *Aspergillus flavus* on three different media, Sabouraud dextrose agar (SDA), nutrient agar (NA) and blood agar (BA). (A) (-ve on SDA); (B) (+ve on SDA); (C) (-ve on NA); (D) (+ve on SDA); (E) (-ve on BA) and (F) (+ve on BA).

Identification of Secondary Metabolites Using QTOF-LCMS

LCMS of *Aspergillus Flavus*

The tentatively identified compounds are Aflavinine,²⁶ Dihydro-24-hydroxyaflavinine,²⁶ Phomaligin A,²⁷ Hydroxysydonic acid,²⁸ Gregatin B,²⁹ Pulvinulin A,²⁹ Chrysogine,³⁰ Aspergillilic acid,³⁰ Aflatoxin B1,³¹ and Aflatoxin G1.³¹ The mean m/z implies the measured m/z (Figure 1) (Table 4).

LCMS *Aspergillus Niger*

The tentatively identified compounds are atromentin,³² fonsecin B,³³ firalenone,³⁴ rubrofusarin,³³ aurasperone E,³⁵ aurasperone D,³⁵ aurasperone C,³⁵ nigerone,³⁶ and $\alpha\beta$ -dehydrocurvularin.³⁷ The mean m/z implies the measured m/z (Figure 2) (Table 5).

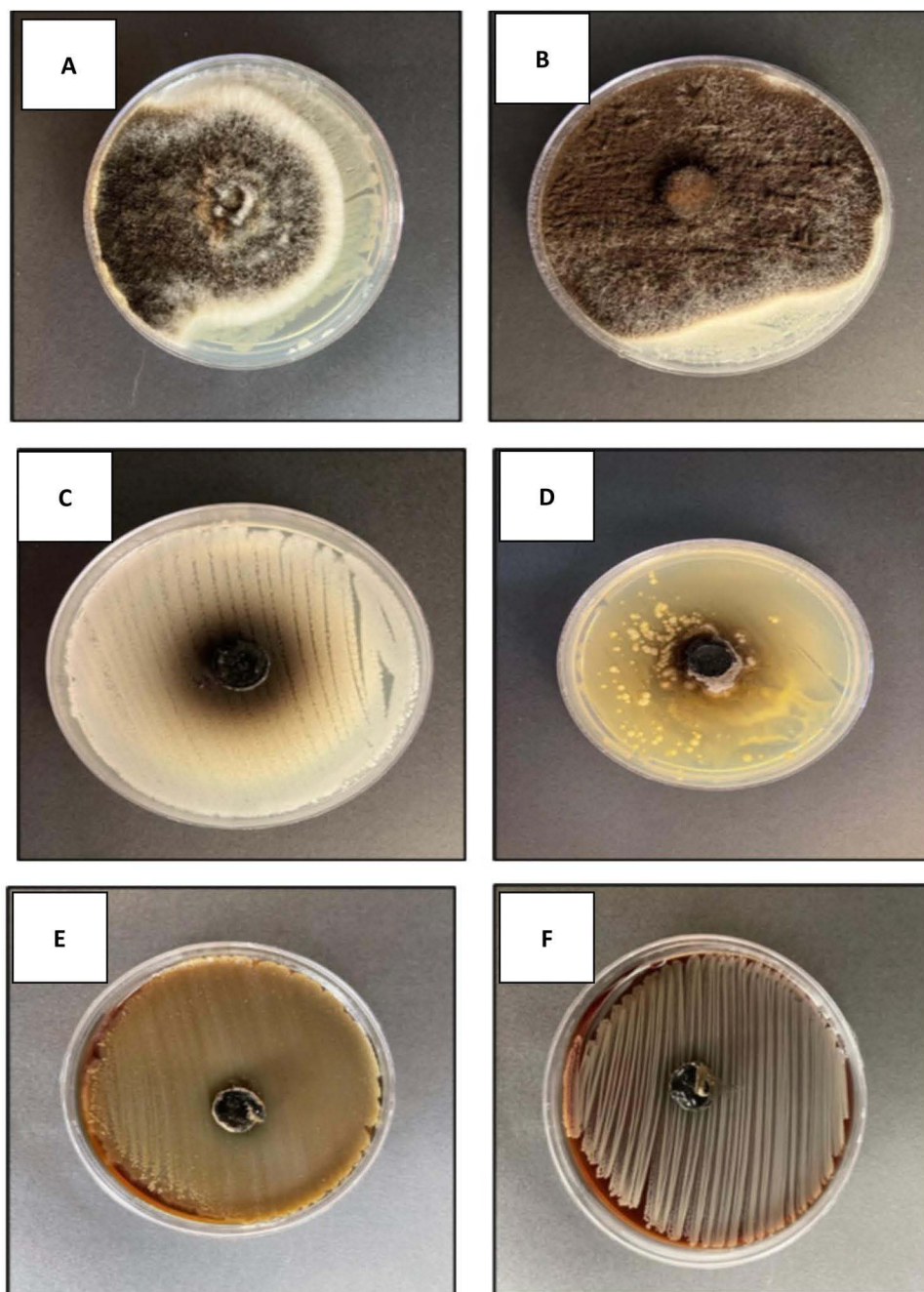


Figure 10 Antimicrobial activity against gram-positive bacteria (+ve) *Staphylococcus aureus* (*S. aureus*) and gram-negative bacteria (-ve) *Escherichia coli* (*E. coli*) of *Aspergillus Niger* on three different media, Sabouraud dextrose agar (SDA), nutrient agar (NA) and blood agar (BA). (A) (-ve on SDA); (B) (+ve on SDA); (C) (-ve on NA); (D) (+ve on SDA); (E) (-ve on BA) and (F) (+ve on BA).

Results for in-silico Predictions

Predicting Anti-Cancer Activities

The anti-cancer activity of the identified metabolites was evaluated using the PassOnline web server, a robust tool for predicting the potential anti-cancer properties of compounds. This analysis was conducted to determine the therapeutic advantages of the identified metabolites and their potential for the development of new anti-cancer drugs sourced from natural origins. The results, outlined in (Table 6), indicated that *Aspergillus Flavus*, along with two metabolites, Gregatin B and Pulvinulin A, exhibited promising anti-cancer activity, with P_a values of 0.902 and 0.880, respectively. Importantly, all the identified metabolites in

Table 3 The IC₅₀ (µg/ml) Values of Different Extracts Against Six Cancer Cell Lines Using the MTT Cell Viability Assay

#	Extracts	IC ₅₀ (µg/mL)											
		KAIMRC1		KAIMRC2		MDA MB 231		HEPG2		HCT8		HCT116	
		IC ₅₀	R ²	IC ₅₀	R ²	IC ₅₀	R ²	IC ₅₀	R ²	IC ₅₀	R ²	IC ₅₀	R ²
1	<i>Aspergillus flavus</i>	20.54	0.93	38.43	0.93	15.63	0.91	32.48	0.95	36.03	0.93	31.81	0.94
2	<i>Aspergillus niger</i>	615.4	0.6	324.9	0.86	98.46	0.91	91.06	0.95	193.5	0.93	176.8	0.91
3	Doxorubicin	0.1234	0.9	3.102	0.92	0.7421	0.9	57.33	0.73	0.6473	0.9	8.134	0.92
4	Mitoxantrone	0.09807	0.94	0.05587	0.83	0.1726	0.91	0.491	0.94	0.1389	0.91	0.443	0.9

Notes: White indicates no effect, Yellow indicates low effect, pink indicates good effect, and red indicates high effect.

Aspergillus niger demonstrated P_a values ranging from 0.709 to 0.854, suggesting that this fungus could be a potential reservoir for discovering highly effective anti-cancer agents.

Evaluation of Pharmacokinetic Parameters

In this section, we showcase the results derived from using the SwissADME online web server, a potent tool for evaluating the pharmacokinetic characteristics of the identified metabolites. Through this thorough analysis, we successfully evaluated diverse pharmacokinetic parameters and ascertained the potential oral activity of the metabolites.

In (Figure 12), the results of this analysis are presented, indicating that most of the metabolites demonstrate pharmacokinetic properties within the recommended range for lipophilicity, molecular weight, solubility, insolubility, insaturation, and flexibility. Notable metabolites showing favorable properties include Phomaligin A, Hydroxysydonic acid, Gregatin B, Pulvinulin A, Aspergillic acid, Aflatoxin B1, Aflatoxin G1, Fonsecain B, and αβ-dehydrocurvularin. However, some of the remaining metabolites exhibit deviations in one or more parameters. Further analysis and

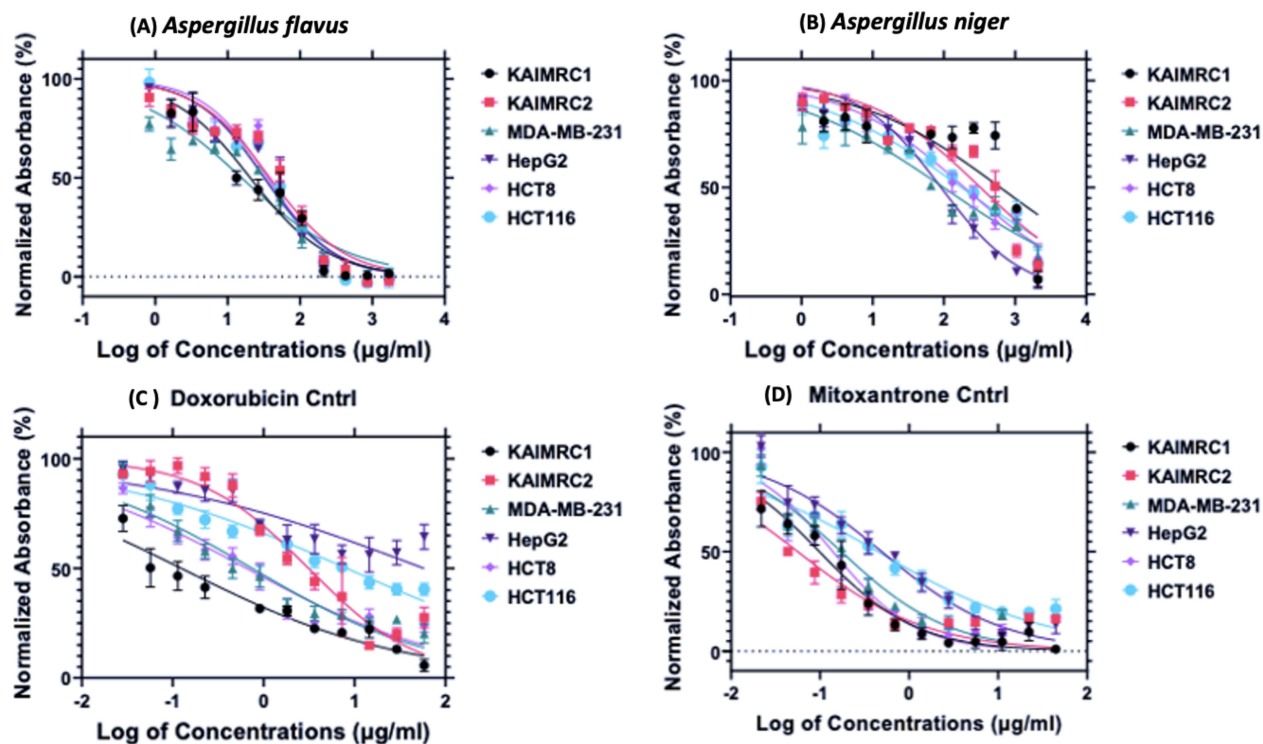


Figure 11 MTT assay on six cell lines. The extracts *Aspergillus flavus* (A) and *Aspergillus Niger* (B) along with Doxorubicin (C) and Mitoxantrone (D) were used. The X-axis shows the log of concentrations of the extracts in µg/mL, and the Y-axis shows normalized absorbance in %.

Table 4 Compounds (M1–M10) Tentatively Identified from *Aspergillus Flavus* Extract

Peak No.	Rt (min)	[M+H] ⁺	[M–H] [–]	Err PPM	Molecular Formula	Tentative Identification	Literature Review of the Compounds
Peak A (M1)	(18.139 — 18.474)	406.3107	405.7224	— 0.650	C ₂₈ H ₃₉ NO	Aflavinine	(21)
Peak B (M2)	(19.375 — 19.806)	423.6327	422.7226	— 0.85	C ₂₈ H ₄₁ NO ₂	Dihydro-24-hydroxyaflavinine	(21)
Peak C (M3)	(20.048 — 20.765)	312.1811	311.568	— 0.65	C ₁₆ H ₂₅ NO ₅	Phomaligin A	(22)
Peak D (M4)	(26.055 — 26.262)	283.1545	282.6022	— 0.85	C ₁₅ H ₂₂ O ₅	Hydroxysydonic acid	(23)
Peak E (M5)	(26.432 — 26.785)	251.1283	250.0682	— 0.75	C ₁₄ H ₁₈ O ₄	Gregatin B	(24)
Peak F (M6)	(26.938 — 27.176)	308.1880	307.6088	— 0.60	C ₁₈ H ₂₈ O ₄	Pulvinulin A	(24)
Peak G (M7)	(27.484 — 27.650)	191.0821	190.6734	— 0.55	C ₁₀ H ₁₀ N ₂ O ₂	Chrysogine	(25)
Peak H (M8)	(28.754 — 28.821)	225.1601	224.7422	— 0.45	C ₁₂ H ₂₀ N ₂ O ₂	Aspergillic acid	(25)
Peak I (M9)	(32.098 — 32.392)	329.0661	328.1667	— 0.30	C ₁₇ H ₁₂ O ₇	Aflatoxin B1	(26)
Peak J (M10)	(33.116 — 33.116)	313.0723	312.0719	— 0.70	C ₁₇ H ₁₂ O ₆	Aflatoxin G1	(26)

Table 5 Compounds (M1–M10) Tentatively Identified from *Aspergillus niger* Extract

Peak No.	Rt (min)	[M+H] ⁺	[M–H] [–]	Err PPM	Molecular Formula	Tentative Identification	Literature Review of the Compounds
Peak A (M1)	(17.347 — 17.878)	325.0697	323.0549	— 0.65	C ₁₈ H ₁₂ O ₆	Atromentin	(27)
Peak B (M2)	(18.347 — 18.804)	291.0861	289.0709	— 0.85	C ₁₅ H ₁₄ O ₆	Fonsecin B	(28)
Peak C (M3)	(19.408 — 19.605)	289.1811	287.568	— 0.70	C ₁₅ H ₁₂ O ₆	Funalenone	(29)
Peak D (M4)	(20.055 — 20.262)	273.2511	271.6022	— 0.40	C ₁₅ H ₁₂ O ₅	Rubrofusarin	(28)
Peak E (M5)	(21.432 — 21.785)	589.1283	587.0682	— 0.75	C ₃₂ H ₂₆ O ₁₁	Aurasperone E	(30)
Peak F (M6)	(21.938 — 21.176)	557.1880	555.6088	— 0.60	C ₃₁ H ₂₄ O ₁₀	Aurasperone D	(30)
Peak G (M7)	(23.484 — 23.650)	592.0821	592.6734	— 0.55	C ₃₁ H ₂₆ O ₁₂	Aurasperone C	(30)
Peak H (M8)	(25.754 — 25.821)	571.1601	569.7422	— 0.45	C ₃₂ H ₂₆ O ₁₀	Nigerone	(31)
Peak I (M9)	(27.098 — 27.392)	291.0661	289.1667	— 0.30	C ₁₆ H ₁₈ O ₅	αβ-dehydrocurvularin	(32)

Table 6 The Predicted Probabilities of Anti-Cancer Activity for the Identified Metabolites in *Aspergillus Flavus*, and *Aspergillus niger* Fungi Samples

Anti-cancer Activity for Metabolites	P _a	P _i
A) <i>Aspergillus Flavus</i>		
1. Aflavinine	0.378	0.113
2. Dihydro-24-hydroxyaflavinine	0.611	0.043
3. Phomaligin A	0.685	0.029
4. Hydroxysydonic acid	0.370	0.036
5. Gregatin B	0.902	0.005
6. Pulvinulin A	0.880	0.005
7. Chrysogine	0.215	0.031
8. Aspergillic acid	0.252	0.041
9. Aflatoxin B1	0.375	0.035
10. Aflatoxin G1	0.280	0.067

(Continued)

Table 6 (Continued).

Anti-cancer Activity for Metabolites	P _a	P _i
B) <i>Aspergillus niger</i>		
1. Atromentin	0.780	0.014
2. Fonsecin B	0.709	0.024
3. Funalenone	0.854	0.006
4. Rubrofusarin	0.772	0.015
5. Aurasperone E	0.720	0.023
6. Aurasperone D	0.794	0.013
7. Aurasperone C	0.762	0.017
8. Nigerone	0.787	0.013
9. $\alpha\beta$ -dehydrocurvularin	0.781	0.014

refinement may be necessary to optimize their pharmacokinetic properties for potential use as anti-cancer agents. These findings provide valuable insights into the development of new orally active anti-cancer drugs derived from natural compounds.

Assessment of Metabolite Toxicity for Anti-Cancer Applications

An extensive evaluation was carried out to assess the toxicity of the identified metabolites to ensure their safety and suitability for anti-cancer applications. The results of the comprehensive toxicity assessment are summarized in (Table 7). Chrysogine, among the analyzed metabolites, demonstrated a predicted hepatotoxicity of 50%. Additionally, three metabolites - Aflatoxin B1, Aflatoxin G1, and $\alpha\beta$ -dehydrocurvularin - were determined to possess a carcinogenicity of 60%. Immunotoxicity analysis revealed significant activity in 12 metabolites, with a probability range from 0.52 to 0.99. Moreover, only 5 metabolites exhibited mutagenic potential, while none were found to be cytotoxic. These findings offer valuable insights into the safety profile of the identified metabolites for further development as anti-cancer agents.

Prediction of Cardiac Toxicity

In order to conduct a thorough assessment of the potential toxicity of the identified metabolites, it was essential to investigate their ability to block the hERG channel, a known cause of cardiac toxicity. Alongside the evaluation of organ and endpoint toxicity, a cardiac toxicity assessment was performed using the pred-hERG web server. This analysis aimed to determine the metabolites' capacity to inhibit the hERG cardiac potassium channel and potentially induce adverse cardiovascular effects. Notably, the majority of the metabolites displayed no indications of cardiac toxicity (refer to Table 8), with the exception of Aflavinine and Aflatoxin G1, which exhibited a 62.81% and 52.47% probability of blocking the hERG channel, respectively.

Discussion

The two species identified belonged to the *Aspergillus* genus, known for their xerophilic nature, thriving in limited water availability.³⁸ Notably, the antibacterial activity of *Aspergillus flavus* and *Aspergillus niger* against *E. coli* underscores their potential as sources of new antibiotics. Furthermore, the varying degrees of antibacterial activity against pathogenic bacteria and the media-dependent nature of this activity open avenues for the development of novel antifungal agents.

In addition to their antibacterial properties, the extracts of *Aspergillus flavus* and *Aspergillus niger* have shown promising antifungal activity against various cancer cell lines. The potent efficacy of these extracts against aggressive and triple-negative breast cancer cell lines highlights their potential in the development of new anti-cancer drugs.^{20,39} Further research into the cytotoxic effects of these extracts at the protein level will provide valuable insights into the underlying mechanisms and pathways involved in their anti-cancer activity.⁴⁰

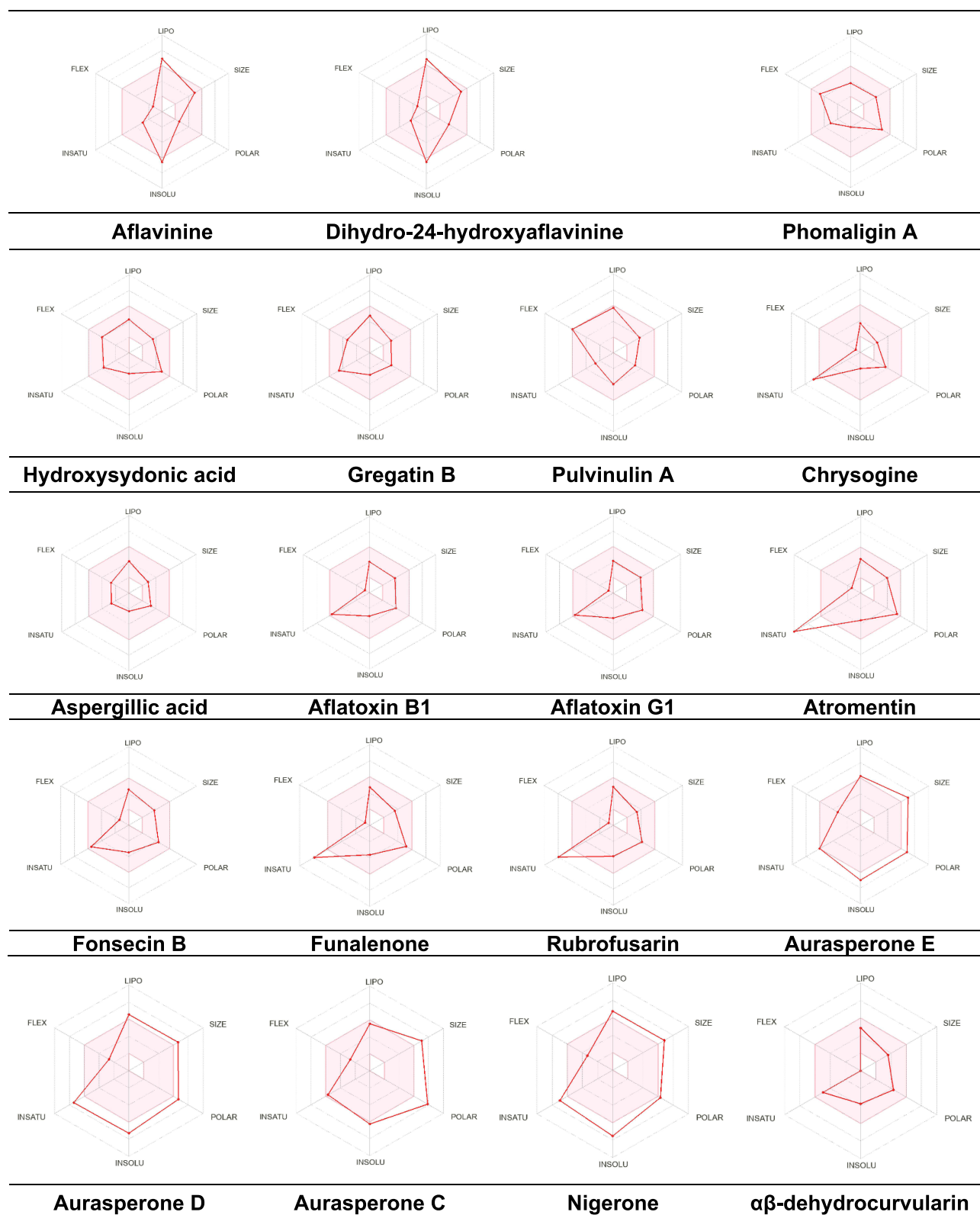


Figure 12 Pharmacokinetic Properties for the Identified Metabolites Using SwissADME Webserver.

Table 7 Comprehensive Toxicity Assessment of the Identified Metabolites Using Protox II Webserver

Metabolite Number	Classification				
	Organ Toxicity (%Probability)	Toxicity Endpoint (% Probability)			
	Hepatotoxicity	Carcinogenicity	Immunotoxicity	Mutagenicity	Cytotoxicity
<i>Aspergillus flavus</i>					
Aflavinine	Inactive (0.72)	Inactive (0.63)	Active (0.92)	Inactive (0.73)	Inactive (0.83)
Dihydro-24-hydroxyaflavinine	Inactive (0.68)	Inactive (0.72)	Active (0.93)	Inactive (0.70)	Inactive (0.83)
Phomaligin A	Inactive (0.76)	Inactive (0.66)	Active (0.88)	Inactive (0.75)	Inactive (0.69)
Hydroxysydonic acid	Inactive (0.58)	Inactive (0.69)	Inactive (0.99)	Inactive (0.77)	Inactive (0.83)
Gregatin B	Inactive (0.64)	Inactive (0.60)	Inactive (0.94)	Active (0.52)	Inactive (0.60)
Pulvinulin A	Inactive (0.75)	Inactive (0.62)	Inactive (0.95)	Inactive (0.79)	Inactive (0.66)
Chrysogine	Active (0.50)	Inactive (0.54)	Inactive (0.98)	Inactive (0.54)	Inactive (0.83)
Aspergillic acid	Inactive (0.52)	Inactive (0.55)	Inactive (0.99)	Inactive (0.59)	Inactive (0.76)
Aflatoxin BI	Inactive (0.67)	Active (0.68)	Active (0.99)	Active (0.95)	Inactive (0.83)
Aflatoxin GI	Inactive (0.71)	Active (0.60)	Active (0.99)	Active (0.81)	Inactive (0.86)
<i>Aspergillus niger</i>					
Atromentin	Inactive (0.57)	Inactive (0.55)	Inactive (0.99)	Inactive (0.85)	Inactive (0.63)
Fonsecin B	Inactive (0.71)	Inactive (0.55)	Active (0.93)	Inactive (0.50)	Inactive (0.81)
Funalenone	Inactive (0.56)	Inactive (0.64)	Active (0.94)	Active (0.50)	Inactive (0.78)
Rubrofusarin	Inactive (0.75)	Inactive (0.60)	Inactive (0.79)	Active (0.78)	Inactive (0.84)
Aurasperone E	Inactive (0.75)	Inactive (0.59)	Active (0.99)	Inactive (0.74)	Inactive (0.82)
Aurasperone D	Inactive (0.78)	Inactive (0.66)	Active (0.99)	Inactive (0.78)	Inactive (0.90)
Aurasperone C	Inactive (0.75)	Inactive (0.59)	Active (0.99)	Inactive (0.74)	Inactive (0.82)
Nigerone	Inactive (0.77)	Inactive (0.53)	Active (0.91)	Inactive (0.55)	Inactive (0.73)
$\alpha\beta$ -dehydrocurvularin	Inactive (0.79)	Active (0.66)	Active (0.88)	Inactive (0.88)	Inactive (0.70)

Table 8 Evaluation of Cardiac Toxicity in the Identified Metabolites Using the Pred-hERG Webserver

Metabolite Name	Activity on hERG Channel	Confiability %
<i>Aspergillus flavus</i>		
1. Aflavinine	blocker	62.81
2. Dihydro-24-hydroxyaflavinine	Non-blocker	81.85
3. Phomaligin A	Non-blocker	65.42
4. Hydroxysydonic acid	Non-blocker	99.34
5. Gregatin B	Non-blocker	80.63
6. Pulvinulin A	Non-blocker	80.07
7. Chrysogine	Non-blocker	65.6
8. Aspergillic acid	Non-blocker	99.48
9. Aflatoxin BI	Non-blocker	59.42
10. Aflatoxin GI	Blocker	52.47
<i>Aspergillus niger</i>		
1. Atromentin	Non-blocker	99.99
2. Fonsecin B	Non-blocker	67.87
3. Funalenone	Non-blocker	97.04

(Continued)

Table 8 (Continued).

Metabolite Name	Activity on hERG Channel	Confiability %
4. Rubrofusarin	Non-blocker	85.04
5. Aurasperone E	Non-blocker	67.04
6. Aurasperone D	Non-blocker	75.25
7. Aurasperone C	Non-blocker	70.12
8. Nigerone	Non-blocker	99.06
9. $\alpha\beta$ -dehydrocurvularin	Non-blocker	74.1

Moreover, the mass spectrometry analysis revealed the identification of several parent compounds, shedding light on the chemical composition of the samples studied. These compounds, such as fonsecin B, rubrofusarin, and aurasperone, hold promise for the discovery of new bioactive molecules with potential pharmaceutical applications in cancer and antibiotic drug development.^{33–37,41}

Furthermore, the comprehensive evaluation of the identified metabolites provides crucial insights into their toxicity profiles, emphasizing the need for further research to understand their toxicological implications. This study lays the foundation for the development of new orally active anticancer drugs derived from natural compounds and informs risk assessment and mitigation strategies for potential health risks associated with these metabolites.^{42,43}

The potential of these findings in the discovery of new anti-cancer and antibiotic drugs, coupled with the importance of continued research to address toxicity implications, positions this study at the forefront of future drug development and risk assessment in the field of cancer therapeutics and antimicrobial agents.

Conclusion

In conclusion, the study highlights the potential of metabolites derived from thermophilic fungi as a source of novel anti-bacterial and anti-cancer agents. The significant biological activity and favorable pharmacokinetic properties of the crude extract of these thermophilic fungi are promising. Certain metabolites indicate their potential for further development, so they should be studied separately. However, further optimization and refinement are necessary to overcome limitations and enhance their efficacy as antibiotics and anti-cancer drugs.

Overall, this study contributes to the ongoing efforts in the discovery and development of effective antibiotics and anti-cancer therapies from natural compounds.

Disclosure

The authors report no conflicts of interest in this work.

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